

# Inflammation and metabolic dysfunction: links to cardiovascular diseases

Annika Taube,\* Raphaela Schlich,\* Henrike Sell, Kristin Eckardt, and Juergen Eckel

Paul Langerhans Group, German Diabetes Center, Duesseldorf, Germany

Submitted 14 September 2011; accepted in final form 14 March 2012

**Taube A, Schlich R, Sell H, Eckardt K, Eckel J.** Inflammation and metabolic dysfunction: links to cardiovascular diseases. *Am J Physiol Heart Circ Physiol* 302: H2148–H2165, 2012. First published March 23, 2012; doi:10.1152/ajpheart.00907.2011.— Abdominal obesity is a major risk factor for cardiovascular disease, and recent studies highlight a key role of adipose tissue dysfunction, inflammation, and aberrant adipokine release in this process. An increased demand for lipid storage results in both hyperplasia and hypertrophy, finally leading to chronic inflammation, hypoxia, and a phenotypic change of the cellular components of adipose tissue, collectively leading to a substantially altered secretory output of adipose tissue. In this review we have assessed the adipo-vascular axis, and an overview of adipokines associated with cardiovascular disease is provided. This resulted in a first list of more than 30 adipokines. A deeper analysis only considered adipokines that have been reported to impact on inflammation and NF- $\kappa$ B activation in the vasculature. Out of these, the most prominent link to cardiovascular disease was found for leptin, TNF- $\alpha$ , adipocyte fatty acid-binding protein, interleukins, and several novel adipokines such as lipocalin-2 and pigment epithelium-derived factor. Future work will need to address the potential role of these molecules as biomarkers and/or drug targets.

adipokines; adipose tissue

THIS ARTICLE is part of a collection on **Cardiovascular Response to Obesity and Diabetes**. Other articles appearing in this collection, as well as a full archive of all collections, can be found online at <http://ajpheart.physiology.org/>.

## Introduction

Obesity is a metabolic disorder of pandemic proportions and is associated with a variety of metabolic dysfunctions like hypertension, dyslipidemia, insulin resistance, and hyperglycemia (199). Thus it is considered a major risk factor for the development of chronic diseases such as type 2 diabetes (145) and cardiovascular diseases (109, 219). Adipose tissue enlargement ensues as a consequence of a persistent positive energy balance resulting from a sedentary lifestyle, conditioned by environmental and genetic factors. Formerly, the function of adipose tissue was thought to be restricted to insulate and cushion the body, store triglycerides during periods of excess energy and provide the body with energy in the form of free fatty acids in states of energy shortage (89). However, it has become increasingly evident that adipose tissue is also a secretory organ able to release various lipid mediators as well as a multitude of bioactive proteins and peptides, collectively referred to as adipokines (275).

Today, it is commonly accepted that adipokines have essential roles in energy homeostasis, glucose and lipid metabolism, cell viability, control of feeding, thermogenesis, neuroendocrine function, reproduction, immunity, and, importantly, car-

diovascular function (88). Accordingly, numerous studies in recent years have demonstrated the pivotal role of adipokines as molecular messengers in the cross talk of adipose tissue with other organs and tissues as well as their contribution to the development of obesity-associated disorders. However, this picture has gained complexity since modern approaches applying highly sensitive analytical techniques have revealed that the adipose tissue output is comprised of hundreds of different factors (4, 130, 226, 334), with additional novel adipokines still being identified (143). Furthermore, it has been described that the adipokine profile may vary between different adipose tissue depots and is altered in pathological conditions such as obesity. In this context, it has been shown that plasma levels of several proinflammatory cytokines as well as acute phase proteins such as C-reactive protein (CRP) are increased in obesity, contributing to a chronic state of low-grade inflammation (67, 326, 327). This systemic inflammatory state has been suggested to be a causative link between obesity and related secondary complications such as cardiovascular diseases (306), by inducing inflammatory processes in the vessel wall. Such processes are considered to be critical determinants of pathological alterations of the vasculature such as thickening of vessel wall, fatty streak formation, or promotion of atherosclerotic plaques.

Previous studies have demonstrated local production of proinflammatory mediators by immune cells of atherosclerotic plaques [for reviews, see Lippy et al. (154) and Zakynthinos and Pappa (329)]; however, in this review we will elucidate the role of adipose tissue-derived factors in the induction of inflammatory processes in the vasculature and focus especially on selected adipokines able to activate nuclear transcription factor- $\kappa$ B (NF- $\kappa$ B) signaling. Furthermore, we will discuss the special role of perivascular fat as a local adipose tissue depot

\* A. Taube and R. Schlich contributed equally to this work

Address for reprint requests and other correspondence: J. Eckel, German Diabetes Ctr., Auf'm Hennekamp 65, D-40225 Duesseldorf, Germany (e-mail: eckel@uni-duesseldorf.de).

and its contribution to the development of cardiovascular diseases. In this context, we propose a list of candidates, including well-known as well as novel adipokines, involved in the induction of inflammatory processes and possibly leading to atherosclerotic lesions as well as cardiovascular complications. With this, we provide new insights into the role of adipokines in the complex interorgan cross talk between adipose tissue and the vasculature. Understanding the molecular mechanisms linking inflammation, metabolic syndrome, and cardiovascular diseases is essential to identify possible biomarkers and potential drug targets. These are important steps to improve diagnosis and treatment of cardiovascular diseases.

