Determinants of valve gating in collecting lymphatic vessels from rat mesentery

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LYMPH PROPULSION IS DETERMINED by the interplay of active and passive forces that combine to propel lymph centrally against a pressure gradient (28, 44, 58). Active lymph propulsion is achieved by the spontaneous, rhythmic contractions of collecting lymphatic vessels (21, 30, 38, 55). Passive factors influencing lymph flow include Starling forces that favor net fluid reabsorption at the level of the lymphatic capillaries, tissue compression from the contractions of nearby skeletal muscle fibers (19, 45), respiratory movements (7, 41, 43), and gravitational forces (45, 50).

Lymphatic valves are essential for minimizing backflow of lymph, and two systems of valves have been described previously (50). Primary lymphatic valves are composed of the overlapping, but discontinuous, junctions of endothelial cells in lymphatic capillaries (8, 9, 40), which function to facilitate preferential movement of interstitial fluid into initial lymphatics. In contrast, secondary lymphatic valves are typically bicuspid structures, spaced every few millimeters along collecting lymphatic vessels. Valve location and development closely correspond with the presence of a muscle cell layer in the vessel wall. Each semilunar valve cusp is lined on both sides by a layer of endothelial cells and anchored at its base by collagen fibers in the wall. The downstream insertion point of a valve leaflet is anchored to the wall by collagen and elastin (1, 26, 54), forming a buttress to prevent inversion when output pressure is elevated (50). Competent secondary valves are essential for ensuring the net proximal flow of lymph toward the heart. A series of dysfunctional valves would exacerbate lymphedema by allowing lymph reflux into peripheral networks of lymphatic vessels, thus promoting elevated hydrostatic pressure in the initial lymphatics, which, in turn, would retard fluid absorption from the interstitium. The consequences of insufficient or malformed secondary valves are readily apparent in genetic conditions involving lymphatic valve malformation (6, 47).

Secondary lymphatic valves are thought to gate passively according to the instantaneous luminal pressure gradient (36, 50), opening and closing with each lymphatic contraction cycle, as fluid is transferred from upstream to downstream lymphangions (18, 59). In some species, a role for active force development by specialized smooth muscle cells or actin filament extensions from these cells into the leaflets (4, 34) has been postulated to facilitate valve closure (5, 31); however, this view is based almost exclusively on structural rather than functional evidence (50). Studies of lymphatic valve ultrastructure abound (1, 26, 54), but we are unaware of published measurements of the forces required to gate secondary lymphatic valves. A passive mechanism for gating is inferred from assumptions based on structure (“funnel-like” shape) and size (37, 50), in which a very low Reynolds number for lymph flow predicts that valve closure is strictly controlled by pressure and viscous forces. Passive movements of the valve leaflets are consistent with the results of micropipette occlusion and fluid injection protocols (27, 29, 52, 53).

Although there is no reason to question the basic assumptions regarding passive lymphatic valve gating (50), several observations in isolated, pressurized collecting lymphatic vessels from rat mesentery led us to suspect that other factors significantly influence this process. First, the valves in passive, single-valve segments are open when pressures in the two cannulation pipettes are equal. A valve can remain open even during a spontaneous contraction because the intraluminal pressure spike associated with systole is blunted by the low
access resistance created by the cannulation pipettes. Second, for strongly pumping segments with two valves, in which intraluminal pressure in the lymphangion is measured with a servo-null system, the output valve is often observed to open at the peak of systole even when peak systolic pressure does not reach or exceed the pressure in the output pipette (unpublished observations), as would be predicted if only the pressure gradient determined valve opening; moreover, the discrepancy between the two pressures appears to depend on the degree to which the vessel is distended.

For these reasons, we hypothesized that the gating of secondary lymphatic valves is governed primarily by the passive, trans-valve pressure gradient but is also influenced by leaflet stiffness or other mechanical factors related to vessel distention. To test this hypothesis, we devised protocols to measure the small pressure gradients required to open or close lymphatic valves under carefully controlled conditions and to determine whether those pressure gradients vary as a function of vessel diameter.

An additional goal of our study was to determine the valve gating patterns during the active, lymphatic contraction cycle. Surprisingly, these patterns have not been documented in detail previously but are assumed to be similar to those observed for heart valves during the cardiac cycle. However, at least two studies have noted the apparent absence of a period of isovolumic systole in spontaneously contracting lymphatic segments (2, 18), when closure of both valves of a lymphangion should be required for intraluminal pressure development (38). Thus we sought to measure valve gating patterns during active lymphatic contractions in relation to pressure and diameter. We predicted that the patterns would be more complex than those of cardiac valves because 1) lymphatic preload and afterload can vary independently or simultaneously depending on the prevailing local pressure conditions, and 2) there are differences in the flow regimes in lymphatic vessels (low Reynolds numbers and highly viscous flow) vs. the heart (large Reynolds numbers and important inertial effects).

**METHODS**

**Vessel isolation.** All animal protocols were approved by the University of Missouri Animal Care and Use Committee and conformed to the Public Health Service Policy for the Humane Care and Use of Laboratory Animals (PHS Policy, 1996).

Male Sprague-Dawley rats (150–300 g) were anesthetized with sodium pentobarbital (60 mg/kg ip), and a loop of intestine from each animal was exteriorized. Lymphatic vessels (80–180 μm inner diameter) were dissected from mesenteric arcades and placed in physiological saline solution (PSS) with albumin (APSS) at room temperature. The animal was exsanguinated from mesenteric arcades and placed in physiological saline solution (PSS) with albumin (APSS) at room temperature. The animal was exsanguinated from mesenteric arcades and placed in physiological saline solution (PSS) with albumin (APSS) at room temperature.

After cannulation, but before the bath temperature was raised to 37°C, the vessel was passively perfused with PSS; it was then placed in the servo-null system. The output signal was filtered at 50 Hz (model LPBF-10G; NPI Electronic, Tamm, Germany). The controller compared the output signal from the pressure transducers to reference voltages specified by a computer (running LabVIEW; National Instruments, Austin, TX). Video images were captured in AVI files along with embedded pressure data, so that all the measured parameters could be replayed offline (using LabVIEW; National Instruments); this method also permitted diameter tracking to be performed at additional sites along the vessel, if needed. The video was compressed (5:1) without degrading the image quality, using a lossless video codec (Lagarith).

