Cyproheptadine prevents pergolide-induced valvulopathy in rats: an echocardiographic and histopathological study

Steven Droogmans,1,4 Bram Roosens,1,4 Bernard Cosyns,1,4 Céline Degaillier,3 Sophie Hernot,4 Caroline Weytjens,1,4 Christian Garbar,3 Vicky Caveliers,2,4 Miriam Pipeleers-Marichal,3 Philippe R. Franken,4 Tony Lahoutte,2,4 Danny Schoors,1 and Guy Van Camp1,4

Departments of 1Cardiology, 2Nuclear Medicine, and 3Pathology, Universitair Ziekenhuis Brussel, and 4In Vivo Cellular and Molecular Imaging, Faculty of Medicine and Pharmacy, Vrije Universiteit Brussel, Brussels, Belgium

Submitted 11 November 2008; accepted in final form 2 April 2009

Droogmans S, Roosens B, Cosyns B, Degaillier C, Hernot S, Weytjens C, Garbar C, Caveliers V, Pipeleers-Marichal M, Franken PR, Lahoutte T, Schoors D, Van Camp G. Cyproheptadine prevents pergolide-induced valvulopathy in rats: an echocardiographic and histopathological study. Am J Physiol Heart Circ Physiol 296: H1940–H1948, 2009. First published April 3, 2009; doi:10.1152/ajpheart.01177.2008.—Serotonergic drugs, such as pergolide, have been associated with the development of cardiac valvular myxoid thickening and regurgitation in humans and more recently in rats. These effects are potentially mediated by the 5-hydroxytryptamine (5-HT2B) receptor (5-HT2BR). Therefore, we sought to determine whether cyproheptadine, a 5-HT2B antagonist, might prevent toxic valvulopathy in an animal model of pergolide-induced valvular heart disease. For this purpose, 50 male Wistar rats received daily intraperitoneal injections of pergolide (0.5 mg/kg, n = 14), pergolide (0.5 mg/kg) combined with cyproheptadine (10 mg/kg, n = 12), cyproheptadine (10 mg/kg, n = 12), or no injections (control, n = 12) for 20 wk. Echocardiography was performed blindly at baseline and at 10 and 20 wk followed by pathology. At baseline, no differences between groups were found with echocardiography. At 20 wk, aortic regurgitation was present in all pergolide-treated animals, whereas it was less frequently observed in the other groups (P < 0.0001). For the other valves, this difference was less pronounced. On histopathology, not only aortic but also mitral valves were thicker, myxoid, and exhibited more 5-HT2B-positive cells in pergolide-treated animals compared with the other groups. Moreover, regurgi
tant aortic and mitral valves were thicker than nonregurgitant aortic and mitral valves. In conclusion, we found that cyproheptadine prevented pergolide-induced valvulopathy in rats, which was associated with a reduced number of 5-HT2B-positive valvular cells. This may have important clinical implications for the prevention of serotonergic drug-induced valvular heart disease.

Address for reprint requests and other correspondence: S. Droogmans, Dept. of Cardiology, Universitair Ziekenhuis Brussel, Laarbeeklaan 101, Brussels 1090, Belgium (e-mail: steven_droogmans@yahoo.com).
Drug Preparation and Administration

Pergolide mesylate (Sigma-Aldrich) was prepared in a 10% ethanolic solution in saline at a concentration of 0.5 mg/ml. Cyproheptadine hydrochloride sesquihydrate (Sigma-Aldrich) was prepared in 25% DMSO (WWR) solution in aqua ad injectabilia at an initial concentration of 40 mg/ml. In the first 2 wk, four animals died of intestinal obstruction in the cyproheptadine-treated groups. This was probably due to the anticholinergic effect at that dosage. Therefore, the concentration was decreased to 10 mg/ml after 2 wk. The treatment identity of the rats was hidden during echocardiography and pathological evaluation.

Echocardiography

Ten minutes before being imaged, the rat was anesthetized with 50 mg/kg pentobarbital sodium (Nembutal, CEVA, Brussels, Belgium) intraperitoneally. Subsequently, the anterior chest wall was shaved, and the rat was placed in left lateral decubitus on a wooden bench to obtain optimal image quality and views, as previously described (3, 32).

