Heightened efficacy of nitric oxide-based therapies in type II diabetes mellitus and metabolic syndrome


1Division of Vascular Surgery and Institute for BioNanotechnology in Medicine, Northwestern University, Chicago, Illinois; 2Basic Research Program, SAIC-Frederick, Inc., Frederick; and 3Laboratory of Comparative Carcinogenesis, National Cancer Institute at Frederick, Frederick, Maryland

Submitted 21 February 2008; accepted in final form 14 October 2008

A enhancing vascular disease and aggressive response to injury is, in part, due to the proinflammatory and proliferative environment potentiated by increased serum levels of insulin and glucose and other biochemical and metabolic derangements (11). Investigators have demonstrated that some of the changes that occur as a result of the diabetic environment include reduced bioavailability of nitric oxide (NO) (12), increased reactive oxygen species (ROS) production (30, 46), a proinflammatory milieu (3), MAPK activation (52), protein kinase C activation (30), and activation of the receptor for advanced glycation end products (4).

In contrast, NO has been shown to possess many different vasoprotective properties (5), including inhibition of platelet aggregation (48), leukocyte chemotaxis (41), vascular smooth muscle cell (VSMC) proliferation and migration (16, 19), and endothelial cell apoptosis (51). Additionally, NO stimulates endothelial cell proliferation (56) and is a potent vasodilator (29). In sum, NO inhibits the components that lead to the development of neointimal hyperplasia in addition to reestablishing native arterial architecture. In fact, numerous investigators have verified the efficacy of NO-based therapies in small and large animal models (9, 10, 18, 25, 34, 42, 43, 49). Yet these therapies aimed at the prevention of neointimal hyperplasia have largely been studied in animal models, where normal blood vessels are subjected to injury, followed by various treatment modalities. It is unclear how the reduced bioavailability of NO, proinflammatory milieu, and increased oxidative stress associated with insulin resistance and hyperglycemia would impact these NO-based therapies. Our hypothesis is that NO will be less effective in an insulin-resistant, hyperglycemic environment. Therefore, in this paper, we sought to determine the efficacy of NO, a well-established inhibitor of neointimal hyperplasia, in the environments of type II DM and metabolic syndrome following arterial injury and...
study the mechanism by which NO exerts its effects in these hostile environments.

MATERIALS AND METHODS

Animal models. Eleven-week-old male Zucker diabetic fatty (ZDF) rats and obese Zucker (OZ) rats [both with a mutation for the leptin receptor (57)] and their genetic control lean Zucker (LZ) rats were obtained from Charles River Laboratories (Wilmington, MA). When fed the Purina 5008 diet, the inbred ZDF male rats exhibited the diabetic phenotype characterized by hyperinsulinemia, hyperglycemia, hypercholesterolemia, and hypertriglyceridemia. The outbred OZ rats when fed normal chow represented metabolic syndrome characterized by obesity, hyperinsulinemia, hypercholesterolemia, and hypertriglyceridemia. The LZ rats represented a genetic control with normal metabolic parameters.

Cell culture. Abdominal aortic VSMC were isolated and cultured from LZ, OZ, and ZDF rodent strains using the collagenase method, described by Gunther et al. (22). Cells were maintained in media containing equal volumes of DMEM-low glucose (SAFC Biosciences, Lenexa, KS) and Ham’s F12 (JRH, Lenexa, KS), supplemented with 10% fetal bovine serum (FBS; Invitrogen, Carlsbad, CA), 100 U/ml penicillin (Invitrogen), 100 μg/ml streptomycin (Invitrogen), and 4 mM l-glutamine (VWR, West Chester, PA) and incubated at 37°C, 95% air and 5% CO₂. Cells were maintained in media from LZ, OZ, and ZDF rodent strains using the collagenase method, described by Gunther et al. (22). Cells were maintained in media containing equal volumes of DMEM-low glucose (SAFC Biosciences, Lenexa, KS) and Ham’s F12 (JRH, Lenexa, KS), supplemented with 10% fetal bovine serum (FBS; Invitrogen, Carlsbad, CA), 100 U/ml penicillin (Invitrogen), 100 μg/ml streptomycin (Invitrogen), and 4 mM l-glutamine (VWR, West Chester, PA) and incubated at 37°C, 95% air and 5% CO₂. VSMC were used between passages 4 and 8 for all experiments.