After initial equilibration, pressure control was switched from standing reservoirs to a custom-made, analog servo-control system (Cardiovascular Research Institute, Texas A&M University) in which servo-null style pumps (model V102; Ling Dynamic Systems, Royston, UK) were connected to the pressure transducers and powered by a hardware-based servo-controller through unitary-gain power amplifiers. The controller compared the output signal from the pressure transducers to reference voltages specified by a computer (running LabVIEW) and adjusted the pump voltages accordingly (57). Pressure steps between 0 and 50 cmH2O could be achieved within 50 ms, and combinations of step- or ramp-pressure waveforms could be applied to the input and/or output ends of the isolated lymphatic segment.

**Servo-null pressure recording.** In segments with two valves, the servo-null method was used to directly measure intraluminal pressure (P<sub>i</sub>). Servo-null micropipettes were pulled from borosilicate glass (1.0/0.5 mm outer diameter/inner diameter, Omega-dot; Frederick Haer, Bowdoin, ME) using a Sutter P-97 puller (Sutter Instrument, Novato, CA). The tips were either gently broken or beveled to an outer diameter of ~2 μm. The pipettes were back-filled with 2 M NaCl and connected to a servo-null micropressure system (model 4A; IPM, La Mesa, CA) to monitor intraluminal pressure in the central lymphangion. Positioning was achieved using a hydraulic micromanipulator (model MO-102; Narishige, East Meadow, NY). The servo-null pump incorporated the same type of low-pressure transducer used in the servo-controller. The system was calibrated using a water manometer along with the other pressure transducers before each experiment. The output signal was filtered at 50 Hz (model LPBF-01G; NPI Electronic, Tamm, Germany).

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vessel wall. Successful micropuncture left the pipette tip free from obstruction in the vessel lumen. Servo-null measurements of $P_i$ were made immediately downstream from the input valve (see image montage in Fig. 1A) to protect the central lymphangion from changes in axial length that occurred upon penetration of the micropipette through the vessel wall. The success rate, without subsequent impairment of contractions after warming, was ~80%. The criteria used to assess valid servo-null pressure recordings were the same as described previously (33). The calibration of each micropipette was adjusted after bath temperature reached 37°C and checked periodically during the experiment if pipette plugging was suspected; the test involved simultaneously raising $P_i$ and $P_{out}$ and adjusting the calibration gain and offset as appropriate to match the pipette pressures.

After micropuncture, the vessel was equilibrated for 30–60 min at 37°C and 3 cmH$_2$O until a stable pattern of spontaneous contractions developed. Subsequently, $P_i$ and $P_{out}$ were lowered to a baseline pressure of 1 cmH$_2$O for an additional 5- to 10-min equilibration period before pressure ramps were imposed. All vessels, even those with two valves (consisting of 1 complete lymphangion surrounded by 2 partial lymphangions), showed nearly synchronized contractions of the entire cannulated segment.

Valve timing protocols. The movements of the input and output valve leaflets relative to changes in diameter and intraluminal pressure were determined at various levels of $P_i$ and $P_{out}$. Initially, $P_{out}$ was set to the same level as $P_i$, and several spontaneous contractions were recorded. Subsequently, a slow, ramp-wise elevation in $P_{out}$ was imposed, with the final pressure being just beyond the level at which the vessel failed to eject during systole. The $P_{out}$ level at which this occurred was quite variable (4–20 cmH$_2$O), as detailed in a separate study (unpublished observations). During these protocols, the video, along with associated analog data, was saved periodically into an AVI file at 28–30 Hz. Movements of the lymphatic valves were detected upon replay of the AVI files, in synchrony with the extracted diameter and pressure data, using a custom computer algorithm (in LabVIEW) to measure the change in image intensity in a region of interest overlaying the base and/or midpoint of a valve leaflet.

The most accurate recordings of valve gating were made in vessels in which closed valve leaflets were oriented perpendicular to the image plane; most output valves were intentionally positioned in this way when the vessel was cannulated to optimize valve tracking. Two densitometric windows were positioned along the midline of the vessel, with the mean pixel intensity of one window used to detect the
amount of overlap between the leaflets when the valve was closed or partially closed (Supplemental Fig. S1, window 4; Supplemental Material for this article is available online at the Am J Physiol Heart Circ Physiol website); this signal was highest when the valve was completely open (Supplemental Fig. S1A) and lowest when it was completely closed. However, pixel intensity within the vessel lumen was also influenced by vessel constriction and/or lateral movements associated with constriction. Therefore, a second window was positioned in the vessel midline (Supplemental Fig. S1, window 3) and used to monitor the change in lumen intensity associated with contraction. The difference between the mean pixel intensities of the two windows (after both were normalized to their control values in diastole) yielded a net densitometer signal that more closely reflected only the movements of the valve leaflets. The baseline level of the net densitometer trace also reflected the backward bulging of the output valve as $P_{out}$ was progressively elevated (e.g., Supplemental Fig. S1, $B-E$, red trace). The degree to which the densitometer output accurately reflected the valve positions was confirmed by replaying the video (at $\approx \frac{1}{3}$ real time) and manually toggling a button to denote whether the valve was open or closed in each frame; the resulting manual and automated traces were virtually identical. A thresholding method was then applied to the net densitometer trace to convert it to a digital recording of valve position, where 1 was open and 0 was closed.

**Valve closure and opening tests.** To determine the trans-valve pressure gradient required to open or close a valve, single valves were studied in isolation (see Fig. 4B). Initial valve gating tests were performed in Ca$^{2+}$-free solution to prevent interference from intraluminal pressure spikes associated with spontaneous contractions. Only one densitometer window was needed under these conditions, with the window positioned along the midline of the closed leaflets.

For valve closure tests, pressures were initially set to equal “baseline” values (typically 3 cmH$_2$O), and the leaflet positions were monitored as $P_{out}$ was slowly ramped (4 cmH$_2$O/min) to a higher value, with $P_{in}$ held constant, until the valve closed. The point of valve closure was evident by a rapid decline in the densitometer signal and also by a smaller but simultaneous drop in upstream diameter. This procedure was repeated three times at each baseline pressure while the video was saved to an AVI file. The baseline pressure was then set to another level, either 0.2, 0.5, 1, 2, 3, 5, 7, 11, 15, or 20 cmH$_2$O for 1–2 min, and the ramp test was repeated in triplicate. A complete set of measurements at all of the above pressures was typically obtained.