An Vivid 7 Pro system (GE Medical Systems, Milwaukee, WI, USA) with a 10-MHz neonatal probe (10S) and 13-MHz linear probe (i13L) was used. Detailed acquisition and processing regarding regurgitant jet evaluation and quantification of left ventricular (LV) systolic and diastolic function have been previously reported (4). Briefly, regurgitant jets were assessed visually in all possible views. The color-Doppler ratio of the regurgitant aortic jet width to the LV outflow tract diameter and the mitral and tricuspid valve regurgitant area ratio (jet/atrium) were calculated. The relative duration of the regurgitant jet on color M-mode to diastole (for pulmonary and aortic valves) or systole (for tricuspid and mitral valves) was also determined. LV volumes were calculated from the M-mode tracings in a short-axis view (papillary muscle level, average of 3 cycles) using the following ellipsoid model: \( \pi \times D^3/3 \) (where \( D \) is the diameter of the ventricle in the short-axis view) (23). The LV ejection fraction (LVEF) was determined from the LV end-diastolic volume (LVEDV) and LV end-systolic volume (LVESV) as follows: LVEF = \((LVEDV - LVESV)/LVEDV\) \times 100. Cardiac output was calculated as stroke volume \( \times \) heart rate, with stroke volume equal to LVEDV minus LVESD. The fractional area change (FAC) was calculated by the LV cross-sectional area at end diastole (LVAd) and LVEDV minus LVESD. The fractional area change (FAC) was determined. LV volumes were calculated from the M-mode tracings in a short-axis view (papillary muscle level, average of 3 cycles) using the following ellipsoid model: \( \pi \times D^3/3 \) (where \( D \) is the diameter of the ventricle in the short-axis view) (23). The LV ejection fraction (LVEF) was determined from the LV end-diastolic volume (LVEDV) and LV end-systolic volume (LVESV) as follows: LVEF = \((LVEDV - LVESV)/LVEDV\) \times 100. Cardiac output was calculated as stroke volume \( \times \) heart rate, with stroke volume equal to LVEDV minus LVESD. The fractional area change (FAC) was calculated by the LV cross-sectional area at end diastole (LVAd) and end systole (LVAs) using the following equation: FAC (in %) = \((LVAd - LVAs)/LVAd\) \times 100 (30). The following LV diastolic parameters were also assessed: peak early velocity, peak late velocity, and mitral medial diastolic annulus velocity.

Influence of Drug Administration on Blood Pressure During Echocardiography

Blood pressure was measured with the advanced autoinflated blood pressure monitor with an infrared sensor and tail cuff from Harvard Apparatus in a subset of 32 rats (8 rats/group). Systolic and diastolic blood pressures were measured after 10 and 20 min after the pentobarbital (50 mg/kg ip) injection. Pentobarbital was administered 30 min after injection of the study drug.

Histopathology

At the end of the experiment, animals were killed with 120 mg/kg pentobarbital sodium intravenously. Hearts were excised, weighed, and fixed in neutral buffered formalin (10%) for 2 h. Hearts were then embedded in paraffin and cut in an axial plane (from base to apex). Three ranges of sections (4–6 \( \mu \)m thickness) were carried out with \( \sim 100 \mu \)m between each range to visualize the maximum amount of valvular tissue, and slides were stained with hematoxylin, eosin, safranin (HES), alcin blue, and Masson’s trichrome. Immunohistochemical staining for 5-HT\(_{3}\)Rs was performed with rabbit anti-5-HT\(_{3}\)R antibody (10 \( \mu \)g/ml, n\(^{a}\)b13292, Abcam, Cambridge, MA) according to the manufacturer’s instructions and as done by Elangbam et al. (8). After incubation with biotinylated donkey anti-rabbit IgG (dilution \( \times 300 \), n\(^{a}\)RP10004, Amersham Biosciences), an alkaline phosphatase detection system (Vectastain ABC-AP standard kit) was used with fucins + substrate chromogen (Dako) without counterstaining. Morphometry was performed by digital image analysis using a personal computer digital image camera (Digital Sight DS-5M, Nikon) mounted on an Axio Lab Zeiss light microscope (Carl Zeiss). We used the NIH Image program (Image-J 1.35d, NIH, Bethesda, MD). The program was calibrated with a graduated slide. Microscopic images were used to blindly evaluate cardiac valves and cardiomyocytes. The maximum thickness of each valve was measured. The width of at least 10 LV cardiomyocytes was measured in each section. Semi-quantitative compositional analysis of the valves was carried out for glycosaminoglycans (GAGs; alcinian blue) and collagen (trichrome). Therefore, microscopic images were taken under the same conditions (magnification: \( \times 100 \)) and were converted to 8-bit images. An appropriate threshold was determined on one slide for GAGs and collagen content, respectively, and subsequently kept constant. Areas of interest were drawn around the valvular leaflets, and the proportion of GAG to collagen was calculated. For immunohistochemistry, the absolute number of 5-HT\(_{3}\)R-positive cells in valvular leaflets were counted in microscopic images taken at a magnification of \( \times 200 \).