#### *Obesity-Associated Alterations of Adipose Tissue*

Enlargement of adipose tissue ensues as a result of an imbalance between energy intake and expenditure. As a consequence, adipocytes undergo hyperplasia and hypertrophy to meet the increased demand for storage capacities (75, 102). However, a persistent state of energy excess represents an increased burden for the lipid storage and processing capacities of the expanding adipose tissue, resulting in various dysfunctions within the tissue like low-grade chronic inflammation and hypoxia (123, 306). These obesity-associated dysfunctions may lead to changes in the cellular composition of the tissue, including alterations in the number, phenotype, and localization of immune, vascular, and structural cells (199), resulting in an altered adipose tissue secretory output. Increased expression of chemoattractant proteins like monocyte chemoattractant protein-1 (MCP-1) induces recruitment and infiltration of additional macrophages. These contribute to the increased expression of proinflammatory cytokines such as tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) (104), thereby further exacerbating the obesity-associated inflamed status of the adipose tissue (301, 306, 313). Additionally, obesity-associated adipocyte hypertrophy has also been associated with a shift of the adipocyte secretome to a more proinflammatory composition (253). In this context, a positive correlation has been described between adipocyte size and secretion of various proinflammatory factors such as TNF- $\alpha$ , IL-6, IL-8, MCP-1, leptin, and granulocyte colony-stimulating factor (253). Whereas the majority of adipokines have been found to be increased in obesity, the expression of adiponectin is decreased (107). Unlike many other adipokines, adiponectin has been correlated to insulin-sensitizing, anti-inflammatory, and anti-proliferative properties (8, 152, 203). Therefore, adiponectin has been attributed a cardioprotective role.

These observations demonstrate that the adipose tissue output dramatically changes in pathological conditions such as obesity. However, the adipokinome may already vary depending on the site of the adipose tissue depots (199). Adipose tissue in the visceral and the subcutaneous compartment are the two most abundant depots, and it has been shown that they produce unique profiles of adipokines (199, 287). In this context, visceral adipose tissue has received special attention since various studies have found a positive correlation between the amount of visceral adipose tissue and cardiovascular diseases (23, 216). Recent studies have even proposed that visceral adiposity, measured as waist circumference, is a more precise risk indicator for type 2 diabetes and cardiovascular diseases than whole body obesity (113, 160, 229, 239). On the

one hand this may be attributed to its location, as it drains directly into the portal vein (9). Special depots of visceral adipose tissue, perivascular and epicardial tissue, might also be located around blood vessels and the heart, respectively, where they specifically affect local tissues, as further discussed below. On the other hand, the visceral adipokinome contains many proinflammatory cardiovascular risk factors, such as IL-6 and plasmin activator inhibitor-1 (PAI-1) (10, 288), which contribute to the close association of visceral adipose tissue and cardiovascular disease.

#### *The Adipo-Vascular Axis*

Obesity is often associated with and represents a major risk factor for the development of cardiovascular diseases. Cardiovascular diseases are responsible for one of the highest mortality rates worldwide, accounting for 16.7 million deaths each year (49, 159), mostly because of the life-threatening complications of coronary artery and cerebrovascular disease (92). While cardiovascular diseases may be characterized by alterations like coronary artery calcification, thickening of vessel wall, formation of fatty streak and atherosclerotic plaques, vessel stiffness, and/or hypertension, atherosclerosis may be considered the principal contributor (2, 24, 228). Originally, atherosclerosis was believed to be a merely passive accumulation of cholesterol in the vessel wall; however, since novel data indicate underlying inflammatory processes to play a major contributing role (203, 221), this review aims to elucidate the role of proinflammatory adipokines in the pathogenesis of atherosclerosis.

To comprehend the influence of adipokines on the development of atherosclerosis, understanding the complex course of events taking place in the pathogenesis of atherosclerosis is important. It has been demonstrated that in the early stages of atherosclerosis, endothelial cells may be activated by various inflammatory stimuli, including a diet rich in saturated fat, hypercholesterolemia, obesity, hyperglycemia, insulin resistance, hypertension, and smoking, triggering the expression of adhesion molecules like vascular cell adhesion molecule-1 (VCAM-1), E-selectin, and intracellular adhesion molecule-1 (ICAM-1) (110, 203, 228). This increases the adherence of monocytes, which infiltrate the subendothelial space and accumulate within the intima (196, 198). In response to overexpression of macrophage colony-stimulating factor in the inflamed intima, which is induced by modified low-density lipoprotein, monocytes are converted to activated macrophages (203). These may then convert to foam cells by receptor-mediated incorporation and accumulation of lipoprotein particles (61, 203). The formation of foam cells and their continued accumulation in the intima, accompanied by proliferation and migration of smooth muscle cells (SMCs) from the media, leads to the first stage of the atherosclerotic lesion, the fatty streak (110). Continued exposure to atherosclerotic factors promotes the progression to more complex atherosclerotic plaques, until destabilizing factors like thinning of the fibrous cap and high foam cell content elicit rupture of a plaque, triggering thrombus formation (110). This in turn may either obstruct the lumen immediately or detach to form an embolus blocking blood flow distal to its origin. Consequently, such atherothrombosis may result in myocardial or brain infarction.