For valve opening tests, $P_{out}$ was set to a level higher than the baseline $P_{in}$ required to close the valve; $P_{in}$ was then ramped up to a higher level, while $P_{out}$ was held constant, until the valve opened. Valve opening was evident by an abrupt increase in the densitometer signal. This procedure was repeated in triplicate for each of the baseline pressures while the video was recorded.

Because the valve tests under passive conditions might not accurately reflect the behavior of the valves under physiological conditions, the closure test was repeated in the presence of imposed basal tone. A number of different agonists potentially could have been used to impose tone (13, 39), but accurate measurement of the small pressure gradients required to test valve gating also required elimination of spontaneous contractions. After several different methods were attempted for simultaneously imposing tone and inhibiting contractions, the one that worked most consistently was to exchange the bath for KPSS + substance P (SP). Elevated [K$^+$] led to cessation of spontaneous contractions and produced a transient constriction, while the simultaneous addition of SP led to the development of stable tone, at least at low pressures (see Supplemental Fig. S2). The SP dose was adjusted to induce various levels of tone, from $\approx 10\%$ tone at 5 $\times$ 10$^{-8}$ to $\approx 80\%$ tone at 1 $\times$ 10$^{-6}$ M. Tone was calculated as the percent diameter decrease at 3 cmH$_2$O compared with that of the passive vessel.

**Particle tracking.** In a few vessels, we sought to confirm that a valve indeed remained open in the face of an adverse pressure gradient by tracking particles that flowed backwards through the valve. For particle tracking, the cannulation pipettes were filled either with unfiltered APSS containing some particulate matter (2 experiments) or with a dilute suspension of red blood cells (1:8 in APSS; 2 experiments) obtained by cardiac puncture of the same rat. When particles or cells exited the output pipette at defined pressure gradients, sequences of AVI images were captured showing the particles moving retrograde through the valve toward the input pipette (Supplemental Fig. S3). Frame subtraction was used to generate a 32-bit image in Image J (National Institutes of Health; http://rsb.info.nih.gov/ij) to give a middle-gray image with bright spots indicating the position to which the particle had moved and dark spots to indicate the position from which the particle had moved. This subtracted image was then overlaid onto the average of the two images used for the subtraction and the particle positions were pseudocolored.

**Confocal microscopy.** Vessel segments were observed at $\times 40$–50 magnification using a Leica AOB5 SP2 Confocal Multiphoton Microscope System. The cells were loaded with CellTracker Green (5 μM in PSS; Molecular Probes). Images were acquired at 0.3-μm intervals in the z-axis (vertically through the lumen) using 489 nm excitation and 508 nm emission wavelengths. The image stacks were reconstructed using Leica confocal software to produce various three-dimensional projections.

**Data analysis.** Custom analysis programs written in LabVIEW were used to extract embedded data from the AVI files, detect valve leaflet movements, and detect diameter and pressure changes off-line. Data were compiled and plotted in Excel (Microsoft, Redmond, WA) and Igor (WaveMetrics, Lake Oswego, OR) and/or Photoshop (Adobe, Mountain View, CA). Typically, one vessel was used for data analysis from each animal. Curve fitting was performed using Igor. Supplemental video files were made using custom LabView programs to crop the video sequences, overlay the embedded data traces, and add appropriate labels. The files were converted from AVI to MP4 format using Aiseesoft Converter Suite (Beijing, China).

**RESULTS**

**Valve gating during the lymphatic contraction cycle.** To determine the sequence of changes in valve position relative to changes in inner diameter and intraluminal pressure, two-valve lymphatic segments were subjected to different combinations of $P_{in}$ and $P_{out}$. A typical recording is shown in Fig. 1B. With $P_{in}$ and $P_{out}$ initially set to equal levels, $P_{out}$ was slowly elevated from 1 to 8 cmH$_2$O at a standard ramp rate of 4 cmH$_2$O/min. $P_{in}$ was held constant as $P_{out}$ was elevated. Typical diameter and pressure responses to a $P_{out}$ ramp are represented by the two bottom traces. The second trace from the top is the net densitometer signal (red is output valve, axis at left; blue is input valve, axis at right) with the shaded area over the densitometer trace representing the range of that signal corresponding to an open valve. The top traces show the binary valve positions (using the same color scheme). Progressive elevation of $P_{out}$ was associated with a progressive decrease in the open times of both valves and a progressive increase in the closed time of the input valve; in this example, the output valve stopped opening halfway through the ramp, as noted below.

Vessel diameter was measured on both sides of the output valve to illustrate the effect of increasing $P_{out}$ on the diameters of the respective segments. As $P_{out}$ increased, the vessel distended progressively on the output (downstream) side of the valve (red trace), as reflected by increases in both end diastolic diameter and end systolic diameter (ESD). In contrast, end diastolic diameter in the central chamber, on the upstream side...
of the output valve (black trace), decreased slightly as $P_{\text{out}}$ increased, reflecting a modest myogenic constriction. Myogenic constriction was a very typical response during $P_{\text{out}}$ ramp protocols and will be described in more detail a separate study. In Fig. 1B, bottom, pressures in the input pipette (blue trace), output pipette (red trace), and central lymphangion (black trace) are overlaid on a common y-axis scale. As $P_{\text{out}}$ was raised from 1 to 3 cmH$_2$O, $P_L$ transiently increased during each contraction such that peak systolic $P_L$ exceeded $P_{\text{out}}$; when this occurred, the output valve opened, as indicated on the “valve position” trace, and partial emptying of the luminal contents resulted. At $P_{\text{out}}$ levels from 4 – 6 cmH$_2$O (denoted by vertical dotted lines), the output valve opened during each spontaneous contraction and some ejection occurred, even though peak $P_L$ did not clearly exceed $P_{\text{out}}$. This phenomenon is discussed in subsequent figures and was not due to unequal calibrations of the two transducers. At $P_{\text{out}}$ levels $>$ 6 cmH$_2$O, the lymphangion continued to contract spontaneously and generated a rise in $P_L$ sufficient to close the input valve, but the peak systolic pressure was insufficient to open the output valve (as was evident by the closed state of the output valve in the valve position trace). Contraction frequency increased by $\sim$70% during this particular $P_{\text{out}}$ ramp, followed by rate-sensitive inhibition (12) that was evident by a long delay before contractions resumed after termination of the ramp. The video sequence is available as Supplemental Video S1.

