Role of interleukin-6 in cardiac inflammation and dysfunction after burn complicated by sepsis

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Zhang H, Wang HY, Bassel-Duby R, Maass DL, Johnston WE, Horton JW, Tao W. Role of interleukin-6 in cardiac inflammation and dysfunction after burn complicated by sepsis. Am J Physiol Heart Circ Physiol 292: H2408–H2416, 2007. First published January 12, 2007; doi:10.1152/ajpheart.01150.2006.—To examine the role of myocardial interleukin-6 (IL-6) in myocardial inflammation and dysfunction after burn complicated by sepsis, we performed 40% total body surface area contact burn followed by late (7 days) Streptococcus pneumoniae pneumonia sepsis in wild-type (WT) mice, IL-6 knockout (IL-6 KO) mice, and transgenic mice overexpressing IL-6 in the myocardium (TG). Twenty-four hours after sepsis was induced, isolated cardiomyocytes were harvested and cultured in vitro, and supernatant concentrations of IL-6 and tumor necrosis factor (TNF-α) were measured. Cardiomyocyte intracellular calcium ([Ca2+]i) and sodium ([Na+]i) concentrations were also determined. Separate mice in each group underwent in vivo global hemodynamic and cardiac function assessment by cannulation of the carotid artery and insertion of a left ventricular pressure volume conductance catheter. Hearts from these mice were collected for histopathological assessment of cardiac inflammation, fibrosis, and apoptosis. In the WT group, there was an increase in cardiomyocyte TNF-α, [Ca2+]i, and [Na+]i, after burn plus sepsis, along with cardiac contractile dysfunction, inflammation, and apoptosis. These changes were attenuated in the IL-6 KO group but accentuated in the TG group. We conclude myocardial IL-6 mediates cardiac inflammation and contractile dysfunction after burn plus sepsis.

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tumor necrosis factor-α; α-myosin heavy chain; cardiac contractile function; fibrosis; apoptosis

PROINFLAMMATORY CYTOKINES have been implicated in cardiac dysfunction after acute injuries, such as burn (13, 25) and sepsis (4). Our laboratory sequentially evaluated the effect of tumor necrosis factor-α (TNF-α), IL-1β, and IL-6 on cardiac contractile dysfunction and showed a complementary role of IL-6 to TNF-α and IL-1β: IL-6 mainly accentuated the myocardial depressive effects of TNF-α and IL-1β but lacked an independent effect (24). As part of the acute phase reaction, IL-6 is acutely upregulated after burn and sepsis. IL-6 is also involved in a wide range of physiological and disease processes, including aging (18). Whether and how such acute or chronic changes in IL-6 regulations mediate myocardial depression after acute injuries remain largely unknown. Currently, there is controversy regarding the exact role of IL-6 in the inflammatory response. Whereas many studies suggest an active role of this cytokine in acute inflammation (32), others indicate its role as a marker (31), or even as an anti-inflammatory mediator (44). The complex roles of IL-6 in mediating essential cell function and the inflammatory response after burn and sepsis prompted the present study examining the effects of IL-6 on cardiac inflammation and function using our clinically relevant model of burn complicated by sepsis (burn plus sepsis) (38, 43). On the basis of our previous findings (24), we hypothesize that IL-6 mediates myocardial dysfunction after burn plus sepsis. We used loss-of-function and gain-of-function approaches to evaluate the role of IL-6 in cardiac inflammation and dysfunction using mice deficient in IL-6 (IL-6 knockout mice, KO) and transgenic mice overexpressing IL-6 specifically in the heart [α-myosin heavy chain (MHC)-IL-6, TG]. TNF-β and IL-6 production were measured in the supernatant of isolated cardiomyocytes. Intracellular calcium and sodium overload was assessed in isolated cardiomyocytes as an indicator of myocardial injury (41, 43). In addition, we measured in vivo hemodynamic and cardiac contractile function, followed by histopathological assessment of cardiac inflammation, fibrosis, and apoptosis after burn plus sepsis.