As mentioned above, a major risk factor for the development of atherosclerosis is obesity and the associated adipokines. To elucidate the molecular basis of this adipo-vascular axis, numerous studies have attempted to assess the impact of various adipokines on the cells of the vessel wall. In this context it has been described that various processes may be affected by adipokines. As abnormal proliferation and migration of SMCs located in the arterial intima has been suggested to be a central event in atherosclerosis (227, 241), adipokines able to induce proliferation or migration have to be considered as potential players in this process. Similarly, the activation of inflammatory signaling by adipokines like TNF- $\alpha$ , leptin, and PAI-1 has been suggested to contribute to the development of cardiovascular diseases (196, 245) by stimulating the generation of endothelial adhesion molecules, proteases, and other mediators, which may enter the circulation in soluble form (203). In this context the transcriptional regulator NF- $\kappa$ B plays a central role, as it mediates the expression of a multitude of genes. Among many others, the expression of adhesion molecules ICAM-1, VCAM-1, and E-selectin has been described to be mediated via NF- $\kappa$ B activation (197, 198). Thus adipokine-induced activation of NF- $\kappa$ B may promote adherence, diapedesis, and accumulation of immune cells such as monocytes or lymphocytes in the vessel wall, which play a central role in atherosclerotic plaque formation (203). Additionally, NF- $\kappa$ B activation is involved in SMC proliferation (144) and mediates the expression of a variety of proinflammatory molecules by macrophages and SMCs (14).

In recent years a number of studies have investigated the impact of selected adipokines on the different steps in the pathogenesis of atherosclerosis mentioned above. However, intensive research on the adipokinome in recent years has 1) demonstrated the very complex nature of the secretory output of adipocytes, 2) suggested novel roles for well-known adipokines, and 3) identified a number of novel adipokines associated with a vasoactive potential. As this multitude of studies conducted in various species and different models makes it difficult to identify promising candidates for drug target or biomarker validation, this review provides a novel summary of available data and evaluation of the vasoactive potential of currently known adipokines based on the evidence found in literature. As a general overview, Table 1 provides a list of adipokines currently described to be associated with cardiovascular disease, including their impact on proliferation, inflammation, and NF- $\kappa$ B activation. However, this list is likely to soon require updating as an increasing number of adipokines are rapidly identified and associated with a vasoactive potential.

The fact that this multitude of adipokines has been found to modulate vascular homeostasis underlines the pivotal role of the adipokinome in the adipose tissue/vessel cross talk. Because of these findings, we propose that obesity-induced inflammatory processes within the adipose tissue and the paracrine action of the associated proinflammatory adipokinome trigger endothelial dysfunction and vascular inflammation, which may ultimately lead to atherosclerosis, heart attack, or stroke (Fig. 1).

#### *Adipokines with a Tight Link to Cardiovascular Disease*

To analyze in more detail the adipokines presented in Table 1, we selected those adipokines with reported effects on NF- $\kappa$ B

activation. Since NF- $\kappa$ B is one of the major transcription factors that has been linked to both cardiovascular health and diseases, it is not surprising that NF- $\kappa$ B has been shown to influence numerous cardiovascular diseases including atherosclerosis (283). The function of NF- $\kappa$ B is largely dictated by the genes that it targets for transcription and varies according to stimulus and cell type (283). We are certainly aware that several adipokines induce cardiovascular diseases independent of NF- $\kappa$ B; however, this is not part of this review.

Here we have distinguished two groups of adipokines. The first group represents adipokines with a tight link to cardiovascular diseases based on evidences from in vitro and clinical studies (Table 2) as well as from animal models (Table 3). The second group includes adipokines with less strong evidence for the development of cardiovascular diseases, since data from animal models are not available either because the animal model has not been generated up till now or the specific knockout is lethal, as for VEGF.

*Adiponectin and leptin: classical adipokines.* Adiponectin, almost entirely produced by adipocytes, is one of the most comprehensively studied adipokines, and in human obese subjects its levels are diminished (176). It has been shown to positively influence energy consumption and fatty acid oxidation in muscle and liver, thereby reducing the triglyceride content (264). Transgenic mice overexpressing adiponectin exert an improved lipid profile (15, 194). Adiponectin has been suggested to be an important factor modulating the cardiovascular system because of its anti-atherogenic and anti-inflammatory effects. In macrophages and endothelial cells, it acts via suppression of TNF- $\alpha$  (196) and proinflammatory cytokines such as IL-6 (308) and directly ameliorates endothelial dysfunction by increasing nitric oxide (NO) production (52, 96). In addition, adiponectin reduces vascular SMC proliferation and migration (207). The role of adiponectin as a cardioprotective adipokine is further supported by results from clinical studies. While increased adiponectin levels are associated with a decreased risk of myocardial infarction (210), hypoadiponectinemia is observed in patients with coronary atherosclerosis and acute coronary syndrome (ACS) (139, 187). In a recent study, it has been shown that serum adiponectin is associated with biomarkers of insulin resistance, inflammation, and endothelial dysfunction, which are independent risk factors for cardiovascular diseases (65). In addition, results from animal studies have revealed that adiponectin exerts beneficial effects at mostly all stages of the atherosclerotic process (200).

Leptin, which directly influences food intake, was the first adipokine identified (90) and is primarily synthesized by white adipose tissue (164). The results of clinical studies investigating the contributions of leptin to the pathophysiology of cardiovascular complications are controversial, leaving the precise role of leptin unclear (266). Some studies have reported elevated leptin levels in patients with ACS (19, 246) and described an association between circulating leptin levels and risk of coronary artery disease (CAD) (289, 305). However, the findings of other studies have indicated no clinically relevant association with risk of CAD (26, 122, 302). The effects of leptin on endothelial cells and vascular SMCs are better investigated and comprise increased NO production via activation of endothelial NO synthase (eNOS) (285) and increased expression and activity of inducible NO synthase (iNOS), respectively (222). However, leptin also increases the expression of