Expanded views of the timing of valve leaflet positions relative to diameter and intraluminal pressure are shown in Fig. 2 when $P_{\text{in}} = P_{\text{out}}$ (A) and later in the ramp when $P_{\text{out}} > P_{\text{in}}$ (B). There were significant differences in the patterns of valve gating under these two conditions. In Fig. 2A, both valves were open before the first period of systole began (systole and diastole are denoted by vertical dotted lines). When systole was initiated, the fall in diameter and rise in $P_L$ essentially coincided. Closure of the input valve coincided with the period in which $P_L$ exceeded $P_{\text{in}}$. The input valve then returned to the open position at the end of systole. In early diastole, when the rate of the diameter change was relatively high, $P_L$ dipped transiently below $P_{\text{in}}$ (and $P_{\text{out}}$). This negative pressure, relative to $P_{\text{in}}$, observed previously in isolated bovine mesenteric lymphangions under similar conditions (24) and that we hereafter call the “suction effect,” pulled the output valve leaflets backward into the closed position, as indicated in the densitometer trace and the valve position trace. This phenomenon was observed in $\sim$75% of vessels when $P_{\text{in}}$ and $P_{\text{out}}$ were equal and at a low level ($\leq$3 cmH$_2$O); diameter typically decreased by $\geq$50% in systole under these conditions, as evident in this example (see also the examples in Supplemental Figs. S4 and S5). The suction effect was diminished somewhat during the second contraction, was only present in four of the seven contractions before the ramp (missing in contractions marked by asterisk in Fig. 1B), and was essentially absent in Fig. 2B when the systolic periods were characterized by smaller amplitude changes.

When $P_{\text{out}}$ was higher than $P_{\text{in}}$, the valve gating pattern was much more consistent with the pattern observed for ventricular valves during the cardiac cycle (56), as illustrated in Fig. 2B. The output valve was closed throughout diastole. At the initiation of systole, the input valve closed, permitting a greater rise in $P_L$ than could be recorded when $P_{\text{in}}$ and $P_{\text{out}}$ were equal (due to partial shunting of the $P_L$ spike through an open output valve into the output cannula). When $P_L$ exceeded $P_{\text{out}}$, the output valve opened. Pressure development in the central lymphangion during the time that both valves were closed implies that a brief period of isovolumic systole occurred (Fig. 2B, inset with expanded time scale at right). During this time, diameter was decreasing at the site immediately upstream from the output valve but the effect on volume was apparently balanced by an increase in diameter at the more proximal end (near the sinus) of the central lymphangion (data not shown); indeed, diameter increases near the sinus were typical during systole when $P_{\text{out}}$ was elevated (data not shown). In midsystole, peak

Fig. 2. Expanded views of selected contractions from the recording in Fig. 1 show the timing of the input and output valves at different levels of $P_{\text{out}}$. Traces are the same as described in Fig. 1. A: lymphatic contraction cycle when $P_{\text{in}} = P_{\text{out}}$ (1 cmH$_2$O). Vertical lines denote the periods of systole, as determined from the diameter recordings. B: lymphatic contraction cycle when $P_{\text{out}} > P_{\text{in}}$. Vertical lines denote the periods of systole, as determined from the diameter recordings. B, inset: first contraction with an expanded time scale; here the dotted vertical lines denote a period of isovolumic systole. ESD, end systolic diameter; EDD, end diastolic diameter.

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systolic $P_L$ clearly exceeded $P_{out}$, but this was not the case at higher levels of $P_{out}$ (see Fig. 3). Systolic $P_L$ peaked before the point in time when ESD was reached. At ESD, $P_L$ was essentially equal to $P_{out}$, at which point the output valve began to close. The input valve remained open until $P_L$ fell below $P_{in}$. Note the slightly longer open time for the input valve and its correspondence with the $P_L$ trace in the second contraction of Fig. 2B, presumably because $P_{out}$ and thus peak $P_L$ were higher so that the fall to $P_{in}$ required more time.

**Discrepancy between the level of peak systolic $P_L$ and $P_{out}$ during ejection.** A common observation during $P_{out}$ ramps is illustrated in Fig. 3. Peak systolic $P_L$ clearly did not exceed or even match $P_{out}$ after $P_{out}$ was greater than $-3$ cmH$_2$O; nonetheless, the output valve opened transiently during each contraction (red valve position trace), as indicated by the horizontal dotted line. At first we suspected transducer drift or partial plugging of the servo-null pipette, which would have altered the $P_L$ calibration. However, a quick calibration test following the $P_{out}$ ramp (at time $=148.1$ min) verified that neither of these problems occurred. It is still possible that the frequency response of the servo-null pipette was not sufficient to accurately detect the $P_L$ peak; however, this behavior was observed in a number of vessels with varying degrees of discrepancy between peak systolic $P_L$ and $P_{out}$. This difference in the two pressures in the face of continued ejection, along with the “bias” of the valves to be open in the absence of a pressure gradient, led us to more carefully examine the pressure gradients required to close or open a valve at different levels of vessel distention.

**Valve closure test using a $P_{out}$ ramp.** A three-dimensional reconstruction of a valve in a rat mesenteric lymphatic vessel, obtained using confocal microscopy, is shown in Fig. 4A. The view is taken from downstream, looking back against the normal flow direction into the valve. Both the valve leaflets and their downstream insertion points are visible, and the bicuspid appearance of the valve is evident. A video that rotates through a 360° view is available as Supplemental Video S2. Based on areas calculated from this image, the ratio of the valve opening to the lumen is 1:7.

A typical valve used for a closure test is shown under brightfield illumination in Fig. 4B, with the valve viewed from the side so that the insertion points lie at the top and bottom surfaces of the vessel. Figure 4, C and D, shows the data obtained during the closure test. Initially, pressures in the cannulation pipettes were equal (in Fig. 4, C and D, middle traces) and the valves were open (densitometer signals were high). As $P_{out}$ slowly increased, the valve closed (denoted by a vertical dotted line). At this time, diameter decreased because pressure on the upstream side of the valve dropped to $P_{in}$ (from approximately half-way between $P_{out}$ and $P_{in}$) at the moment of closure. The magnitude of the diameter change depended on the degree to which the vessel was distended initially.