Tritiated [3H]thymidine incorporation assay. [3H]Thymidine incorporation was assessed as a surrogate for cellular proliferation. Of note, cell counting was performed, and the results of cell counting correlated with [3H]thymidine incorporation. Aortic VSMC plated in 12-well plates (4 × 10⁵ cells/well) were growth arrested for 24 h, after which they were exposed to media, with or without the NO donor S-nitroso-N-acetylpenicillamine (SNAP; 250–1,000 μM), with or without the NO donor (Z)-1-[N-(2-aminoethyl)-N-(2-ammonioethyl)amino]diazan-1-ium-1,2-diolate (DETA/NO; 500–1,000 μM), with or without glucose (25 mM), with or without insulin (200 nM) in the presence of [3H]thymidine (5 μCi/ml, PerkinElmer, Wellesley, MA) for an additional 24 h. [3H]Thymidine incorporation into trichloroacetic acid-precipitated DNA was quantified by scintillation counting. SNAP releases 1 mol of NO per mole of SNAP. DETA/NO releases 2 mol of NO per mole of DETA/NO.

Cell death assay. Aortic VSMC plated in six-well plates (1 × 10⁶ cells/well) were growth arrested for 24 h, after which they were exposed to media with or without the NO donor SNAP (250–1,000 μM) in 10% FBS for 24 h. Cells were trypsinized, collected, pelleted, and then resuspended in 250-μl phosphate-buffered saline (PBS). Forty microliters of this suspension were added to 160 μl of Guava ViaCount Reagent (Guava Technologies, Hayward, CA), and cell death was assessed using Guava PCA (Guava Technologies).

Flow cytometry. Aortic VSMC plated in six-well plates (1 × 10⁶ cells/well) were growth arrested for 24 h, after which they were exposed to media with or without the NO donor DETA/NO (0–1,000 μM) for 24 h. Trypsinized cells were resuspended in 50-μl PBS and fixed with 450 μl of ice-cold 70% ethanol, followed by resuspension in a propidium iodide staining solution [1× PBS (pH 7.4), 50 μg/ml propidium iodide (Invitrogen), 204 μg/ml RNase A (Sigma, St. Louis, MO), 0.1% Triton X-100 (Fisher Biotech, Fair Lakes, NJ)]. Samples were analyzed on a Coulter Epic XL flow cytometer. Analysis was performed using ModFit 3.1 LT (Verity, Topsham, ME).

Western blot analysis. VSMC were collected after 24 h of exposure to the different treatment groups by scraping and were resuspended in 20 mM Tris with 100 μM phenylmethylsulfonyl fluoride (Sigma), 1 μM leupeptin (Sigma), and 1 μM sodium orthovanadate (Sigma). Protein was quantified with the bicinchoninic acid protein assay, according to manufacturer’s instructions (Pierce, Rockford, IL). Whole cell samples (20 μg) were subjected to sodium dodecyl sulfate-polyacrylamide gel electrophoresis on 13% gels and transferred to nitrocellulose membranes (Schleicher & Schuell, Keene, NH). Membranes were hybridized with a rabbit polyclonal anti-p27 or anti-p27 antibody (1:1,000; Santa Cruz, Santa Cruz, CA), followed by horseradish peroxidase-linked goat anti-rabbit immunoglobulin (1: 10,000; Pierce). Proteins were visualized using chemiluminescent reagents, according to the manufacturer’s instructions (Supersignal Substrate; Pierce), and the membranes were exposed to film and developed. Western blot films were scanned to JPEG images, and densitometry was performed on representative images using ImageJ (NIH, Bethesda, MD).

NO-eluting gel. The NO-eluting gel was made by combining 100 mg of disodium 1-[2-(carboxylato)pyrrolidin-1-yl]diazen-1-ium-1,2-diolate (PROLI/NO) powder with 0.6 g of polaxamer 407 (BASF, Mt. Vernon, NJ). Reacted with 2 mol of NO per mole of SNAP. DETA/NO releases 2 mol of NO per mole of DETA/NO.

Fig. 1. Nitric oxide (NO) inhibits [3H]thymidine incorporation to a greater extent in type II diabetes mellitus (DM) and metabolic syndrome compared with control. A: rat aortic vascular smooth muscle cells (VSMC) harvested from lean Zucker (LZ, control), obese Zucker (OZ; metabolic syndrome), and Zucker diabetic fatty (ZDF; type II DM) rodents were exposed to starvation media [0% fetal bovine serum (FBS)], control media (10% FBS), or varying concentrations of NO donor S-nitroso-N-acetylpenicillamine (SNAP; 0,–1,000 μM) for 24 h and [3H]thymidine incorporation was assessed. *P < 0.05 compared with 10% FBS for each group. B: LZ cells were exposed to control media or hyperglycemic (G, 25 mM) or hyperinsulinemic (I, 200 nM) environment and then treated with the NO donor (Z)-1-[N-(2-aminoethyl)-N-(2-ammonioethyl)amino]diazan-1-ium-1,2-diolate (DETA/NO). [3H]Thymidine incorporation was assessed at 24 h. *P < 0.001 vs. control; **P < 0.007 vs. DETA/NO. 0.5–1.0 mM. Each group was assessed in triplicate. Values are representative of 5 independent experiments.
Olive, NJ), as previously described (47). This mixture was then combined with 2 ml of cold PBS and placed on ice in the 4°C refrigerator. The mixture was agitated every 5 min for ~30 min until a gel formed. To measure NO release from the gel, 200 µl of the gel (the same amount placed in the neck of each rat) were place into 500 µl of PBS, with dedicated tubes set up for each time point. At the appropriate time point, the supernatant was removed, and nitrite release was measured using the Greiss reaction, as previously described (36).