Statistical Analysis

Data are presented as means \( \pm \) SD. Comparisons between groups were performed using the Mann-Whitney U-test for continuous variables and Fisher’s exact test for categorical variables. All \( P \) values were calculated two tailed. \( P \) values of \( <0.05 \) were considered significant. Statistical analysis was done with SPSS software (version 16.01, SPSS, Chicago, IL).

RESULTS

General Characteristics

In the pergolide + cyproheptadine-treated group, three animals died (two animals within 2 wk and one animals at week 4), whereas three animals in the cyproheptadine-treated group died (two animals within 2 wk and one animals at week 9). At necropsy, the four rats that died before the first 2 wk showed an intestinal obstruction at the level of the colon due to fecal impaction. In the other two rats, no clear cause of death could be detected. After a reduction to 10 mg/kg, cyproheptadine was well tolerated by the other animals. Animals that received pergolide showed hyperactivity, poor grooming, aggressive behaviour, and increased gnawing activity, which was more pronounced during the first week of the study. This was also seen in the pergolide + cyproheptadine-treated group. On the other hand, the cyproheptadine-treated group showed no general behavioral changes.

Echocardiography

Valves. All valves were visualized with echocardiography during the study. At baseline, there were no differences in valvular regurgitation between groups (data not shown). The occurrence and severity of valvular regurgitation in the different groups are shown in Tables 1 and 2. One rat in the cyproheptadine-treated group at 20 wk was excluded from the echographic analysis because of too-deep sedation leading to bradycardia. At 20 wk, aortic regurgitation was present in all 14 pergolide-treated rats compared with 1 rat in the pergolide + cyproheptadine-treated group (11%), 2 rats in the cyproheptadine-
treated group (25%), and 1 rat in the control group (8%, \( P < 0.0001 \)). Aortic regurgitation was already more frequent at 10 wk in pergolide-treated rats compared with the other groups.

At 20 wk, mitral regurgitation was found in seven rats in the pergolide-treated group (50%) compared with four rats in the pergolide + cyproheptadine-treated group (44%, \( P = 1.00 \)), none in the cyproheptadine-treated group (0%, \( P = 0.022 \)), and four rats in the control group (33%, \( P = 0.453 \)). Although the mean mitral regurgitant area ratio was not different between groups, the mean relative duration of the mitral regurgitation to systole was higher in the pergolide-treated group compared with the other groups (Table 2).

Tricuspid and pulmonary regurgitation were already found in the cyproheptadine-treated group (44%, \( P = 0.108 \)), and control (25 ± 18/81 ± 12 mmHg) rats (mean blood pressure was measured after 10 and 20 min after the pentobarbital injection, given 30 min after the study drug).

**Pathology**

*Postmortem body weight.* Body weights of pergolide-, pergolide + cyproheptadine-, and cyproheptadine-treated animals (540 ± 44 g, \( P = 0.003 \); 519 ± 61 g, \( P = 0.001 \); and 523 ± 51 g, \( P = 0.002 \), respectively) were lower compared with control animals (597 ± 30 g).

**Valves.** On histopathological examination, pergolide-treated rats had thickened aortic and mitral valves compared with the other groups. Aortic and mitral valve thickening were prevented with cyproheptadine (Fig. 1). The tricuspid valve was not clearly affected by pergolide. In addition, regurgitant aortic and mitral valves were thicker than nonregurgitant aortic (273 ± 176 vs. 180 ± 84 \( \mu \)m, \( P = 0.029 \)) and mitral (419 ± 152 vs. 260 ± 118 \( \mu \)m, \( P = 0.003 \)) valves (Fig. 2).