At the low baseline pressure used for the test in Fig. 4C, the minimal pressure gradient ($P_{out}$ − $P_{in}$) required to close the valve was 0.17 cmH$_2$O. In contrast, when the closure test was repeated on the same valve, at a higher baseline pressure (Fig. 4D), a minimal pressure gradient of 1.27 cmH$_2$O was required for closure. The set of three closure tests for this vessel at the lower pressure level are available as Supplemental Video S3.

Particle tracking during the imposition of an adverse pressure gradient confirmed that valves with the morphology shown in Fig. 4B, top, were indeed open. Supplemental Fig. S3A shows a vessel segment with an apparently open valve when $P_{in} = P_{out} = 5$ cmH$_2$O. As $P_{out}$ was elevated ramp wise from 5 to $-6.7$ cmH$_2$O, with $P_{in}$ held constant, a single particle emerged from the “output” pipette (point #1) and moved backwards through the valve, down the prevailing pressure gradient, until it exited the “input” pipette at point #17. The positions of the particle at sequential times in the AVI file are shown by the yellow dots (1–17). Supplemental Fig. S3B shows particle velocity as a function of the relative position, and Supplemental Fig. 3C shows the diameter, pressure and densitometer data during the $P_{out}$ ramps associated with this test. The times at which the movements of this particle were...
recorded correspond to AVI frame numbers (1,156–1,295, with total duration = 4.8 s) in the second ramp; the pressure gradient was ~1 cmH2O when this particle moved across the field of view. In other experiments (see Supplemental Video S4), red blood cells introduced into the pipette system were observed to move in the same direction through an open valve and to stop abruptly the instant the valve closed.

Valve opening test using a Pout ramp. An example of the test devised to measure the minimal pressure gradient required to open a valve is shown in Fig. 5. With Pout held constant, Pin was raised ramp wise until the valve opened. Pin and Pout levels at the instant of closure are recorded and the difference (Pout − Pin) is the minimal pressure gradient required for closure. Diameter traces on both sides of the valve are recorded at sites corresponding to the colored windows in B. Test in C starts at the lowest baseline pressure level used (0.2 cmH2O), whereas the test in D starts at the highest level of baseline pressure used (20 cmH2O). Both recordings are from the same vessel. Note the difference in the diameter and pressure scales between C and D.

Summary analyses for valve gating tests. The data sets from the valve closure and opening tests, performed at 10 different baseline pressures, were analyzed in two different ways. In Fig. 6A, the pressure gradient required for valve closure (filled circles, axis at left) is plotted as a function of baseline pressure. The reference pressure was designated as the side on which the pressure ramp was imposed (output); thus P was calculated as Pout − Pin for the valve closure test. Over the lower end of the baseline pressure range from 0.1 to 3 cmH2O, the pressure gradient required for valve closure increased from 0.1 to 1.4 cmH2O. However, as baseline pressure increased from 3 to 20 cmH2O, the pressure gradient required for valve closure increased only an additional 30% (1.4 to 2.1 cmH2O). Interestingly, the change in slope of the curve fitting these data roughly corresponded to the “elbow” in the passive pressure-diameter relationship for the same vessel (red), which is plotted on the axis at right in Fig. 6A. The data from the valve opening tests on the same vessel are also plotted in Fig. 6A (open circles). The reference pressure was designated as the side...
on which the pressure ramp was imposed (input); thus ΔP was calculated as \( P_{in} - P_{out} \) for the valve opening test.

Although the general shapes of ΔP-baseline pressure curves were fairly representative of those for most vessels, plotting data sets from multiple vessels in this manner resulted in substantial scatter in the data along the baseline pressure axis. Because baseline pressure determined the passive vessel diameter, the data were expressed as a function of vessel diameter. In Fig. 6B, the pressure gradient required for valve closure (closed circles) and valve opening (open circles) for the same vessel are plotted as a function of the normalized passive diameter (D/D\(_{max}\)), where D\(_{max}\) is the maximum passive diameter in Ca\(^{2+}\)-free PSS at 20 cmH\(_2\)O. Here the ΔP for each test is defined as in Fig. 6A. It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O) as the diameter approached the maximal passive required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O). It is evident that the pressure gradient required for valve closure increased 23-fold (0.1 to 2.3 cmH\(_2\)O).

The results of valve closure tests performed on 17 vessels and valve opening tests performed on 10 vessels are summarized in Fig. 7. All tests were performed in Ca\(^{2+}\)-free solution with the x-axis representing normalized diameter. Higher baseline pressures are associated with progressive distention of the vessel up to the maximum diameter recorded at 20 cmH\(_2\)O, and it is apparent that a valve in a highly distended vessel can remain open in the face of a substantial adverse pressure gradient (on average 2.2 cmH\(_2\)O, but as high as 5 cmH\(_2\)O; Fig. 7A). Likewise, the opening tests in Fig. 7B show that valve opening occurs at a progressively more negative pressure gradient (\( P_{in} - P_{out} \)) as a vessel distends. However, the range of the pressure gradient required for valve opening changes only approximately sixfold with vessel diameter compared with the >20-fold range of the pressure gradient required for valve closing (only one vessel required a pressure gradient more negative than −1 cmH\(_2\)O for valve opening). The data sets in Fig. 7 were fit with the power functions listed in Supplemental Table S1. We impute no physiological or mathematical significance to the power functions; they merely provided better fits of the data sets than exponential or third order polynomial functions.

**Effect of vessel tone on valve closure.** We were concerned that valve gating tests under passive conditions might not accurately reflect the behavior of the valves under physiological conditions; e.g., that the relationships in Fig. 7 might be altered by the presence of spontaneous vessel tone. An effect of tone on gating would be predicted if smooth muscle cells directly or indirectly controlled tension of the valve leaflets. The graph in Fig. 8A shows the results of valve closure tests in four vessels using KPSS + 1 × 10\(^{-7}\) M SP to impose tone (open circles), along with tests performed subsequently on the same valves in Ca\(^{2+}\)-free solution (closed circles). The tone induced by this dose of SP (20.4 ± 2.2%) slightly exceeded the upper limit of basal tone observed under several physiological conditions (11, 13, 22, 23). Tone was relatively stable at low pressures but did not completely recover after tests at baseline pressures >10 cmH\(_2\)O. Thus, at some pressures, tone changed during the set of three closure tests at each baseline pressure. For this reason, all of the data points (rather than only the average of 3 trials at each baseline pressure) were plotted in Fig. 8. There appeared to be slight difference in the curves fitting these data sets, with the curve for the SP data shifted slightly to the left of the Ca\(^{2+}\)-free curve. Curve fits of the individual (paired) data sets (not shown) confirmed that the SP