Animal surgery. All animal procedures were performed in accordance with the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health (NIH Publication 85-23, 1996) and approved by the Northwestern University Animal Care and Use Committee. Rats were fasted overnight before the procedure and blood draw. Rats were anesthetized with inhaled isoflurane (0.5–3%). Atropine was administered subcutaneously (0.1 mg/kg) to decrease airway secretions. Weight was documented, and blood glucose was measured. Following a midline neck incision, the rat carotid artery balloon injury model was performed using a 2F Fogarty catheter, as previously described (36, 50). After injury and restoration of blood flow, 200 µl of the NO-eluting gel (containing 10 mg PROLI/NO) were applied evenly to the external surface of the injured common carotid artery of rats in the treatment group, and the neck incision was closed. The control group received poloxamer gel alone. Rats were killed at 2 wk (LZ n = 11, OZ n = 12, ZDF n = 13), and blood was collected to measure insulin, glucose, cholesterol, and triglyceride levels. Again, the rats were fasted overnight before this blood draw.

Tissue processing. Carotid arteries were harvested following in situ perfusion fixation with cold PBS (250 ml) and 2% paraformaldehyde (500 ml). Vessels were placed in paraformaldehyde at 4°C for 1 h and then 30% sucrose in PBS at 4°C overnight. The tissue was quick-frozen in Optimal Cutting Temperature compound (Tissue Tek, Hatfield, PA), and 5-µm sections were cut throughout the entire injured segment of the common carotid artery.

Morphometric analysis. Carotid arteries harvested at 2 wk were examined histologically for evidence of neointimal hyperplasia with hematoxylin-eosin-stained sections using a standardized methodol-

![Diagram](https://example.com/diagram.png)

**Fig. 2.** NO causes a greater G0/G1 cell cycle arrest in type II DM and metabolic syndrome compared with controls. Rat aortic smooth muscle cells harvested from LZ (control), OZ (metabolic syndrome), and ZDF (type II DM) rats were exposed to control media (10% FBS) or NO donor DETA/NO (1,000 µM) for 24 h. From left to right: representative flow cytometry analysis of control cells, then DETA/NO treated cells, followed by Modfit quantification of the percentage of cells in each stage of the cell cycle for LZ (control) (A), OZ (metabolic syndrome) (B), and ZDF (type II DM) (C) cells. *P < 0.05 compared with uninjured. Each group was assessed in triplicate. Values are representative of 4 independent experiments.
greater degree in metabolic syndrome and type II DM. To assess proliferation was conducted using [3H]thymidine incorporation in VSMC in all three rat strains compared with baseline levels and LZ controls (Fig. 1A, P < 0.05). The percent reduction between the control group and the 1,000 μM SNAP treatment group was 61 and 56% for OZ (metabolic syndrome) and ZDF (type II DM) VSMC, respectively, which was statistically greater than the 27% reduction observed in the LZ (control) VSMC (P < 0.05). Thus, while the control rats did not demonstrate a reduction in proliferation with the addition of SNAP, the inhibition of VSMC proliferation was most dra-

Apoptosis. Apoptosis was evaluated in sectioned carotid arteries harvested at 2 wk by terminal deoxynucleotidyl transferase-mediated dUTP-biotin nick-end labeling (TUNEL) using a commercially available system, according to the manufacturer’s instructions (DeadEnd Colorimetric TUNEL system; Promega, Madison, WI). The positive control consisted of a sample treated with DNase I to cause DNA fragmentation. The negative control consisted of a sample treated without the TdT enzyme.