MATERIALS AND METHODS

IL-6 knockout and transgenic mice. Male, 8- to 10-wk-old, wild-type (WT, C57BL/6J) and IL-6 knockout (IL-6 KO, C57BL/6J-Il6tm1Kopf) mice were obtained from the Jackson Laboratory (Bar Harbor, ME). The IL-6 KO mice have a targeted mutation in the IL-6 gene and do not produce IL-6 mRNA or protein. The colony has been maintained by the Jackson Laboratory for many years and used in numerous studies. The low-level IL-6 activity detected in our immunoassay in the cultured cardiomyocyte supernatant (see RESULTS) probably represents cross-reactivity with other structurally or functionally similar cytokines in the IL-6 family. To generate transgenic mice overexpressing myocardial IL-6 (TG mice), a regulatory region derived from the α-MHC promoter is used to drive IL-6 expression (37). A Flag-tagged form of mouse IL-6 cDNA (gift from Dr. Jian Zhang, University of Michigan) was placed downstream of the α-MHC promoter (11), followed by a 0.6-kb fragment of the human growth hormone intron and polyA signal sequences. The UT Southwestern Transgenic Core Facility generated three founder α-MHC-IL-6 TG mouse lines. Genotyping was performed by PCR using DNA extracted from a tail biopsy with the following primers (forward primer: 5'-GACGAAATTCGCCCTTCTCTTAGCTACAAG-3' and reverse primer: 5'-AATTCGGCCGCTAGGTTTTGCGGCGTTAGA-3'). Forward and reverse primers were designed to target corresponding IL-6 and human growth hormone DNA sequence, respectively (Fig. 1). Positive offspring was crossed with C57BL/6J mice for at least nine generations. Mice were handled in accordance with institutional and National
IL-6 Transgene → Control WT KO TG

Fig. 1. PCR analysis of DNA isolated from tails of wild-type (WT), IL-6 knockout (KO), and α-myosin heavy chain (MHC)-IL-6 transgenic (TG) mice shows insertion of IL-6 transgene. Control is α-MHC-IL-6 transgene DNA.

Institutes of Health guidelines. The study protocol was approved by the Institutional Animal Care and Use Committee. A total of 14–19 mice were used in each group, and at least 8 mice were assigned to in vivo hemodynamic and cardiac functional assessment. The rest of the mice were used for in vitro Western blot analysis of myocardial IL-6, cardiomyocyte supernatant cytokine secretion, and intracellular calcium ([Ca$^{2+}$]$_i$) and sodium concentrations ([Na$^+$]). The entire experimental procedure is illustrated in Fig. 2.

Burn plus sepsis injury model. We used our previously established model of 40% total body surface area (TBSA) contact burn followed by Streptococcus pneumoniae sepsis induced at 7 days after the initial burn (38, 43). We used this model because our institutional data showed most infectious complications occurred between 4 and 7 days after burn injury (9), and our experimental data showed a maximum myocardial depression after this injury model of burn complicated by sepsis (38, 43). The total body weight (TBW) of each mouse was obtained before the injury. The mice were anesthetized by placement in a plastic chamber supplied with 2.5% isoflurane in oxygen. Brussel probes preheated to 100°C were applied to the torso back of mice to induce the 40% TBBSA third-degree burn. The mice were given 4 ml·kg$^{-1}$·%TBBSA burn$^{-1}$ intraperitoneal lactated Ringer solution for resuscitation, and 0.05 mg·kg$^{-1}$ buprenorphine was administered every 30 min and centrifuged at 10,000 for 10 min at 4°C. Protein concentration was determined by the Bradford assay, using BSA for the standard curve (Bio-Rad Protein Assay Reagents, Hercules, CA). The protein samples were separated on a 4–20% SDS-polyacrylamide gel. The protein was transferred to a polyvinylidene difluoride membrane (Millipore, Bedford, MA). The membrane was blocked for 1 h in 5% nonfat milk and probed for 1 h with goat polyclonal mouse IL-6 antibody (1:1,000, cat. no. SC-1265, Santa Cruz Biotechnology, Santa Cruz, CA). After the blot was washed, donkey anti-goat IgG-horseradish peroxidase antibody was added (cat. no. SC-2033, Santa Cruz Biotechnology). After three washes (20 mM Tris, 135 mM NaCl, 0.1% Tween, pH 7.6), the bound antibody was visualized by enhanced chemiluminescence (ECL system, Amersham Biosciences, Piscataway, NJ).