Table 1. Overview of adipokines associated with cardiovascular disease

Adipokine	Proliferation	Inflammation	NF- $\kappa$ B Activation	Depot
Adiponectin	↓ (8, 144, 180)	↓ (197)	↓ (197)	↑ v(192, 258); ↑ sc(68, 155)
Adipsin	—	↑ (53, 276)	—	↑ v(10)
A-FABP	↑ (58)	↑ (152)	↑ (152)	↑ v(89)
Angiotensin II	↑ (47, 180)	↑ (236, 314)	↑ (279)	—
ANGPTL2	—	↑ (193, 199)	—	↑ sc(78)
Apelin	—	↑ (53)	—	—
Chemerin	↑ (125)	↑ (93) ↓ (318)	↑ (242)	↔ (84)
CXCL5	—	↑ (199)	—	—
DPP-4	↑ (143)	—	—	↑ v(143)
G-CSF	↑ (40)	↑ (262)	—	—
IL-1 $\beta$	↑ (180, 276)	↑ (53, 276)	↑ (307, 314)	↔ (233)
IL-4	↑ (274) ↓ (282)	↑ (53)	↔ (307)	—
IL-6	↑ (180, 276)	↑ (53, 195, 199)	—	↑ v(62, 73)
IL-8	↑ (180)	↑ (53)	↑ (172)	↔ (233); ↑ v(28)
IL-10	↓ (180)	↓ (337)	↓ (178, 337)	(—)
IL-18	↑ (37)	↑ (53, 199)	↑ (37)	(—)
Leptin	↑ (108, 180, 304)	↑ (195, 199, 276)	↑ (108)	↑ sc(10, 182)
Lipocalin 2	—	↑ (199) ↓ (330)	↓ (60)	—
MCP-1	↑ (180, 276)	↑ (53, 199)	—	↑ v(29)
MIF	↑ (180) ↔ (240)	↑ (53, 240)	↑ (232)	↑ v(5); ↔ (254)
MIP-1a	↑ (162, 276)	↑ (98)	—	—
Nesfatin-1	—	↑ (318)	—	↑ sc(213)
Omentin	—	↓ (318, 319)	—	↑ v(321)
PAI-1	↑ (41, 54, 180); ↓ (137)	↑ (203) ↓ (212)	—	↑ v(62, 245); ↑ sc(59); ↔ (3)
PDGF	↑ (333)	—	—	—
PEDF	↑ (64)	↑ (64)	↑ (64)	↑ v(291)
RANTES	↔ (244)	↑ (177)	—	↑ sc(163)
RBP4	—	↑ (111, 199)	—	↑ v(60); ↑ sc(233)
Resistin	↑ (180)	↑ (22, 141, 199)	↑ (22, 247)	↔ (179)
Sfrp5	—	↓ (195, 199)	—	—
TGF $\beta$	↑ (277)	↑ (276)	—	↔ (3)
TNF $\alpha$	↑ (180, 276)	↑ (195, 199)	↑ (198, 307)	↔ (10, 89)
TSP1	↑ (260) ↓ (265)	↓ (260)	↓ (271)	↑ v(214)
Vaspin	—	↓ (208, 318)	↓ (208)	↑ v(272)
VEGF	↑ (32, 66, 125)	↑ (129)	↑ (175)	↑ v(62)
Visfatin	↑ (180)	↑ (199, 225)	↑ (225)	↔ (17)

A-FABP, adipocyte fatty acid-binding protein; ANGPTL2, angiopoietin-like 2; CXCL5, C-X-C motif chemokine 5; DPP-4, dipeptidyl peptidase-4; G-CSF, granulocyte colony-stimulating factor; IL, interleukin; MCP-1, monocyte chemoattractant protein-1; MIF, macrophage migration inhibitory factor; MIP-1 $\alpha$ , macrophage inflammatory protein-1 $\alpha$ ; PAI-1, plasminogen activator inhibitor-1; PDGF, platelet-derived growth factor; PEDF, pigment epithelium-derived factor; RANTES, regulated upon activation, normal T-cell expressed, and secreted; RBP4, retinol binding protein 4; Sfrp5, secreted frizzled-related protein 5; TGF- $\beta$ , transforming growth factor- $\beta$ ; TNF- $\alpha$ , tumor necrosis factor- $\alpha$ ; TSP1, thrombospondin 1; VEGF, vascular endothelial growth factor; ↑, parameter activated; ↓, parameter decreased; ↔, parameter unchanged; ↑ v, adipokine predominantly expressed in visceral adipose tissue; ↑ sc, adipokine predominantly expressed in subcutaneous adipose tissue.

PAI-1 (250) and CRP (249) in human vascular endothelial cells.

Furthermore, leptin-deficient mice, which are extremely obese, are protected from atherosclerosis despite other metabolic risk factors, indicating that this adipokine contributes directly to cardiovascular diseases (95).

**TNF- $\alpha$  and macrophage migration inhibitory factor: macrophage-associated cytokines.** TNF- $\alpha$  is one of the most extensively examined proinflammatory cytokines that plays an important role in atherosclerosis as well as in other inflammatory and metabolic disorders, which are known risk factors for cardiovascular diseases. Upregulation of TNF- $\alpha$  in the vascular wall of carotid and coronary arteries promotes endothelial apoptosis thus leading to impairment of endothelial function (45, 46). Moreover, TNF- $\alpha$  induces phenotypic changes in vascular SMCs (263) as well as their migration (82, 83, 118, 263), proliferation (215), and apoptosis (300), which are all critical for the initiation and progression of vascular lesions. In TNF- $\alpha$ /apolipoprotein E (ApoE) double knockout mice most proatherosclerotic factors such as IL-1 $\beta$ , MCP-1, and NF- $\kappa$ B

are downregulated (309). Furthermore, TNF- $\alpha$  plasma concentrations are positively correlated with carotid intima-media thickness (IMT) (252) and increased in patients with premature CAD (118), thus further emphasizing the important role of TNF- $\alpha$  for the development of cardiovascular diseases.