ROS. The in situ presence of superoxide was evaluated in carotid artery sections harvested at 2 wk using the oxidative fluorescent dye dihydroethidium (DHE), as described previously (27). Five hundred microliters of DHE (2 μM) were applied to the frozen 5-μm carotid sections, cover slipped, and incubated for 30 min. Positive control consisted of sections exposed to hydrogen peroxide (200 μM). Fluorescent images of DHE staining throughout the entire cross section of the artery were obtained with the Nikon inverted microscope eclipse TE2000-U (Melville, NY) using excitation and emission wavelengths of 488 and 585 nm, respectively. DHE staining was quantified by 10 independent blinded observers using a standardized grading scale from 0 to 3 as follows: 0 = no positive staining, 1 = minimal positive staining, 2 = moderate positive staining, and 3 = intense positive staining.

Nitrotyrosine staining. Carotid arteries were examined for evidence of nitrotyrosine expression using immunoperoxidase staining. Briefly, immunohistochemistry was carried out using the HRP Polymer & DAB Plus Chromogen UltraVision LP Value Detection System (Thermo Scientific, Fremont, CA). Endogenous peroxidase activity was blocked by incubating the slides in a hydrogen peroxide blocking reagent (provided in kit) for 15 min. To block nonspecific background staining, specimens were incubated in Ultra V Block (provided in kit) for 10 min. Nitrotyrosine expression was detected using a monoclonal anti-nitrotyrosine antibody (1:200, Cayman Chemical, Ann Arbor, MI) for 1 h at room temperature. Specimens were then incubated in the Value Primary Antibody Enhancer (provided in kit) for 20 min, rinsed, and incubated in Value HRP Polymer (provided in kit) for 30 min. The specimens were incubated in DAB Plus Chromogen and DAB Plus Substrate (provided in kit) for 3 min. Counterstaining was performed with Gill 2× hematoxylin (Protocol, Fisher Scientific, Waltham, MA). For negative controls, the primary antibody was omitted.

Statistical analysis. Results are expressed as means ± SE. Differences between multiple groups for the in vitro studies were analyzed using the one-way repeated-measures ANOVA with the Student-Newman-Keuls post hoc test for all pairwise comparisons (SigmaStat; SPSS, Chicago, IL). The one-way ANOVA was used to assess significance for the in vivo animal studies. Statistical significance was assumed when P < 0.05.

RESULTS

NO inhibits [3H]thymidine incorporation in vitro to a greater degree in metabolic syndrome and type II DM. To better characterize the effects of NO on proliferation of the three strains of VSMC (LZ, OZ, ZDF), an in vitro assay to assess proliferation was conducted using [3H]thymidine incorporation. Of note, the results of these experiments on [3H]thy-
matic in VSMC harvested from rats demonstrating metabolic syndrome and type II DM.

Given that the above experiment examined the effect of NO on VSMC harvested from each animal model and not the effect in the diabetic milieu of type II DM, we wanted to evaluate the effect of NO on VSMC in a hyperinsulinemic and hyperglycemic environment. In support of the data above, NO was found to inhibit [3H]thymidine incorporation to a greater extent in LZ cells in a hyperinsulinemic (200 nM) and hyperglycemic (25 mM) environment (89% inhibition) compared with VSMC in control media with no added glucose or insulin (only 63% inhibition, \( P < 0.007 \); Fig. 1B). Thus NO is more effective at inhibiting proliferation in VSMC harvested from the animal models of type II DM, as well as in VSMC exposed to the metabolic conditions of type II DM.

**NO induces minimal VSMC cell death in vitro.** To determine whether NO induces VSMC death in vitro, death was assessed using the Guava PCA. As expected, VSMC maintained in starvation media (0% FBS) did induce a statistically significant increase in VSMC death in all three cell lines compared with VSMC maintained in regular growth media (LZ 39 ± 3.4%, OZ 36 ± 1.4%, ZDF 48 ± 0.9%, \( P < 0.05 \)). The addition of varying concentrations of SNAP (0–1,000 \( \mu \)M) to regular growth media resulted in no statistically significant impact on cell death (LZ 9 ± 1.3%, OZ 14 ± 2.3%, ZDF 14 ± 1.1%, \( P = \) nonsignificant) compared with VSMC maintained in growth media alone, as measured in the three cell lines (LZ 16 ± 3.0%, OZ 11 ± 1.5%, ZDF 9 ± 1.6%, \( P = \) nonsignificant). These data are representative of five independent experiments.