Cardiomyocyte isolation, culture, and supernatant cytokine measurements. Twenty-four hours after S. pneumoniae instillation, mice were heparinized and decerebrated, and the heart was removed through a median sternotomy. The heart was immediately placed in ice-cold calcium-free Tyrode solution, followed by cannulation of the aorta and retrograde perfusion with calcium-free Tyrode solution equilibrated with 95% O$_2$–5% CO$_2$. The heart then underwent enzymatic digestion by perfusion with 80 ml of calcium-free Tyrode solution containing 20 mg collagenase A (Boehringer-Mannheim, Indianapolis, IN) and 2 mg of protease (polysaccharide XIV, Sigma, St. Louis, MO). The ventricular tissue was gently minced in Tyrode solution containing 100 μM calcium. The myocyte suspension was filtered, and the cells were allowed to settle. The supernatant was removed and cells resuspended in 50 ml Tyrode solution. The rinsing and settling steps were repeated as calcium concentration gradually increased from 100 μM to 1.88 mM. Cell viability was measured with Trypan blue dye exclusion. Cardiomyocytes were plated 50,000 cells·ml$^{-1}$·well$^{-1}$ and incubated at 37°C in 5% CO$_2$ for 18 h. Microtiter plates were removed from the incubator, and the supernatant was harvested to measure secreted TNF-$\alpha$ and IL-6 using ELISA kits (TNF-$\alpha$, Endogen, Woburn, MA; IL-6, Biosource, Camirillo, CA) (42).

Cardiomyocyte [Ca$^{2+}$]$_i$ and [Na$^+$]$_i$ measurements. Cardiomyocyte harvested from above were loaded with fura-2 AM for 45 min or sodium-binding benzofurazan isophthalate for 60 min at room temperature in the dark. They were suspended in Tyrode solution containing 1.88 mM calcium and washed to remove extracellular dye. Cardiomyocytes were placed in a perfusion chamber under an inverted microscope and stimulated (0.5 s) with platinum electrodes. Fluorescent images at 340- and 380-nm wavelengths in response to cell excitation were captured with [Ca$^{2+}$]$_i$ and [Na$^+$]$_i$ quantified by InCyt Im 2 Imaging System (Intracellular Imaging, Cincinnati, OH) (42).

Global hemodynamics and cardiac contractile function. Twenty-four hours after S. pneumoniae instillation, mice were anesthetized.
with 1.5% to 2% inhaled isoflurane in oxygen. Tracheostomy was performed to allow endotracheal intubation and positive pressure mechanical ventilation. The left carotid artery was cannulated for mean arterial blood pressure measurements and administration of lactated Ringer solution. A clamshell thoracotomy incision was performed to expose the heart. The pericardium was excised, and a 6-0 silk tie was placed around the inferior vena cava (IVC). The left ventricular (LV) pressure-volume catheter (SPR 839; Millar Instruments, Houston, TX) was inserted through the apex of the heart. Steady-state hemodynamic variables were measured, followed by contractility parameters with preload reduction achieved by transient IVC occlusion while suspending mechanical ventilation. Data were digitally converted and displayed using Chart software (version 4.12; ADInstruments, Castle Hill, Australia). Steady-state hemodynamic variables included heart rate (HR), stroke volume (SV), cardiac output (CO), and systemic vascular resistance (SVR). Variables obtained during IVC occlusion included LV end-systolic pressure-volume relationship (ESPRV), preload-recruitable stroke work (PRSW), and time-varying maximum elastance ($E_{max}$). All data were analyzed with pressure-volume analysis software (PVAN version 3.1; Millar Instruments) (38).