Macrophage migration inhibitory factor (MIF) is expressed in various tissues such as adipose tissue and regulates acute inflammatory as well as adaptive immune reactions (70). MIF expression is induced by proatherogenic stimuli, such as oxidized low-density-lipoprotein (31), and it has been shown to become upregulated in macrophages, endothelial cells, and SMCs during the development of atherosclerotic lesions (18, 31). Its expression correlates with increased IMT and lipid deposition in the aorta of mice and in advanced human carotid artery plaques (136, 238). A recent study suggested high MIF levels as an independent risk factor for future coronary events in CAD patients with impaired glucose tolerance/type 2 diabetes mellitus (166). MIF influences the proliferation and migration of macrophages and vascular cells (205, 240), and MIF-deficient SMCs display impaired proliferation. MIF defi-

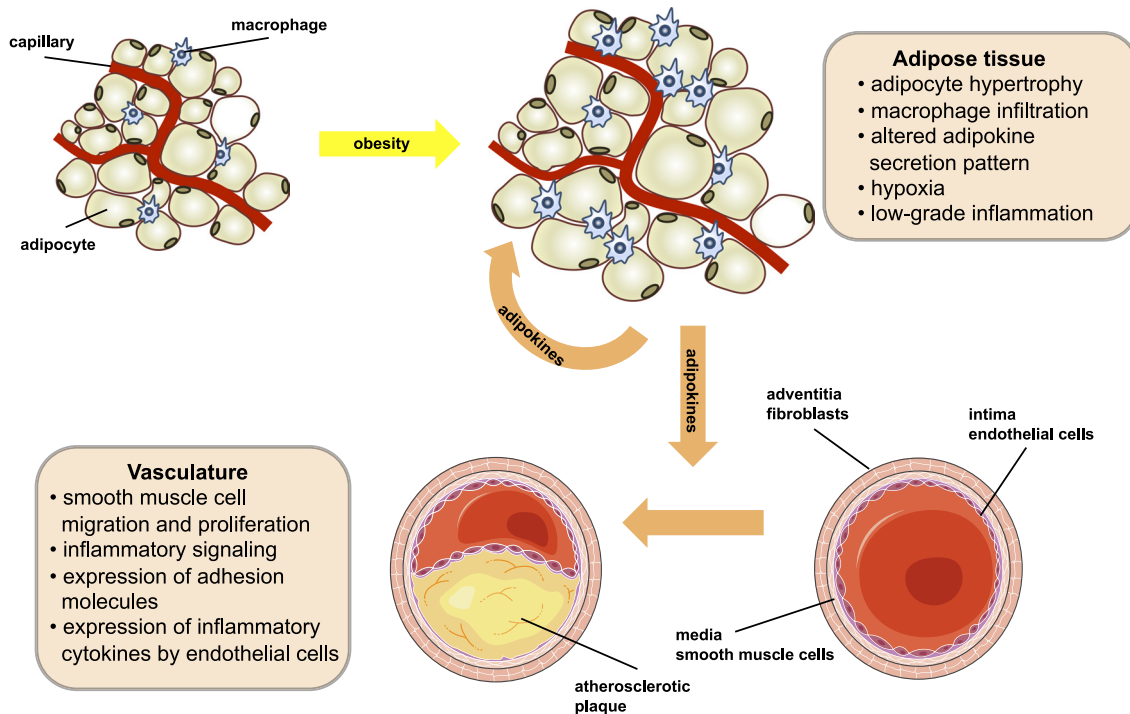


Fig. 1. Obesity-induced adipose tissue inflammation triggers endothelial dysfunction and vascular inflammation.

ciency in LDL receptor-knockout mice prevented diet-induced atherogenesis, as shown by decreased IMT and lipid deposition in the aorta (205). Taken together, these results suggest that MIF could be an important player in the pathogenesis of atherosclerosis and may represent a potential drug target for the treatment of inflammatory and cardiovascular diseases (184).

**Adipocyte fatty acid-binding protein and lipocalin-2: small lipid-binding proteins.** Adipocyte fatty acid-binding protein (A-FABP) is one of the most abundant intracellular lipid transport proteins in adipocytes (167, 312), regulating lipid metabolism by promoting diffusion, sequestration, and transport of long-chain fatty acids (71). In addition, A-FABP is secreted and it has been shown that high levels are associated with a worse cardiometabolic risk profile (311). Furthermore, A-FABP serum levels are positively associated with the metabolic syndrome (106), CAD (220), and carotid IMT (324), whereas they are inversely associated with endothelial function (7). In human endothelial cells, the expression of A-FABP can be induced by VEGF-A, bFGF (58), and lipids, leading to reduced activity of eNOS and NO production (146). In addition, knockdown of A-FABP reduced endothelial cell proliferation (58).