**NO causes a G0/G1 cell cycle arrest.** To further investigate the heightened efficacy of inhibition of VSMC proliferation by NO, the percentage of cells in the various stages of the cell cycle was analyzed. For all three animal models, the majority of VSMC at baseline was found in G0/G1 phase (Fig. 2). The addition of the NO donor DETA/NO resulted in an increase in the percentage of VSMC found in the G0/G1 phase, with a corresponding decrease in the percentage of cells in the G2/M and S phases. However, following exposure to DETA/NO, a larger percentage of cells was arrested in G0/G1 in OZ (80%) and ZDF (87%) vs. LZ VSMC (72%, \( P < 0.05 \)). Accordingly, the addition of DETA/NO increased the expression of the cyclin-dependent kinase cell cycle inhibitory proteins p21 in all three cell lines, but resulted in the greatest increase in p21 from baseline in ZDF cells (Fig. 3). DETA/NO increased p27 expression in all three cell lines, but no significant difference was noted between the cell types.

**NO inhibits neointimal hyperplasia more effectively in metabolic syndrome and type II DM.** We then examined the effect of NO on neointimal hyperplasia following rat carotid artery balloon injury in animal models of type II DM (ZDF, \( n = 13 \)) and metabolic syndrome (OZ, \( n = 12 \)), as well as control animals (LZ, \( n = 11 \)). Animal weights, serum glucose, insulin, cholesterol, and triglycerides from the animals are summarized in Table 1. These data confirm that the rat models exhibited a biochemical profile in accordance with their intended disease. The OZ rats, which represent metabolic syndrome, demonstrated hyperlipidemia, obesity, and insulin resistance without hyperglycemia. The ZDF rats, which represent type II diabetes, demonstrated hyperlipidemia, insulin resistance, and hyperglycemia. Due to glucose spilling and the nature of the ZDF model, these rats often are of normal weight. As expected, the addition of the NO donors did not demonstrate any significant effect on these parameters. While there was a trend toward increased insulin levels following treatment with PROLI/NO, this did not reach statistical significance.

Balloon injury produced reproducible neointimal lesions at 2 wk following injury in the rat carotid artery (Fig. 4A). Injury increased the intimal area to the greatest degree in OZ rats compared with LZ and ZDF rats (\( P < 0.05 \)). The application of PROLI/NO significantly reduced intimal area and intima-to-media area ratio (I/M) in all three rat strains compared with injury alone (\( P < 0.05 \)) (Fig. 4, B and D). Treatment with PROLI/NO resulted in a trend toward decreased medial area in all of the rat strains. However, these results were not statistically significant (Fig. 4C). The NO-based therapy showed a dramatic 93% reduction in the I/M in type II DM rats (0.42 ± 0.09 vs. injury alone, \( P < 0.05 \)) and a 69% reduction in the I/M in metabolic syndrome rats (0.08 ± 0.03 vs. injury alone, \( P < 0.05 \)). These reductions are significantly greater than the 46% reduction in I/M seen in the control rats (0.43 ± 0.07, \( P < 0.05 \)). Thus NO robustly inhibited neointimal hyperplasia in the rat models of metabolic syndrome and type II DM compared with control rats.

NO release was measured from the PROLI/NO and control gels using the Greiss reaction, which measures nitrite. The control gel contained no PROLI/NO. Similar to our laboratory’s prior publications with NO-eluting gels (33), the PROLI/NO gel released a high amount of nitrite initially, with continued release out to 6 days (Fig. 5).

**NO induces minimal VSMC apoptosis in vivo.** Similar to the in vitro studies, we sought to determine whether NO limited the formation of neointimal hyperplasia by inducing VSMC cell death.

### Table 1. Baseline characteristics of LZ (control), OZ (metabolic syndrome), and ZDF (type II diabetic) rats

<table>
<thead>
<tr>
<th></th>
<th>LZ Injury Alone</th>
<th>LZ Injury + PROLI/NO</th>
<th>OZ Injury Alone</th>
<th>OZ Injury + PROLI/NO</th>
<th>ZDF Injury Alone</th>
<th>ZDF Injury + PROLI/NO</th>
</tr>
</thead>
<tbody>
<tr>
<td>( n )</td>
<td>5</td>
<td>6</td>
<td>6</td>
<td>6</td>
<td>6</td>
<td>7</td>
</tr>
<tr>
<td>Weight (OD), g</td>
<td>323 ± 12</td>
<td>318 ± 7</td>
<td>496 ± 1*</td>
<td>502 ± 11*</td>
<td>356 ± 7*</td>
<td>359 ± 11*</td>
</tr>
<tr>
<td>Weight (Sac), g</td>
<td>336 ± 11</td>
<td>346 ± 7</td>
<td>542 ± 4*</td>
<td>543 ± 11*</td>
<td>356 ± 20*</td>
<td>389 ± 18*</td>
</tr>
<tr>
<td>Glucose (OD), mg/dl</td>
<td>122 ± 5</td>
<td>NR</td>
<td>129 ± 1</td>
<td>459 ± 30*</td>
<td>416 ± 24*</td>
<td>101 ± 19*</td>
</tr>
<tr>
<td>Glucose (Sac), mg/dl</td>
<td>108 ± 7</td>
<td>111 ± 3</td>
<td>102 ± 6</td>
<td>96 ± 5</td>
<td>467 ± 27*</td>
<td>345 ± 56*</td>
</tr>
<tr>
<td>Insulin, pM</td>
<td>41 ± 10</td>
<td>30 ± 0</td>
<td>88 ± 29*</td>
<td>113 ± 26*</td>
<td>87 ± 15*</td>
<td>101 ± 19*</td>
</tr>
<tr>
<td>Cholesterol, mg/dl</td>
<td>74 ± 4</td>
<td>71 ± 3</td>
<td>116 ± 4*</td>
<td>117 ± 5.1*</td>
<td>158 ± 4*</td>
<td>148 ± 12*</td>
</tr>
<tr>
<td>Triglycerides, mg/dl</td>
<td>85 ± 8</td>
<td>69 ± 27</td>
<td>369 ± 13*</td>
<td>370 ± 18*</td>
<td>333 ± 20*</td>
<td>366 ± 45*</td>
</tr>
</tbody>
</table>