**Histopathology.** Immediately after cardiac function assessment, the heart was removed and placed in ice-cold lactated Ringer solution. The ventricles were isolated at the level of the atrioventricular annulus, rinsed, and weighed. The ventricle was cut transversely at the midpapillary level. Tissue blocks were fixed in 10% PBS-buffered neutral formaldehyde overnight, dehydrated with ethanol, and embedded with paraffin. They were sliced into 5-μm thickness sections according to standard protocols. Routine histomorphology was examined with hematoxin-eosin-stained slides. A semiquantitative grading system described by Nyska (26) was employed to evaluate the extent of cardiac inflammation: minimal (grade 1) changes involved 1–10%
of the section; mild (grade 2) involved 11–40%; moderate (grade 3) involved 41–80%; and severe (grade 4) involved 81–100%. Programmed cell death (apoptosis) of cardiomyocytes was assessed by terminal deoxynucleotidyl transferase (TdT)-mediated dUTP nick-end labeling (TUNEL) assay using the Promega Fluorescein Apoptosis detection kit (Promega, Madison, WI). Briefly, tissue sections were deparaffinized, preincubated with the equilibration buffer for 5 min, and incubated with TdT in the presence of digoxigenin-conjugated deoxyuridine triphosphate for 1 h at 37°C. The reaction was terminated by the stopping buffer. The sections were incubated with fluorescein-labeled anti-digoxigenin antibody (green, nuclear stain). An excess (~100 μl) of 1% propidium iodide (red) was added to each slide for nuclear counterstain. The slides were examined using 470-nm (FITC-green) and 535-nm (Cy-3, red) fluorescence microscope. All cell nuclei were stained red, but nuclei of apoptotic cells were stained green, and an overlap of green and red generated an orange image indicating the apoptotic cell. The distinction between apoptotic cardiomyocytes from noncardiomyocytes was made based on the morphology and location of the nuclei. Ten random high-power fields (~800) representing ~4,000 cells were counted to enumerate TUNEL-positive cells expressed as percentage of total cells (17). Normal thymus tissue was used as positive control, and negative controls were conducted in the same fashion as positive controls, except no TdT was added.

Cardiac fibrosis was assessed with slides stained with Masson’s Trichrome according to protocols described (1). Collagen was stained blue, and myocytes were stained red. A scoring system was used to define the degree of myocardial fibrosis as described by Shimizu et al. (34): grade 0, no fibrosis; grade I, focal fibrosis; grade II, moderate fibrosis; and grade III, diffuse fibrosis. Four different areas (left, anterior, posterior walls, and septum) were analyzed from each section, and the results were averaged.

Statistical analysis. All values are expressed as means ± SE. For each variable, comparisons were made among all groups using the WT group as control by one-way analysis of variance and Dunnett’s multiple comparisons test. In addition, Student’s t-test was performed for each variable within each group between sham and burn plus sepsis mice. A value of P < 0.05 was considered statistically significant.

RESULTS

There were no differences in cardiac structure and function between the commercial WT (C57BL/6J) mice and the WT littermates of the IL-6 TG mice. Therefore, only the WT group was used as the control group.

Myocardial IL-6 protein by Western blot analysis. Myocardial IL-6 protein content in WT, IL-6 KO, and TG groups is shown in Fig. 3. There was no detectable IL-6 protein in the KO group. Mice in the TG group had an abundance of IL-6 compared with the WT group (Fig. 3).

Cardiomyocyte supernatant TNF-α and IL-6 levels. In WT mice, both TNF-α and IL-6 increased after burn plus sepsis. In the IL-6 KO group, there was a low level of IL-6 activity at baseline and after burn plus sepsis, but the level was lower than the WT group. There was an increase in TNF-α levels in this group after burn plus sepsis injury. In TG mice, there was an increase in IL-6 and TNF-α levels after sham injury compared with the WT mice. After burn plus sepsis injury, both IL-6 and TNF-α further increased compared with those in the WT mice (Fig. 4).