Pergolide-treated animals exhibited an increased number of 5-HT_{3b}R-positive interstitial and endothelial cells in the aortic and mitral leaflets (Fig. 3) compared with the other groups. Most of the 5-HT_{3b}R-positive cells were scattered throughout the fibromyxoid sponge layer.

**Table 1. Occurrence of TR, PR, MR, and AR at 10 and 20 wk in all groups**

<table>
<thead>
<tr>
<th></th>
<th>Cyproheptadine</th>
<th>Pergolide + Cyproheptadine</th>
<th>Pergolide</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Control, %</td>
<td>P value vs. control</td>
<td>P value vs. control</td>
</tr>
<tr>
<td></td>
<td>P value vs. control</td>
<td></td>
<td></td>
</tr>
<tr>
<td>10 wk</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>TR</td>
<td>33</td>
<td>1.00</td>
<td>30</td>
</tr>
<tr>
<td>PR</td>
<td>33</td>
<td>0.338</td>
<td>20</td>
</tr>
<tr>
<td>MR</td>
<td>25</td>
<td>0.229</td>
<td>30</td>
</tr>
<tr>
<td>AR</td>
<td>0</td>
<td>1.00</td>
<td>0</td>
</tr>
<tr>
<td>20 wk</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>TR</td>
<td>42</td>
<td>0.650</td>
<td>56</td>
</tr>
<tr>
<td>PR</td>
<td>58</td>
<td>0.650</td>
<td>44</td>
</tr>
<tr>
<td>MR</td>
<td>33</td>
<td>0.117</td>
<td>44</td>
</tr>
<tr>
<td>AR</td>
<td>8</td>
<td>0.537</td>
<td>11</td>
</tr>
</tbody>
</table>

Shown are percentage of rats with valvular regurgitation in each group as well as the corresponding \( P \) values. TR, tricuspid regurgitation; PR, pulmonary regurgitation; MR, mitral regurgitation; AR, aortic regurgitation. *Statistically significant difference.

**Table 2. Severity of valvular regurgitation in the different treatment groups at 20 wk**

<table>
<thead>
<tr>
<th></th>
<th>Cyproheptadine</th>
<th>Pergolide + Cyproheptadine</th>
<th>Pergolide</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Control, means ± SD</td>
<td>Means ± SD</td>
<td>P value vs. control</td>
</tr>
<tr>
<td>AR jet width/LVOT</td>
<td>23</td>
<td>29 ± 6</td>
<td>0.667</td>
</tr>
<tr>
<td>Relative duration of AR to diastole, %</td>
<td>17</td>
<td>17 ± 1</td>
<td>1.00</td>
</tr>
<tr>
<td>Mitral regurgitant area ratio</td>
<td>11 ± 3</td>
<td>17 ± 1</td>
<td>1.00</td>
</tr>
<tr>
<td>Relative duration of MR to systole, %</td>
<td>12 ± 3</td>
<td>16 ± 2</td>
<td>0.841</td>
</tr>
<tr>
<td>Tricuspid regurgitant area ratio</td>
<td>9 ± 3</td>
<td>9 ± 2</td>
<td>0.841</td>
</tr>
<tr>
<td>Relative duration of TR to systole, %</td>
<td>14 ± 3</td>
<td>16 ± 2</td>
<td>0.286</td>
</tr>
<tr>
<td>Relative duration of PR to diastole, %</td>
<td>19 ± 3</td>
<td>22 ± 4</td>
<td>0.267</td>
</tr>
</tbody>
</table>

LVOT, left ventricular (LV) outflow tract diameter. *Statistically significant difference.
Valvular thickening by pergolide was due to myxoid transformation of the sponge layer with an increase of the GAG-to-collagen ratio (Fig. 4). This was most pronounced in the mitral valve. GAG deposits were also observed in the other groups but were smaller and localized at the distal free edge of the valvular leaflets. In contrast, pergolide-treated rats showed mostly diffuse thick myxoid changes reaching the base of the cusps. Chondroid metaplasia was found in all groups.
the attachments side of the left coronary aortic cusp and anterior mitral leaflet.

LV myocytes. LV myocytes were thicker in the pergolide-treated group compared with the other groups. LV fibrosis was not found (Fig. 4).