Values are means ± SE; \( n \), no. of rats. LZ, lean Zucker; OZ, obese Zucker; ZDF, Zucker diabetic fatty; PROLI/NO, disodium 1-[2-(carboxylato)pyrrolidin-1-yl]diazen-1-ium-1,2-diolate; OD, operative date; Sac, sacrifice date; NR, not recorded. *\( P < 0.05 \) vs. LZ; †\( P < 0.05 \) vs. OZ.

_J. Appl. Physiol._ • Vol. 295 • December 2008 • www.ajpheart.org

Downloaded from http://ajpheart.physiology.org/ by 10.220.33.4 on July 1, 2017
in vivo. Sections stained using TUNEL did not reveal a distinct pattern of apoptosis among the different treatment groups (data not shown). These data are consistent with the in vitro data in which NO did not induce significant VSMC death.

NO reduces ROS more in rodent models of type II diabetes and metabolic syndrome in vivo compared with controls. To investigate the heightened efficacy of NO-mediated inhibition of VSMC proliferation, we utilized the ROS stain DHE, which predominantly detects superoxide. Balloon injury produced a reproducible increase in ROS that was inhibited by the application of NO-eluting PROLI/NO gel (Fig. 6). Injury increased the amount of ROS more in OZ and ZDF rats compared with LZ rats (P < 0.05), and the application of PROLI/NO gel significantly reduced the amount of ROS in OZ (43% reduction) and ZDF (30% reduction) rats compared with LZ rats (23% reduction) (Fig. 6B).

**DISCUSSION**

We examined the effect of NO on VSMC proliferation, cell cycle regulation, and cell death in vitro and neointimal hyper-

---

Fig. 4. NO inhibits neointima formation following arterial injury more effectively in type II DM and metabolic syndrome. NO treatment groups received a topical application of NO donor disodium 1-[2-(carboxylato)pyrrolidin-1-yl]diazen-1-ium-1,2-diolate (PROLI/NO); arteries were harvested 2 wk after injury and analyzed morphometrically. A: representative hematoxylin-eosin-stained sections from LZ (control, n = 11), OZ (metabolic syndrome, n = 12), and ZDF (type II DM, n = 13) rats: uninjured, injury alone, or injury plus application of PROLI/NO gel. Intimal area (B), medial area (C), and intima-to-media area ratios (IM) (D) are shown for rodents subjected to vascular injury alone and vascular injury followed by application of PROLI/NO gel. *P < 0.05 compared with injury alone; **P < 0.05 compared with LZ injury alone.
plasia and ROS production in vivo. In summary, we found that NO was a more effective therapy in the altered milieu of type II DM and metabolic syndrome compared with control animals, thereby disproving our hypothesis. This is important because DM and metabolic syndrome predispose patients to vascular disease. Consequently, many of these patients undergo treatment of severe vascular disease with interventions such as angioplasty, endarterectomy, and bypass grafting. Unfortunately, the vasculopathy of DM and metabolic syndrome results in greater rates of failure and restenosis after these interventions. As mentioned previously, NO is vasoprotective and inhibits many of the events that lead to the development of neointimal hyperplasia. The decreased bioavailability of NO in DM may leave these patients without physiological protection, not only for the normal day-to-day rigors of vascular activity, but also for derangements inherent to vascular interventions (3). This study demonstrates that a NO-based therapy has enhanced potency in the environments of DM and metabolic syndrome, and these data suggest a clinical role for NO-eluting vascular therapies in these disease states.