The expression of lipocalin-2 in adipocytes was first described by Lin and colleagues (157), and is markedly induced during differentiation of preadipocytes to adipocytes (16). Clinical as well as experimental studies indicate an important role of lipocalin-2 as an inflammatory adipokine in obesity and related complications (34, 60, 297). In patients with CAD lipocalin-2 levels are increased and independently associated with systolic arterial blood pressure, insulin resistance, and decreased HDL cholesterol levels (43). In addition, a high expression of lipocalin-2 has been shown in atheromatous human plaques that were associated with increased matrix metalloproteinase-9 activity (273). Serum lipocalin-2 levels are

also elevated in various obese rodent models and human obesity (297, 320, 330) and positively correlated with adiposity, hypertriglyceridemia, hyperglycemia, insulin resistance, and high-sensitive CRP (297). In vascular SMCs, mRNA and protein expression of lipocalin-2 is increased upon IL-1 $\beta$  treatment in a NF- $\kappa$ B-dependent manner (30). Furthermore, lipocalin-2 knockout mice are protected against diet-induced endothelial dysfunction (158). However, lipocalin-2 may also have anti-inflammatory effects since it suppresses LPS-induced cytokine production in macrophages and antagonizes effects of TNF- $\alpha$  on adipocytes and macrophages (330).

A-FABP as well as lipocalin-2 are adipokines linking obesity with vascular diseases and are involved in the pathogenesis of atherosclerotic plaque. Interestingly, serum levels of A-FABP are positively associated with those of lipocalin-2 (63, 181, 278, 311). The increased release of these two adipokines in conditions of obesity may contribute to the pathogenesis of endothelial dysfunction and atherosclerosis.

**Interleukins: family of immune system's messengers.** Proinflammatory cytokines of the interleukin family are considered to be key players in the chronic vascular inflammation that is typical for atherosclerosis and cardiovascular diseases (171). IL-1 $\beta$  is a prototypic proinflammatory cytokine with different biological functions, inducing the production of different cytokines and chemokines. In vitro studies demonstrated that IL-1 $\beta$  increases the expression of cell adhesion molecules (295), MCP-1 (156), and lipocalin-2 (30) in vascular SMCs. In addition, it stimulates the migration (299) and proliferation of these cells (128, 234). In the ApoE/IL-1 $\beta$  double knockout mouse model, IL-1 $\beta$  deficiency decreases the severity of atherosclerosis (121, 134), further supporting the important role of IL-1 $\beta$  in vascular disorders.

IL-4 is a proinflammatory cytokine and plays a critical role in the progression of atherosclerosis. In endothelial cells, IL-4

Table 2. Overview of adipokines with a tight link to cardiovascular diseases based on data obtained in *in vitro* and *in vivo* studies