Cardiomyocyte $[Ca^{2+}]_i$ and $[Na^+]_i$. Burn plus sepsis caused an increase in cardiomyocyte $[Ca^{2+}]_i$ and $[Na^+]_i$, in the WT mice. In IL-6 KO mice, baseline $[Ca^{2+}]_i$ and $[Na^+]_i$ were not different from those in the WT group. After burn plus sepsis, $[Ca^{2+}]_i$ and $[Na^+]_i$ both increased, but the levels were lower than those of the WT group. The TG group had higher levels of $[Ca^{2+}]_i$ and $[Na^+]_i$ at baseline, and these levels were higher than those of the WT group after burn plus sepsis (Fig. 5).

Global hemodynamics and cardiac contractile function. In the WT group, there was a decrease in HR and CO after burn plus sepsis; however, mean arterial pressure was maintained with an increase in SVR during steady-state hemodynamic measurements. All variables obtained during IVC occlusion showed decreased contractility after the injury. In the IL-6 KO group, the global hemodynamic and cardiac contractile changes were attenuated. In the TG group, hemodynamic changes were accentuated after burn plus sepsis, with a further reduction in CO and an increase in SVR, and all contractility variables showed myocardial depression compared with those in the WT group (Table 1). Representative pressure-volume tracings during IVC occlusions are shown in Fig. 6.

Heart weight, total body weight, and histopathological evaluation. The heart weight, TBW, and the heart-to-total body weight ratio (H/TBW) are summarized in Table 2. The IL-6 KO mice had a similar heart weight and TBW compared with the WT mice. In the TG mice, the TBW was similar to the WT mice, but the heart weight was higher. As a result, H/TBW was higher in the TG group than in the WT group. The LV wall thickness was similar in the WT and IL-6 KO groups but higher in the TG group.

Cardiac inflammation, apoptosis, and fibrosis are summarized in Table 3. In the WT group, there was no inflammation

Table 1. Changes in global hemodynamics (steady state) and cardiac contractility (IVC occlusion)

<table>
<thead>
<tr>
<th></th>
<th>WT</th>
<th>IL-6KO</th>
<th>TG</th>
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<tbody>
<tr>
<td></td>
<td>Sham</td>
<td>B+S</td>
<td>Sham</td>
</tr>
<tr>
<td>Global hemodynamics (steady state)</td>
<td></td>
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<td></td>
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<tr>
<td>HR, beats/min</td>
<td>562±12</td>
<td>494±9†</td>
<td>559±13</td>
</tr>
<tr>
<td>MAP, mmHg</td>
<td>86±3</td>
<td>85±2</td>
<td>88±5</td>
</tr>
<tr>
<td>CO, ml/min</td>
<td>9.1±0.4</td>
<td>5.0±0.4†</td>
<td>9.2±0.8</td>
</tr>
<tr>
<td>SVR, mmHg/ml^{-1}·min</td>
<td>9.4±0.4</td>
<td>17.5±1.7†</td>
<td>9.6±0.9</td>
</tr>
<tr>
<td>Cardiac contractility (IVC occlusion)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>ESPVR, μl/mmHg</td>
<td>19.1±9.0</td>
<td>5.1±1.1†</td>
<td>19.3±4.3</td>
</tr>
<tr>
<td>PRSW, μl/mmHg</td>
<td>106±6</td>
<td>39±2†</td>
<td>109±11</td>
</tr>
<tr>
<td>$E_{max}$, μl/mmHg</td>
<td>41.5±3.5</td>
<td>16.9±2.9†</td>
<td>43.4±2.3</td>
</tr>
</tbody>
</table>