To understand the role of NO in vessels from DM and metabolic syndrome, a better understanding of the pathophysiology of diabetic vascular disease must be gained. How insulin resistance and hyperglycemia create a pathological vascular environment in DM is not completely understood, but extensive research has elucidated several factors that contribute to the vasculopathy seen in DM. The bioavailability of NO represents a key marker in vascular health. Endothelial cells constitutively produce NO, and NO has many beneficial effects on the blood vessel. However, endothelial dysfunction, which is associated with impaired endothelium-dependent NO production, has been demonstrated in experimental models of diabetes, as well as in patients (17, 54). This dysfunction results from multiple derangements, but ROS play a predominant role. The diabetic environment activates systems like the MAPK pathway, protein kinase C, receptor for advanced glycation end products, and nuclear factor-κB in VSMC that augment the production of superoxide (6). Furthermore, insulin directly stimulates superoxide production through activation of NADPH oxidase, and glucose directly blocks electron transfer in the mitochondria, both of which result in further superoxide production (23, 46, 55). The increased superoxide, in turn, results in further decreases of NO production by uncoupling endothelial NO synthase (eNOS); uncoupled eNOS in turn produces more superoxide (2). Lastly, hyperinsulinemia stimulates angiotensin II, which results in inhibition of Akt (39). This further diminishes production of NO by preventing phosphorylation of eNOS by Akt (15). Thus this hyperinsulinemic and hyperglycemic environment is fraught with a decreased bioavailability of NO and a heightened ROS presence, both of which contribute to the altered response to vascular injury (7).

Oxidative stress has been shown to stimulate VSMC proliferation and lead to greater neointimal hyperplasia (20, 21). Jacobson et al. (31) delivered a NADPH oxidase inhibitor to rodents systemically following arterial injury. They observed decreased superoxide levels in the arterial wall, as well as diminished neointimal hyperplasia. Dourron et al. (14) delivered the NADPH oxidase inhibitor to the periadventitial surface of the artery, similar to our present study, and also demonstrated reduced superoxide levels in the arterial wall and decreased neointimal hyperplasia at 2 wk. Tempol, a ROS scavenger, has also reduced neointimal hyperplasia in diabetic rodents (32). Thus it is clear that ROS play an important role in the arterial injury response. Since hyperinsulinemia and hyperglycemia are known to stimulate ROS production directly and indirectly, and less NO is present, the arterial wall is at a significantly disadvantaged state. Small changes in the arterial injury response, especially in the early phase, may result in significant changes in the end biological response. Thus the provision of excess NO, which decreases production and increases degradation of ROS, to the arterial wall following injury can throw off the negative balance and shift the environment to a more vasoprotective state. However, the effect of NO on ROS is unlikely to be the only mechanism by which NO is more effective in the diabetic milieu, since NO is known to affect many different aspects of the arterial injury response, including platelets, leukocytes, VSMC, endothelial cells, and fibroblasts. Further study is warranted to delineate these different regulatory pathways.

While we are unaware of any studies that have investigated the efficacy of a NO-based therapy to inhibit neointimal hyperplasia in a diabetic environment, Barbato et al. (3) examined the effect of inducible NO synthase (iNOS) gene transfer in rodents exhibiting metabolic syndrome. They demonstrated that NO inhibited neointimal hyperplasia to a greater extent in metabolic syndrome rats, which showed a 66% reduction in the I/M after infection with an adenovirus carrying the iNOS gene, while control rats demonstrated a 50% reduction with NO (3). The vascular injury response in the face of metabolic syndrome was associated with increased adhesion molecule expression, inflammatory cell infiltration, and proliferation (3). In their study, iNOS was able to inhibit this heightened injury response and reduce neointimal formation in this proinflammatory environment (3). In fact, our reduction in neointimal hyperplasia with the application of NO in the environment of metabolic syndrome (69%) is consistent with that observed by Barbato et al. (3) (66%).

Our present study is not without limitations. Our results verify the efficacy of NO in inhibiting neointimal hyperplasia in the type II DM rodent arterial injury model. However, it remains to be seen what effect NO would have in a large-animal model of type II diabetes and metabolic syndrome. Another limitation to our study is that we used two different classes of NO donors: S-nitrosothiols (i.e., SNAP) and diaze-
NITRIC OXIDE-BASED THERAPIES IN DIABETES MELLITUS