**DISCUSSION**

In this study, we found that cyproheptadine had a preventive effect against the development of pergolide-induced valvulopathy in a rat model. This was demonstrated by a reduction of incidence of aortic regurgitation on echocardiography and by less myxoid aortic and mitral thickening on histology. This effect was related to a reduction of 5-HT2BR-positive valvular cells. Our data further indicated that echocardiography was a reliable tool to assess the presence of pergolide-induced valvulopathy and follow treatment with cyproheptadine and also correlated well with histopathology.

Cyproheptadine protected against pergolide-induced aortic regurgitation in this study, whereas its effect on the incidence of mitral regurgitation was not significant. This could be due to the relatively lower frequency of mitral regurgitation detected in the pergolide-treated group compared with previous work (50% vs. 67%) (3). On the other hand, the relative duration of mitral regurgitation was more pronounced in pergolide-treated animals compared with the other groups. On histopathology, pergolide + cyproheptadine-treated rats had clearly less affected aortic and mitral valves compared with the pergolide-treated group, which demonstrates the protective effect of cyproheptadine on left-sided heart valves. Moreover, on histology, regurgitant aortic and mitral valves were thicker than nonregurgitant valves. The absence of a significant change in blood pressure between the different groups suggests that the observed valvular regurgitations are not importantly determined by blood pressure and rather due to true valvular lesions.

5-HT3AR activation is a common feature of all valvulopathic drugs (or their metabolites), whereas this was not found with drugs not associated with valvular heart disease (10, 25, 28, 29). Moreover, some of these valvulopathic drugs have been shown to induce mitogenesis in cultured interstitial cells from human cardiac valves by activating the 5-HT2BR (28). In the present study, we found that cyproheptadine reduced the number of 5-HT2BR-positive cells in the aortic and mitral valvular leaflets when given with pergolide, which further suggests the involvement of the 5-HT3AR in the pathogenesis of drug-induced valvulopathy. This is also strengthened by the observation that chemically similar drugs, such as lisuride and terguride, which are agonists for 5-HT2CRs and 5-HT2ARs and antagonists for 5-HT2BRs, were not associated with valvulopathy (15, 24). Similarly, no increase in the risk of valvulopathy was observed in patients treated with the nonergot-derived dopamine agonist pramipexole, which has a low affinity for the 5-HT2BR (26, 34). Moreover, 5-HT injections in rats also led to increased 5-HT2BR mRNA expression in the affected valves (7). In another animal study (13), terguride inhibited the long-term hypertrophic organ effects of 5-HT. Young et al. (33) recently published a full binding profile of cyproheptadine. Besides the 5-HT3AR, cyproheptadine also antagonizes several other 5-HT receptors. Hence, the exact mechanism by which cyproheptadine prevented pergolide valvulopathy in this study needs to be further investigated.

We could not observe a clear toxic effect of pergolide on the tricuspid valve, as it was less affected in histology and echocardiography. Moreover, toxic right-sided valvular heart disease is more difficult to diagnose and evaluate, as functional (physiological) regurgitations are more pronounced in both humans and rats. This is also true for the pulmonary valve. With our technique, this valve was very difficult to visualize on histopathology because of its more anterior-superior position.
As in our previous study (3), no sufficient pulmonary valves could be histologically evaluated to draw meaningful conclusions.

In aging rats, spontaneous nodular and segmental myxoid valvular thickening has been described (5, 6). This was also found in the control rats of this study. However, compared with the pergolide-treated group, valvular thickening was less pronounced and more localized toward the distal free edges of the valvular leaflets. This can as well explain the progressive increase in the number of tricuspid, pulmonary, and mitral regurgitations in control rats during the study. On the other hand, serotonergic compounds such as serotonin and pergolide led to fibroblast stimulation with a more pronounced and diffuse accumulation of GAGs throughout the entire valve. Similar histological changes have also been described in anorexigen-induced valvulopathy and floppy mitral valves in humans, as these valves contained more GAGs than normal (16). Several studies (3, 7, 12) have shown that serotonin injections also led to a similar valvulopathy in rats. A recent study (7) by Elangbam et al. has shown that rats treated with high-dose 5-HT exhibited, after only 7 days, thickened aortic and mitral valves with increased GAG and decreased collagen content. In humans, normal valves are composed of equal amounts of GAG and collagen (17). Similar observations were made in this study, where the thickened valves (especially the mitral valve) of pergolide-treated rats had an increased GAG-to-collagen content, whereas the other groups had an equal distribution of GAG and collagen.