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**Fig. 5.** Valve opening tests performed on a single-valve vessel in Ca\(^{2+}\)-free solution. A: valve opening test at a low baseline pressure. Densitometer, pressure, and diameter traces are similar to those described in previous figures. Blue traces are the respective recordings on the input side of the valve, and red traces are the respective recordings on the output side of the valve. Test begins with \( P_{in} \) sufficiently higher than \( P_{out} \) to maintain a closed valve. A ramp-wise increase in \( P_{in} \) is imposed until the valve opens. \( P_{out} \) and \( P_{in} \) levels at that instant are recorded and the difference (\( P_{in} - P_{out} \)) is the minimal pressure gradient required for opening. This value was typically negative as shown here. B: valve opening test is repeated at the highest level of \( P_{out} \) used (20 cmH\(_2\)O). Both A and B are from the same vessel. Note the difference in the diameter and pressure scales between the 2 panels.
and Ca\textsuperscript{2+}-free curves essentially overlapped in three of the four vessels.

The valve closure tests were repeated on four additional vessels in which the SP dose was adjusted (from 6 × 10\textsuperscript{-7} M to 1 × 10\textsuperscript{-6} M) to impose ~50\% tone (51.3 ± 1.6\%). In this case, the curve that fit the aggregate closure test data for SP was more noticeably shifted to the left compared with the curve for the aggregate data in Ca\textsuperscript{2+}-free solution (Fig. 8B). Curve fits of the individual (paired) data sets (not shown) confirmed that three of four vessels showed substantial leftward shifts in the curves fitting the SP data relative to the curves fitting the Ca\textsuperscript{2+}-free data. The data sets in Fig. 8 were fit to power functions, with coefficients listed in Supplemental Table S1. A leftward shift in the ΔP vs. D/D\textsubscript{max} relationship suggests that the induction of tone increases the pressure gradient necessary to close the valve at any given baseline pressure, compared with a passive vessel.

**DISCUSSION**

**Valve gating patterns during the contraction cycle.** This study is, to our knowledge, the first detailed description of the timing of lymphatic valve movements relative to pressure and diameter changes during the lymphatic contraction cycle. Thus Fig. 2 is the equivalent of the “Wigger’s diagram” (32) for the lymphangion. In the rat, the timing of lymphatic valve leaflet movements during the contraction cycle closely resembles that of cardiac valves if, and only if, P\textsubscript{out} > P\textsubscript{in} (Fig. 2B). At the onset of systole, the input valve closes first, allowing pressure in the chamber to rise during contraction, after which the output valve opens when P\textsubscript{L} exceeds P\textsubscript{out} (at low pressures). At higher pressures, the output valve will typically open even though P\textsubscript{L} does not reach P\textsubscript{out}, according the relationship defined in Fig. 7B. In either case, once open the output valve closes at the end of systole when P\textsubscript{L} falls sufficiently below P\textsubscript{out}. Finally, the input valve reopens after diastolic P\textsubscript{L} has returned to the baseline level. If diastolic P\textsubscript{L} does not return to baseline before the next contraction is initiated, the input valve remains closed and a slow rise in diastolic P\textsubscript{L} ensues, as can be observed just before the onset of valve insufficiency (unpublished observations).

**Fig. 6.** Summary of valve closure and opening tests for one lymphatic vessel. A, left axis: pressure gradient required for valve closure (P\textsubscript{out} – P\textsubscript{in}; ●) or opening (P\textsubscript{in} – P\textsubscript{out}; ○) plotted as a function of baseline pressure. Each point is the average ΔP (P\textsubscript{out} – P\textsubscript{in}) for 3 trials. Right axis: continuous pressure diameter relationship for the same vessel determined from a simultaneous P\textsubscript{in} + P\textsubscript{out} ramp from 0.2 to 20 cmH\textsubscript{2}O in Ca\textsuperscript{2+}-free PSS after valve gating tests were completed; data were normalized to the passive diameter at 20 cmH\textsubscript{2}O. B: pressure gradient required for valve closure (P\textsubscript{out} – P\textsubscript{in}; ●) or opening (P\textsubscript{in} – P\textsubscript{out}; ○) plotted as a function of normalized diameter. All measurements were made in Ca\textsuperscript{2+}-free PSS. All curve fits are power functions. D/D\textsubscript{max}, passive diameter/maximum passive diameter.

**Fig. 7.** Summary of measurements describing the pressure gradient required for valve closure (A; n = 17) or valve opening (B; n = 10) as a function of normalized diameter.
Flow measurements are usually included in cardiac cycle diagrams (32). Unfortunately, we were not able to measure lymphangion flow accurately under conditions in which diameter, pressure, and valve movements were simultaneously measured. Although centerline velocity can be measured using red blood cells introduced into the input pipette (see Supplemental Video S4), velocity during much of systole is often too high (15, 16) to be accurately determined at ~30 frames per second with the camera used in the present experiments. Flow measurements in conjunction with valve gating tests could increase our understanding of some aspects of valve behavior but would not likely result in highly detailed flow information such as the presence of secondary flow structures near the valve leaflets. This kind of information is currently better obtained from carefully constructed computational flow simulations, such as we have performed for the straighter regions of lymphangions (49). The outcomes of those simulations may then be used to attempt to design better experimental approaches to measure flow patterns around the valves.