Values are means ± SE. B+S, burn plus sepsis; IVC, inferior vena cava; WT, wild-type; IL-6 KO, IL-6 knockout; TG, transgenic; HR, heart rate; MAP, mean arterial pressure; CO, cardiac output; SVR, systemic vascular resistance; ESPVR, end-systolic pressure-volume relationship; PRSW, preload-recruitable stroke work; $E_{max}$, maximum elastance. *P < 0.05 vs. WT group; †P < 0.05 vs. sham.
associated with sham injury, but inflammation increased to 
grade 2 after burn plus sepsis injury. In the KO group, the 
inflammatory changes were attenuated after burn plus sepsis. 
In the TG group, there was basal inflammation associated 
with sham injury, and the inflammatory response increased to 
grade 3 after burn plus sepsis injury (Table 3, Fig. 7). The apoptosis 
was increased after burn plus sepsis injury in the WT group, 
but such changes were attenuated in the KO group. Apoptosis 
was most pronounced in the TG group (Fig. 8). There was no 
discernible fibrosis in both WT and KO mice, but there is 
moderate collagen (blue) deposition (grade 2) in the TG group 
(Table 3 and Fig. 9).

**DISCUSSION**

Myocardial depression after acute injuries such as burn or 
burn plus sepsis is largely considered to be mediated by TNF-α 
and IL-1β, which initiate a cascade of events, including in-
duced production of nitric oxide and free radicals (21), altered 
calcium and sodium handling (2, 43), and increased myocardial 
apoptosis (22). Although IL-6 is increased as part of the acute 
phase reaction, its role in mediating myocardial depression 
after acute injuries is not as clearly defined as the roles of 
TNF-α and IL-1β. We focused on the role of IL-6 in cardiac 
inflammation and function after burn plus sepsis as a clinically 
relevant model of injury where our laboratory has repeatedly 
shown that myocardial depression after this two-hit injury is 
exaggerated compared with burn alone or sepsis alone (33, 38, 
42, 43).

Results in our study support the hypothesis that IL-6 is an 
active mediator in the inflammatory response of the heart: mice 
deficient in IL-6 show attenuated inflammation, reduced apop-
tosis, and improved contractile function, whereas mice over-
expressing myocardial IL-6 showed more pronounced inflam-
mation, increased apoptosis, and depressed contractile func-
tion. These results are consistent with previous findings that 
IL-6 mediates the inflammatory response. For example, intrin-
sic production of IL-6 by cardiomyocytes was increased after 
burn plus sepsis (43) or hemorrhagic trauma (45). Treatment 
with anti-IL-6 antibodies reduced mortality after endotoxin injec-
tion alone (36) or after burn followed by endotoxin injec-
tion (27, 28). More recently, the IL-6 gene has been shown to 
be upregulated in patients suffering from meningococcal septic 
shock, and removal IL-6 from serum of these patients reversed 
the negative inotropic effects on rat cardiomyocytes in vitro 
(29). The fact that IL-6 increases in parallel with other proin-
flammatory cytokines, most notably TNF-α and IL-1β, makes 
it difficult to study the effect of an increase in IL-6 alone. It is 
generally believed that IL-6 participates in a wide range of 
immunological actions, such as activation of intracellular ad-
hesion molecule-1 (20), stimulation of antigen-presenting mac-
rophages and dendritic cells, recruitment of leukocytes, all 
promoting the inflammatory process (16). Whether and how 
IL-6 independently promotes myocardial inflammatory re-
sponses and cardiac dysfunction have not been well studied. In 
a model of isolated cardiomyotes, Yu et al. (46) showed 
incubation of cardiomyocytes with IL-6 increased inducible 
nitric oxide synthase activity and decreased cardiomyocyte 
shortening. Finkel et al. (8) had previously demonstrated the

![Graph](image_url)

**Table 3. Quantification of inflammation, apoptosis, and fibrosis**

<table>
<thead>
<tr>
<th></th>
<th>WT</th>
<th>KO</th>
<th>TG</th>
</tr>
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<tbody>
<tr>
<td>Grade of inflammation</td>
<td>Sham</td>
<td>B+S</td>
<td>Sham</td>
</tr>
<tr>
<td>Myocardial, %</td>
<td>0</td>
<td>0.17</td>
<td>&lt;0.1</td>
</tr>
<tr>
<td>Nonmyocardial, %</td>
<td>0</td>
<td>0.23</td>
<td>&lt;0.1</td>
</tr>
<tr>
<td>Grade of fibrosis</td>
<td>0</td>
<td>0</td>
<td>II</td>
</tr>
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B+S, burn plus sepsis.