Adipokine	In Vitro Studies	Clinical Studies
Adiponectin	<ul style="list-style-type: none"> <li>- Suppression of inflammatory cytokines (196, 308)</li> <li>- EC: <math>\uparrow</math> NO production (97), <math>\downarrow</math> NO inactivation (52), <math>\downarrow</math> apoptosis (131, 310, 332)</li> <li>- VSMC: <math>\downarrow</math> proliferation (8, 144, 207), <math>\downarrow</math> migration (8, 165)</li> </ul>	<ul style="list-style-type: none"> <li>- Inversely associated with markers of insulin secretion, endothelial function, and inflammation (65)</li> <li>- Hypoadiponectinemia in patients with coronary atherosclerosis (139) and ACS (187)</li> <li>- Increased levels associated with a decreased risk of myocardial infarction in healthy men (210)</li> </ul>
A-FABP	<ul style="list-style-type: none"> <li>- EC: expression induced by VEGF-A, bFGF (58), and lipids (146), associated with <math>\downarrow</math> phosphorylated eNOS and <math>\downarrow</math> NO production (146), <math>\uparrow</math> proliferation (58)</li> </ul>	<ul style="list-style-type: none"> <li>- High levels associated with worse cardiometabolic risk profile (311)</li> <li>- Serum levels positively associated with the Metabolic Syndrome (106), CAD (220), carotid IMT (324)</li> <li>- Circulating levels inversely associated with endothelial function (7)</li> <li>- High levels of IL-1<math>\beta</math> in patients with unstable angina (202)</li> </ul>
IL-1 $\beta$	<ul style="list-style-type: none"> <li>- EC: <math>\uparrow</math> VCAM-1 ectodomain release (251) and <math>\uparrow</math> expression of MCP-1 (174)</li> <li>- VSMC: <math>\uparrow</math> expression of cell adhesion molecules (296), MCP-1 (156), and lipocalin-2 (30)</li> <li>- VSMC: <math>\uparrow</math> migration (299); <math>\uparrow</math> proliferation (128, 234)</li> </ul>	<ul style="list-style-type: none"> <li>- <math>\uparrow</math> Levels of IL-1<math>\beta</math> mRNA in coronary arteries of patients with ischemic heart disease (76)</li> </ul>
IL-4	<ul style="list-style-type: none"> <li>- EC: <math>\uparrow</math> expression of inflammatory mediators (39, 224), <math>\uparrow</math> ROS generation (149), <math>\uparrow</math> apoptosis (148)</li> <li>- VSMC: <math>\uparrow</math> proliferation and 12-lipoxygenase expression (190), <math>\uparrow</math> migration (294)</li> </ul>	<ul style="list-style-type: none"> <li>- Circulating levels higher in patients with CAD (115)</li> <li>- IL-4 mRNA rarely (74) or not observed in atherosclerotic plaques (280)</li> </ul>
IL-8	<ul style="list-style-type: none"> <li>- Modulator of monocyte-endothelial interaction under flow conditions (81)</li> <li>- EC: expression induced by TNF<math>\alpha</math> (322) and homocysteine (79), <math>\uparrow</math> proliferation (204), <math>\uparrow</math> migration (267, 338)</li> <li>- VSMC: <math>\uparrow</math> proliferation and 12-lipoxygenase expression (190), <math>\uparrow</math> migration (298, 328), regulation of VCAM-1 (94, 331)</li> </ul>	<ul style="list-style-type: none"> <li>- Concentrations significantly higher in CAD patients (100)</li> <li>- Levels predicts cardiovascular events (112)</li> <li>- Higher levels in patients with early coronary atherosclerosis (85)</li> <li>- Expression increased in coronary atherectomy tissue (248) and atherosclerotic plaques (124)</li> </ul>
IL-10	<ul style="list-style-type: none"> <li>- Macrophages: inhibition of inflammatory molecules, <math>\downarrow</math> apoptosis (91)</li> <li>- EC: inhibition of TNF<math>\alpha</math>-, IL-1<math>\beta</math>- or LPS-induced expression of IL-6 and IL-8 (39), <math>\uparrow</math> eNOS expression, <math>\uparrow</math> NO production (35)</li> <li>- VSMC: inhibition of TNF<math>\alpha</math>- and bFGF-stimulated proliferation and migration (178, 243)</li> </ul>	<ul style="list-style-type: none"> <li>- Elevated plasma levels predict long-term adverse outcomes in ACS (36)</li> <li>- Some studies report higher levels in patients with ACS (133, 183), while other studies report lower levels in patients with CAD (115) and unstable angina (256)</li> </ul>
IL-18	<ul style="list-style-type: none"> <li>- EC: secretion induced by CRP (317), <math>\uparrow</math> apoptosis (37, 335)</li> <li>- VSMC: <math>\uparrow</math> proliferation (37, 217), <math>\uparrow</math> migration (37), <math>\uparrow</math> expression of IL-6, IL-8 and MCP-1 (230)</li> </ul>	<ul style="list-style-type: none"> <li>- Expressed in human carotid atherosclerotic plaques (169)</li> <li>- Serum levels as a strong independent predictor of death from cardiovascular causes in patients with CAD (20)</li> <li>- Serum levels associated with carotid IMT (315)</li> <li>- Elevated plasma levels in patients with ACS (19, 246)</li> </ul>
Leptin	<ul style="list-style-type: none"> <li>- EC: <math>\uparrow</math> PAI-1 (250) and CRP expression (249), <math>\uparrow</math> eNOS activation and NO production (285)</li> <li>- VSMC: <math>\uparrow</math> NOS expression and activity leading to <math>\uparrow</math> NO production (222)</li> </ul>	<ul style="list-style-type: none"> <li>- Some studies report an association with risk of CAD (289, 305), while other studies found no association with risk of CAD (26, 122, 302)</li> </ul>
Lipocalin-2	<ul style="list-style-type: none"> <li>- VSMC: <math>\uparrow</math> expression after IL-1<math>\beta</math> treatment via NF-<math>\kappa</math>B (30)</li> <li>- Macrophages: suppression of LPS-induced cytokine production (330)</li> </ul>	<ul style="list-style-type: none"> <li>- Levels positively associated with e.g., adiposity, hypertriglyceridemia, high sensitivity CRP (297), and CAD (43)</li> <li>- High expression in atheromatous human plaques, associated with increased MMP-9 activity (273)</li> </ul>
MIF	<ul style="list-style-type: none"> <li>- EC and macrophages: induction by oxLDL (31)</li> <li>- VSMC: <math>\uparrow</math> migration after short-term exposure (240)</li> <li>- MIF-deficient smooth muscle cells: impaired proliferation and lower proteolytic capacity (205)</li> </ul>	<ul style="list-style-type: none"> <li>- Upregulated during progression of atherosclerosis toward inflammatory stages (31)</li> <li>- High levels as independent risk factor for future coronary events in CAD patients with IGT/T2DM (166)</li> </ul>
PDF	<ul style="list-style-type: none"> <li>- EC: inhibition of VEGF-induced proliferation and migration (56)</li> <li>- VSMC: <math>\uparrow</math> proliferation, activation of inflammatory signaling pathways (64)</li> <li>- VSMC: inhibition of PDGF-BB-induced proliferation and migration (186, 293)</li> </ul>	<ul style="list-style-type: none"> <li>- Higher levels in T2DM patients (114)</li> <li>- Levels strongly associated with the Metabolic Syndrome (316), vascular inflammation, and carotid IMT (268)</li> </ul>
TNF- $\alpha$	<ul style="list-style-type: none"> <li>- EC: <math>\uparrow</math> apoptosis (46)</li> <li>- VSMC: induction of phenotypic changes (263), <math>\uparrow</math> migration (83, 118), <math>\uparrow</math> proliferation (215), <math>\uparrow</math> apoptosis (300)</li> </ul>	<ul style="list-style-type: none"> <li>- Increased plasma concentrations in patients with premature coronary artery disease (117)</li> <li>- Levels positively correlated with carotid IMT (252)</li> </ul>
TSP-1	<ul style="list-style-type: none"> <li>- EC: <math>\uparrow</math> expression of cell adhesion molecules (189)</li> <li>- VSMC: stimulation of chemotaxis (147, 191)</li> <li>- <math>\uparrow</math> Aggregation and stability of platelet aggregates (188)</li> </ul>	<ul style="list-style-type: none"> <li>- No data available</li> </ul>

ACS, acute coronary syndrome; bFGF, basic fibroblast growth factor; CAD, coronary artery disease; CRP, C-reactive protein; EC, vascular endothelial cells; eNOS, endothelial nitric oxide (NO) synthase; IGT, impaired glucose tolerance; IMT, intima-media thickness; iNOS, inducible NO synthase; MMP, matrix metalloproteinase; oxLDL, oxidized low-density lipoprotein; ROS, reactive oxygen species; T2DM, type 2 diabetes mellitus; VSMC, vascular smooth muscle cells.