Unlike the heart, where \( P_{\text{out}} \) always exceeds \( P_{\text{in}} \), lymphatic valves can exhibit multiple gating patterns depending on the prevailing \( P_{\text{in}} \) and \( P_{\text{out}} \) (see Figs. 1–2). When \( P_{\text{in}} = P_{\text{out}} \), the input valve typically closes at the onset of systole and opens at the end of systole. Because the output valve is open when the trans-valve pressure gradient is zero (Fig. 7), it can occasionally remain open throughout the entire contraction cycle (e.g., contractions marked by asterisk in Fig. 1). More typically, the output valve closes transiently in early diastole as \( P_{\text{in}} \) dips below the level of \( P_{\text{in}} \) and \( P_{\text{out}} \) (Fig. 2A). The dip in intraluminal pressure is associated with contractions of large amplitude that typically occur at low baseline pressure levels (<3 cmH2O) and is the subject of another study (14). In the heart, isovolumic systole is a brief period of time when both input and output valves of the ventricle are closed, allowing pressure development during ventricular contraction. This phase may or may not occur in a lymphatic vessel. Benoit et al. (2) noted, from in vivo observations of lymphatics in the rat mesentery, that there was no discernable period of isovolumic systole. The vessels in their experiments, which were performed in supine animals without an externally imposed afterload, most likely exhibited valve gating patterns resembling those shown in Fig. 2A rather than Fig. 2B. Experiments with single, isolated bovine mesenteric lymphangions studied at equal diastolic \( P_{\text{in}} \) and \( P_{\text{out}} \) also demonstrated the absence of a period of isovolumic systole (20). In contrast, our recordings (Fig. 2B, inset) clearly show a brief period of isovolumic systole when \( P_{\text{out}} > P_{\text{in}} \). Differences in the level of afterload possibly could reconcile the apparent discrepancy between the observations of Benoit et al. (2) and Gashev (20) with those of McHale and Roddie (38, 39) with respect to resolving a period of isovolumic systole in the lymphangion contraction cycle.

Still other valve gating patterns can occur in lymphatic vessels under conditions where \( P_{\text{in}} \) and \( P_{\text{out}} \) change, conditions never experienced by the heart. For example, in edematous tissues, lymphatic \( P_{\text{in}} \) may exceed \( P_{\text{out}} \) and the lymphangion may behave more as a conduit, with continuous forward flow, than as a pump (23, 48); similar behavior may occur normally in intestinal lymphatics during high levels of lymph formation. We are able to simulate this condition by setting \( P_{\text{in}} \) higher than \( P_{\text{out}} \) and thereby imposing a favorable gradient for flow (23, 25). When \( P_{\text{in}} \) is increased ramp wise while \( P_{\text{out}} \) is held constant, both valves typically remain open throughout the ramp (see Supplemental Fig. S4). In contrast, when \( P_{\text{in}} \) and \( P_{\text{out}} \) are simultaneously elevated, both valves will gate sequentially during each contraction cycle until \( P_{\text{in}} \) and \( P_{\text{out}} \) exceed a certain level (Supplemental Fig. S5). The simultaneous \( P_{\text{in}} + P_{\text{out}} \) ramp might simulate a condition where a lymphatic vessel would be pumping against a gravitational load in the face of developing edema.

Effect of vessel distention on valve gating. Our measurements of valve gating in single-valve segments verify previous predictions that the gating of lymphatic valves is primarily (or solely) determined by the trans-valve pressure gradient. Predictions about passive valve gating have been made largely on the basis of valve ultrastructure, which show that the leaflets are composed of two thin endothelial cell sheets containing...
elastin and collagen fibers (26). Schmid-Schönbein (50) noted that the small size of the vessel and low Reynolds numbers for lymph flow support the concept that valve leaflet movements are dictated solely by trans-valve pressure gradients and viscous forces. Although such gradients are predicted to be small, we could find no published measurements to validate this assumption, apart from the work of Lobov (35) in an unpublished Russian thesis. Lobov reported that an adverse pressure gradient of \( \approx 0.4 \) cmH\(_2\)O was sufficient to close secondary valves in bovine mesenteric lymphatic vessels at a baseline pressure of 3 cmH\(_2\)O; although a direct comparison with our results is not possible without knowing \( D/D_{\text{max}} \) for bovine vessels in those particular experiments, the absolute value is well within the range of the pressure gradients for closure of rat mesenteric lymphatic valves shown in Fig. 7A.

A novel and important finding from our study is that the trans-valve pressure gradient required for valve closure (and to a lesser extent for valve opening) depends strongly on vessel distension, the level of which is determined by the baseline pressure at which the test is performed. At low pressures (<2 cmH\(_2\)O) that are within the physiological range of rat mesenteric vessels (2), \( \approx 0.1 \) cmH\(_2\)O pressure gradient (\( P_{\text{out}} - P_{\text{in}} \)) is required for valve closure, whereas at higher pressures (e.g., >10 cmH\(_2\)O), a trans-valve gradient of 2.2 cmH\(_2\)O (and in some cases >5 cmH\(_2\)O; Fig. 7A) is required. Thus there is a >20-fold range in the pressure gradient determining valve closure over the range of pressures the vessel would experience normally and under conditions of edema (44, 46). The baseline pressure at which the \( \Delta P \) for closure substantially increases (typically 2–3 cmH\(_2\)O) corresponds approximately to the “elbow” in the passive pressure-diameter curve of the vessel (Fig. 6A) and suggests that the increase in the pressure gradients required for opening/closing may be determined by tensioning of the valve leaflets as the vessel progressively distends. The \( \Delta P \)-diameter relationship (Fig. 7A) explains why valves do not always close when a vessel segment with a single-valve contracts under conditions where \( P_{\text{in}} = P_{\text{out}} \) (as occurs in Fig. 1 before the \( P_{\text{out}} \) ramp begins). At baseline pressures >2 cmH\(_2\)O, the small fluctuations in intraluminal pressure associated with contraction are insufficient to generate even a transient back-pressure large enough to close the valve, since the pressure spikes are largely shunted out the open cannulation pipettes. The graph in Fig. 7A also explains why reverse flow (through an open valve) can sometimes be observed in vivo (Supplemental Video S7). The graph in Fig. 7B explains why an output valve can open even if internal (developed) pressure does not exceed output pressure (Fig. 3).

There was a distinct lack of symmetry in the ranges of pressure gradients necessary to open (6-fold) and close (20-fold) the valve leaflets as transmural pressure increased (Fig. 6). This is perhaps related to the bias of the valves to remain in the open position. Furthermore, increasing the transmural pressure likely results in less contact area between the leaflets and makes coaptation more difficult. This would further bias the valve to the open position. We occasionally noted that there appeared to be less contact area between the leaflets at larger vessel diameters, but the imaging techniques used here did not allow for accurate assessment of contact area. It is also possible that, like the vessels themselves (Fig. 6A), the valve leaflets become stiffer with increasing diameter as the wall distends and pulls radially on the base and/or buttress of the leaflets.