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Table 2. Heart weight, total body weight, heart-to-total body weight ratio, and LV wall thickness

<table>
<thead>
<tr>
<th></th>
<th>Heart, g</th>
<th>TBW, g</th>
<th>Heart/TBW, %</th>
<th>LV Wall Thickness, mm</th>
</tr>
</thead>
<tbody>
<tr>
<td>WT</td>
<td>0.119±0.006</td>
<td>26.33±0.75</td>
<td>0.451±0.021</td>
<td>1.500±0.020</td>
</tr>
<tr>
<td>KO</td>
<td>0.110±0.007</td>
<td>24.69±0.82</td>
<td>0.446±0.015</td>
<td>1.583±0.224</td>
</tr>
<tr>
<td>TG</td>
<td>0.124±0.008*</td>
<td>26.27±1.05</td>
<td>0.472±0.012*</td>
<td>1.887±0.259*</td>
</tr>
</tbody>
</table>

Values are means ± SE. TBW, total body weight; LV, left ventricle. *P < 0.05 vs. WT group.
relationship between IL-6-induced nitric oxide synthase activity and altered myocardial tension in vitro. Using isolated heart preparations, our group has shown IL-6 possessed little direct myocardial depressive effect, but its synergism with TNF-α and IL-1β exaggerated their myocardial depressive effects (24). Our present study further demonstrates the role of IL-6 in exacerbating cardiac dysfunction in intact subjects.

The different levels of inflammation and apoptosis after burn plus sepsis among the three experimental groups in our study are well illustrated. These findings further support the theory that the heart is not only a mechanical pump but also an immune active organ (39). Locally produced proinflammatory cytokines, including TNF-α and IL-1β, probably mediate a large part of the myocardial dysfunction associated with injury and disease. In the present study, IL-6 likely mediated myocardial dysfunction either through its own action or by promoting the production of and synergizing with TNF-α and IL-1β. Indeed, myocardial IL-6 overexpression was associated with increased TNF-α production even after sham injury, and this increase was even more pronounced after burn plus sepsis.

The inflammatory response after burn plus sepsis was accompanied by intracellular calcium and sodium overload. Such an overload was attenuated in the KO mice but accentuated in the TG mice. Intracellular calcium and sodium overload has been shown in a variety of disease models, including cardiac hypertrophy and failure (30) and ischemia-reperfusion injury (7). It is considered a major indicator of myocardial cell injury. Our group has shown that intracellular calcium and sodium overload is a contributing factor to myocardial contractile dysfunction after burn or burn plus sepsis (2, 12, 14, 15). The mechanisms of increased [Ca^{2+}], and [Na^{+}], are not completely understood: burn and/or sepsis may alter sarcoplasmic calcium ATPase expression and function (2), causing an increase in [Ca^{2+}], with a compensatory increase in [Na^{+}], as a result of accelerated Ca^{2+}/Na^{+} exchange, or such injuries induce intracellular acidification, which results in increased Na^{+}/H^{+} exchange and, subsequently, Ca^{2+}/Na^{+} exchange (35).

In addition to promoting the inflammatory response, IL-6 also appears to be associated with myocardial and nonmyocardial cell apoptosis. It has been well established that apoptosis is a major event after myocardial injury such as ischemia-reperfusion (40), and it has been implicated in endotoxin-induced myocardial dysfunction (19). Our coworkers have demonstrated the importance of apoptosis in myocardial dysfunction after burn (5, 23) and endotoxemia (6). Similar to findings by Carlson et al. (6), the percentage of cells showing positive TUNEL stains was numerically too low to correlate with the degree of functional deterioration, which could be explained in part by the rapid clearance of apoptotic cells by macrophages. In addition, proinflammatory cytokines, especially TNF-α, not only participate in triggering apoptosis (6, 19) but also initiate a wide range of other inflammatory processes, including activation of inducible nitric oxide synthase (21). It was not clear whether the different levels of apoptosis in our study was the result of IL-6 itself or different activities of TNF-α and other inflammatory cytokines. The observed apoptosis may serve more as a marker of the inflammatory activity than as a critical component of contractile dysfunction. Nonetheless, it is reasonable to link the existence or overabundance of IL-6 to the more pronounced apoptotic activities after burn plus sepsis.