However, all three NO donors reliably release NO, and our aim is to examine the effect of NO on neointimal hyperplasia in the midst of insulin resistance and hyperglycemia. One mole of NO is released per 1 mol of SNAP, while 2 mol of NO are released per 1 mol of DETA/NO. We have experience using S-nitrosothiols and diazeniumdiolates for in vitro and in vivo experiments; both inhibit VSMC proliferation and induce G0/G1 cell cycle arrest, with no significant differences between them (33, 35, 37, 47). Furthermore, diazeniumdiolates were utilized for the present in vivo experiments due to our laboratory’s previously published studies and, therefore, our familiarity with their in vivo characteristics (35, 51). DETA/NO was chosen instead of PROLI/NO for the in vitro studies simply because of its longer half-life in solution at 37°C and pH 7.4 (half-time = 20 h vs. 2 s, respectively). PROLI/NO would be an inappropriate choice for a 24-h in vitro assay. Lastly, while our gel produced significant results in the short term, the long-term results of NO application and the use of improved vehicles of NO delivery (like NO-eluting stents) should be researched further.

Another aspect of our study that deserves discussion is the efficacy of periadventitial application of the NO-eluting gel to the surface of the artery. We, and others, have previously shown that periadventitial application of a NO donor is effective at inhibiting neointimal hyperplasia at 2 wk (9, 18, 33, 34, 43, 47). The efficacy of this approach is most likely due to the highly diffusible nature of NO, allowing this molecule to...
penetrate through the layers of the arterial wall, reaching even the inner endothelial cell layer. However, it has also been established that the adventitial fibroblasts contribute significantly to the development of neointimal hyperplasia. Thus periadventitial application has the added benefit of greatly affecting the adventitial fibroblasts, as a diffusion gradient is likely to exist throughout the wall. Lastly, we previously demonstrated that the short-acting PROLI/NO was more effective at inhibiting neointimal hyperplasia compared with the long-acting NO donor diazeniumdiolated poly(acrylonitrile) (47). We hypothesized that this was due to the larger release of NO initially from PROLI/NO compared with the long-acting NO donor. This large release would have an overall greater effect on the arterial injury cascade, given that the injury response begins within seconds to minutes following injury with platelet aggregation and leukocyte chemotaxis.

Lastly, how a short-acting NO donor such as PROLI/NO can have such a lasting effect on the arterial wall should be addressed. At physiological pH and temperature, low concentrations of PROLI/NO have been shown to have a very short half-life, on the order of seconds (53). However, we have shown in this paper, as well as prior publications, that, when shielded inside a gel at higher basic concentrations, PROLI/NO has an extended half-life of NO release (33). This is not surprising, given the kinetics of NO release from diazeniumdilates. They are highly sensitive to pH, temperature, concentration, and the surrounding milieu of hydrogen ions (28). Furthermore, the gel acts to shield the PROLI/NO from hydrogen ions, thereby prolonging the half-life even further. Lastly, while it is difficult to equate the NO release from the PROLI/NO gel in vivo to the NO release from the NO donors in vitro, the highest concentration of NO delivered to the cells in vitro (i.e., 1 mM) is ~1/20th as much PROLI/NO as is placed in the neck of the animals. Thus the amount of NO delivered to the carotid artery in vivo is reasonable and may even be on the low side, given the surface area of cells exposed to NO in vitro. Ultimately, however, our goal is the end biological effect, i.e., inhibition of neointimal hyperplasia, so that a therapy can be developed and used in all patients, regardless of their disease states, to prevent restenosis following vascular interventions.

In conclusion, periadventitial application of a NO-eluting gel onto the surface of an injured rat carotid artery inhibited neointimal hyperplasia more effectively in animal models of type II DM and metabolic syndrome compared with controls. While PROLI/NO inhibited neointimal hyperplasia in all three rodent models, the inhibition was nearly twice as great in type II DM vs. control. Before this therapy can be utilized in the clinical arena, it must be evaluated in the large-animal model, and long-term studies should be conducted. Overall, this therapy has promising clinical potential for an ever-growing population of patients with diabetic vascular disease.

ACKNOWLEDGMENTS

The authors express thanks to the Northwestern University Institute for BioNanotechnology in Medicine and to Lynnette Dangerfield for administrative support.

GRANTS

This work was supported in part by funding from the NIH (1K08HL084203 to M. Kibbe); the American Vascular Association, the Department of Veterans Affairs (VA Merit Review Grant to M. Kibbe); the American Medical Association Foundation (2007 Seed Grant to S. S. Ahanchi); and through the generosity of Hilda Rosenbloom and Eleanor Baldwin. In addition, part of this research was supported with federal funds from the National Cancer Institute, NIH under contract NO1-CO-12400 with SAIC-Frederick, Inc. (L. K. Keefer), and by the Intramural Research Program of the NIH, National Cancer Institute, Center for Cancer Research (L. K. Keefer).
REFERENCES