While direct measurements of stiffness in this setting would be extremely difficult to make, we can qualitatively visualize leaflet behavior under various flow conditions. In some experiments, the leaflets exhibit measureable “flutter” in response to rapid pressure oscillation at both low and high baseline transmural pressures (Supplemental Video S6), suggesting that they remain reasonably compliant even when a vessel is near maximally distended. A fuller assessment of this phenomenon through high resolution three-dimensional imaging and computational modeling would be beneficial. In the meantime, the findings of our study should provide valuable information for models of lymphatic function such as the recent model developed by Bertram et al. (3).

**Influence of vessel tone on valve gating.** The idea that active force generation by muscle cells in the lymphatic wall can control valve gating is controversial. In bovine lymphatics, some anatomical evidence exists for the presence of muscle cells or muscle cell extensions into the valve leaflets (4, 5), yet no functional data are available to support the concept that muscle contraction per se can initiate valve closure/opening. In this regard, we never observed valves to close without the imposition of an appropriate trans-valve pressure gradient, even in the presence of tone. However, the relationships described in Fig. 8 and Supplemental Fig. S6 clearly demonstrate that tone has an influence, albeit an indirect one, on valve closure. The reciprocal implication is that high levels of tone will lead to enhanced valve opening when \( P_{\text{in}} < P_{\text{out}} \) (Fig. 7B), i.e., in the presence of high tone an output valve will open well before \( P_{\text{L}} \) reaches \( P_{\text{out}} \), meaning that active force developed by lymphatic muscle during systole would facilitate valve opening. It should be noted that “tone” in this context reflects the effect of steady-state stimulation of the vessel with KPSS + SP in our protocols. Whether the same principle also applies to the transient, active force development by lymphatic muscle during the contraction cycle will be difficult to determine using methods similar to the ones we employed. Nevertheless, our results (Fig. 8) are partially consistent with the idea of an active
muscle component previously proposed to be involved in the gating of bovine lymphatic valves (4, 5, 42), although we anticipate that this is an indirect effect transmitted possibly through collagen fibers at the base or buttress of the valve leaflets rather than by muscle cells within the leaflets.

Physiological implications. The lymphatic system typically works against an opposing standing pressure gradient to move lymph from a low pressure environment (the interstitium) to a higher pressure environment (outflow into the venous network). An important conclusion from the present study is that, despite the pressure gradient against which the lymphatic network works, secondary lymphatic valves in rat mesenteric lymphangions nevertheless have a “bias” to be in the open position (Figs. 1, 2, 6, and 7). However, the extent of this “open bias” is strongly influenced by the hydrodynamic conditions that the valve experiences on a moment-to-moment basis. When lymph pressures are low and there is little or no adverse pressure gradient (e.g., $P_{in} = P_{out} = 1$ cmH$_2$O), the valves are open the majority of the contractile cycle, ~85 and 75% of the duty cycle for the input and output valves, respectively (Figs. 1 and 2A).

However, when the vessel must work against a pressure gradient, the amount of time the valve spends in the open position significantly declines, to ~75% and 15% of the duty cycle for the input and output valves, respectively, when $P_{out} = ~2$ cmH$_2$O and $P_{in} = 1$ cmH$_2$O (Figs. 1 and 2B). When the opposing pressure gradient exceeds the limit of the pump, the output valve does not open and the duty cycle of the input valve falls substantially (to ~30% in the example shown in Fig. 1).

The open bias of the valve potentially could translate into several physiological advantages to enhance the efficiency of lymphatic function, including the predominant role of the valve to minimize backflow of lymph. We have also demonstrated that valve gating is different at low vs. high pressures (Figs. 6 and 7). When pressure is low, the vessel has a relatively small diameter and the trans-valve pressure difference needed to close the valve is relatively small. However, when the pressure is high, the vessel is distended and the $\Delta P$ required for closure significantly increases. Additionally, we have shown that this relationship is altered by lymphatic tone (Fig. 8). Thus a potential benefit of the residual lymphatic muscle tone between contractions could be to help maintain the vessel at a relatively small diameter (compared to the passive diameter at the same pressure), thereby allowing the valve to close properly during diastole under a small adverse pressure gradient. This is illustrated clearly in Fig. 9, which depicts the $\Delta P$ required to close a valve at different baseline pressures in a single vessel with high tone (KPSS + $10^{-6}$M SP) and in the same vessel with no tone (Ca$^{2+}$ free PSS). With loss of tone (vertical arrow), a larger $\Delta P$ is required for valve closure. For example, at a baseline pressure of 2 cmH$_2$O, the $\Delta P$ required to close the output valve in the vessel with high tone (~102 $\mu$m average diameter) is ~0.3 cmH$_2$O, whereas in the same vessel with no tone (~136 $\mu$m average diameter), the $\Delta P$ required to close the output valve is approximately threefold higher. Figure 9 illustrates this relationship for a single vessel where the effect is easier to see, but the entire data set shows a similar relationship (see Supplemental Fig. S6), both for moderate and high levels of tone.

An extension of the idea described in the preceding paragraph is that valves will become more susceptible to becoming insufficient in distended vessels, because a greater $\Delta P$ is required for closure. Thus, if diastolic pressure rises, pressure will be more likely to equalize through an open valve, rendering that valve insufficient. Indeed, we have observed this sequence of events frequently under conditions that mimic those seen in lymphedema (a simultaneous ramp increase in $P_{in}$ and $P_{out}$, with $P_{out} > P_{in}$; unpublished observations) and insufficient valves have been reported to be associated with distended lymphatic vessels in vivo, including vessels in patients with lymphedema (44, 51). A potential means to combat this problem and minimize lymph reflux could be to increase the resting tone of the collecting lymphatic vessels.

In summary, our findings provide several new insights into the function of secondary lymphatic valves. 1) We have documented in detail for the first time the temporal sequence of events among diameter, pressure, and valve gating during the lymphatic contraction cycle. The timing of the lymphatic valve movements is similar to that of cardiac valves only under the specific condition where lymphatic afterload is elevated. 2) We find that lymphatic vessel distention exerts a profound influence on the pressure gradient required for valve opening/closing. 3) We provide an explanation for why lymphatic valves in mesenteric vessels have a natural tendency to remain open when the trans-valve pressure gradient is zero or near zero. 4) Finally, we present functional evidence to support the concept that lymphatic muscle tone exerts at least an indirect effect on valve gating.

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DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the author(s).

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