Besides functional and morphological valvular alterations, pergolide also gave rise to myocardial hypertrophy without fibrosis. This was accompanied with a decrease of LV function. Similar findings were observed in our previous study (3). The absence of LV hypertrophy in the pergolide/cyproheptadine-treated group may be explained by the 5-HT2BR antagonistic properties of cyproheptadine, since selective blocking of this receptor also prevented isoproterenol- and angiotensin II-induced cardiac hypertrophy (14, 19). In clinical practice, ergotamine-derived dopamine agonists have not been associated with LV dysfunction (21). This apparent contradiction might be due to the relative high dosage of pergolide injected in these animals (21). Also, body weights in cyproheptadine-treated animals were lower than in control animals, which was not expected (11). This could be due to the intraperitoneal administration of cyproheptadine, which might have led to a de-

Fig. 3. Top: 5-hydroxytryptamine (5-HT)2B receptor (5-HT2BR) immunohistochemistry of the aortic and mitral valvular leaflets in the different treatment groups. 5-HT2BR-positive cells are red (arrows). Scale bar = 50 μm. Magnification: ×200. Bottom: absolute numbers of 5-HT2BR-positive cells per ×200 field in the aortic and mitral valvular leaflets were higher in the pergolide-treated group. *P < 0.05; **P < 0.01.
creased intestinal absorption of nutrients in these rats by its anticholinergic effect. This administration route was chosen to give a corrected dosage at the same time. Although the cyproheptadine-treated animals had lower body weights, this did not influence valvular function and morphology, which confirms the usefulness of this rat model for studying drug-induced valvular heart disease.

We have to acknowledge the limitation that no separate control groups for each vehicle were studied. However, in our previous study (3), neither injections with 10% ethanolic solution (pergolide vehicle) nor injections with physiological saline led to valvular regurgitations or thickening. Moreover, in a pilot study (dose finding for cyproheptadine and DMSO), we did not observe valvular toxicity in rats only receiving DMSO. Also, DMSO is considered a relative safe vehicle with a low or no toxicity profile. In agreement, no valvular toxicity was found in the cyproheptadine-treated group (containing DMSO).

Cyproheptadine is a cheap, safe, off-patent, and widely available drug currently mainly used for its antihistaminic, antiallergic properties (20). It also inhibits several 5-HT receptors (33), which is useful for the symptomatic treatment of metastatic carcinoid syndrome (18). It has even been used for the treatment of neuroleptic-induced akathisia because of its anticholinergic properties (9). Therefore, cyproheptadine might be useful as a prophylactic agent for pergolide-induced valvular heart disease, all the more so, because it lacks and even counteracts the extrapyramidal side effects induced by atypical antipsychotic drugs (3). In addition, the combination of pergolide and cyproheptadine is often used for the treatment of Cushing disease in horses (27).

The findings of this study may have important implications for the practical use of serotonergic compounds such as anti-obesity drugs, amphetamine derivatives [i.e., 3,4-methylenedioxymethamphetamine (“Ecstasy”)], antimigraine drugs (ergotamine), and Parkinson’s disease (pergolide and cabergolide) treatments. These drugs have some unique therapeutic effects but have been associated with valvular heart disease (2, 26, 31). The simultaneous use of these drugs with a 5-HT antagonist, such as cyproheptadine, may prevent the develop-
ment toward toxic valvulopathy. Moreover, by blocking the 5-HT receptor of valvular fibroblasts, further deterioration of existing drug-induced valvular heart disease might be stopped and perhaps regression may occur. In that way, it might facilitate the regression of valvular heart disease even after removal of the causal drug. Of course, these hypotheses need to be confirmed in prospective clinical trials.

In conclusion, we found that cyproheptadine prevented pergolide-induced valvulopathy in a rat model, which was associated with a reduced number of 5-HT<sub>2B</sub>-R-positive valvular cells. These findings may have important clinical implications for treatments with serotoninergic drugs.

**GRANTS**

S. Droogmans is an Aspirant of the Research Foundation-Flanders [Fonds Wetenschappelijk Onderzoek (FWO)]. T. Lahoutte is a Senior Clinical Investigator of the Research Foundation-Flanders (Belgium). The research at In Vivo Cellular and Molecular Imaging is funded by the Interuniversity Attraction Poles Programme (Belgian State, Belgian Science Policy).

**REFERENCES**