All three groups of mice showed similar global hemodynamic and cardiac contractile function after sham injury, although TG mice showed mild inflammation with increased TNF-α production and cardiomyocyte [Ca^{2+}], and [Na^{+}]. The reason for the normal function in sham TG mice despite the
mild inflammation is not clear. It is possible that the basal inflammatory state that occurred as a result of IL-6 overexpression produced numerous compensatory responses in the intact animals, masking mild or modest cardiac contractile dysfunction. In addition, IL-6 overproduction in the myocardium does not simulate a true pathological insult, such as burn injury, with regard to signaling derangements (3) that contribute to myocardial dysfunction. Whereas a rise in basal IL-6 production is seen in a variety of processes, including aging (10), major organ dysfunction predominantly occurs after major insults such as burn or burn plus sepsis (9).

In the present study, the KO mice were deficient in IL-6 in all tissues, whereas TG mice overexpressed IL-6 in the myocardium only. We chose total body IL-6 knockout because IL-6 is produced by many organs and tissues, including liver, gut, macrophages, leukocytes, and Kupffer cells. IL-6 produced in these tissues after burn plus sepsis will most likely circulate to the heart, making it impossible to investigate the effect of the loss of IL-6 on the myocardium. On the other hand, we had to use myocardium-specific IL-6 overexpressing mice because IL-6 exerts a wide range of physiological effects, including participation of total body immune response, regulation of body mass, and more importantly, stimulation of adrenocorticotropic hormones and interaction with estrogen (18). Selective cardiac overexpression of IL-6 enables us to focus on its role on myocardial structure and function without affecting other organ systems. We believe such selection of genetic groups of mice allowed us to better address the role of IL-6 in the myocardium after burn plus sepsis.

In summary, we showed myocardial IL-6 production was increased after burn plus sepsis injury, along with intracellular calcium and sodium overload, myocardial inflammation and

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Fig. 8. Representative images of heart tissue apoptosis. After burn plus sepsis (B+S), there was apoptotic activity in the WT, but not in the IL-6 KO group. In the TG group, there was apoptosis after sham, and apoptosis became more significant after B+S (TUNEL, original magnification ×800. All nuclei were stained red, and apoptotic nuclei were stained green. Overlapping the red and green images generated orange nuclei, indicating the type and location of cells undergoing apoptosis).

Fig. 9. Representative histograms of collagen deposition in the heart. Cardiac fibrosis was more significant in the TG mice (Masson’s Trichrome, original magnification ×400).
apoptosis, and depressed myocardial contractile function. These effects were partially attenuated by deficiency in IL-6 and accentuated by myocardial IL-6 overexpression. Myocardial overexpression of IL-6 was also associated with elevated basal inflammation as evidenced by more prominent inflammatory infiltration and apoptosis and increased cardiomyocyte basal inflammation. Overexpression of IL-6 was also associated with elevated and accentuated by myocardial IL-6 overexpression. Myocardial inflammation, apoptosis, and depressed myocardial contractile function.

There are limitations to the present study. Although we documented histopathological and functional changes associated with IL-6 after burn plus sepsis, there were no data that directly explain the molecular mechanisms of IL-6 in mediating myocardial dysfunction. In addition, we have noted changes in heart weight and LV wall thickness in the TG group, but we have not initiated the detailed analysis of the mechanism of such structural changes. Future studies aimed at the mechanisms of IL-6 will help us understand how IL-6 mediates the inflammatory response in the heart and explore interventions directed at preserving cardiac function after burn plus sepsis.

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