Table 3. Overview of adipokines with a tight link to cardiovascular diseases based on data obtained in animal models

Animal Studies	
Adiponectin	
KO model	- ↑ Leukocyte-endothelial cell interactions; ↑ E-selectin and VCAM-1 expression; ↓ endothelial NO production (200)
Overexpression	- ↓ Adiposity, altered expression of lipogenic enzymes; ↑ expression of uncoupling proteins (15) - ↓ Fat storage, morbidity and mortality, oxidative DNA damage upon high-fat diet (194)
A-FABP	
KO model	- ↑ Obesity, but no insulin resistance or diabetes, no TNF- $\alpha$ expression in adipose tissue (105) - Improved peripheral insulin resistance, beneficial effect on pancreatic $\beta$ -cell function and lipid metabolism (281)
IL-1 $\beta$	
KO model	- ↓ Aortic sinus lesions and lesion area (121, 134) - ↓ VCAM-1 and MCP-1 expression (134)
IL-4	
KO model	- ↓ Plaque area (50) - In contrast, no protection from early atherosclerosis; no differences in the presence and activity of 12/15-lipoxygenase in macrophages (80)
IL-8	
KO model	- ↓ Atherosclerotic lesions (21)
IL-10	
KO model	- ↑ Lesions (209); ↑ susceptibility to atherosclerosis, lipid accumulation, and T-cell infiltration ↓ collagen content (168)
Overexpression	- ↓ Lesions (209)
IL-18	
KO model	- ↓ Lesion size with a more stable phenotype; ↑ serum cholesterol (57)
Leptin	
KO model	- ↑ Plasma cholesterol and triglyceride levels; extensive atherosclerotic lesions throughout the aorta (95)
Lipocalin2-	
KO model	- ↓ Endothelial dysfunction; ↑ basal and insulin-stimulated Akt/eNOS phosphorylation in the aorta (158)
MIF	
KO model	- ↓ Abdominal aorta lipid deposition and IMT, marked retardation of atherosclerosis, ↓ cell proliferation (205)
PEDF	
KO model	- ↑ Microvascular density, excessive angiogenesis, epithelial cell hyperplasia (55)
TNF- $\alpha$	
KO model	- ↓ Fatty-streak lesions; ↓ expression of proatherosclerotic factors (309)
TSP-1	
KO model	- ↑ Postinfarction inflammatory response, infiltration of neighboring noninfarcted area with macrophages and myofibroblasts; ↑ left ventricular remodeling (72)

KO, knockout.

increases the expression of inflammatory mediators (39, 224), the generation of reactive oxygen species (149), as well as apoptosis (148), whereas it induces proliferation (190) and migration (294) in vascular SMCs. A recent study described higher circulating IL-4 levels in patients with CAD (116), whereas IL-4 mRNA is rarely (74) or not observed in atherosclerotic plaques (280). Data obtained in animal models are controversial. Whereas Davenport and Tipping (50) showed that IL-4 deficiency reduced plaque area and atherosclerotic lesions, George et al. (80) observed no protection from early atherosclerosis.

IL-8 is another cytokine that plays a role in the context of cardiovascular diseases (6). It has been described as a modulator of monocyte-endothelial interaction (81), and its expression is induced by TNF- $\alpha$  (322) and homocysteine (79). In addition, IL-8 promotes endothelial cell proliferation (204) and migration (267, 338), the latter via activation of phosphatidylinositol 3-kinase-Rac1/RhoA pathway (142). In vascular SMCs, IL-8 has been shown to influence VCAM-1 expression (94, 331), proliferation (190), and migration (298, 328).

Moreover, IL-18 act as a proatherogenic factor, as supported by data from IL-18-deficient mice that display reduced atherosclerosis (57). A recent study demonstrated that IL-18 induced proliferation of human SMCs and matrix metalloproteinase induction (217).

In contrast, IL-10 is an anti-inflammatory cytokine, which is thought have an anti-atherogenic potential. IL-10 inhibits TNF-

$\alpha$ -, IL-1 $\beta$ -, or LPS-induced expression of IL-6 and IL-8 in endothelial cell (39) and increases eNOS expression and NO production (35). In vascular SMCs, IL-10 prevents migration and proliferation, partially mediated by NF- $\kappa$ B inactivation (178, 243). Furthermore, IL-10 transgenic mice showed reduced atherosclerosis, whereas IL-10-deficient mice exhibited increased early atherosclerotic lesion formation (193), illustrating the anti-atherogenic potential of IL-10. However, results from clinical studies are not consistent as some report higher IL-10 levels in patients with ACS (133, 183), whereas other report lower levels in patients with CAD (115) and unstable angina (256).

*Thrombospondin-1 and PEDF: proteins associated with angiogenesis.* Thrombospondin-1 (TSP-1) is a member of a protein family that mediates cell-matrix and cell-cell interactions. Experimental data have shown that TSP-1 stimulates the aggregation and stability of platelet aggregates (188), induces the expression of cell adhesion molecules (189) in endothelial cells, and stimulates chemotaxis in vascular SMCs (147, 191). In TSP-1 knockout mice, more extensive postinfarction remodeling has been observed compared with wild-type mice (72). Until now, no clinical data are available for TSP-1.

PEDF, first identified in retinal pigment epithelial cells, is a multifunctional, pleiotropic protein and is expressed in various tissues such as adipose tissue. The role of PEDF in cardiovascular diseases is not completely understood. PEDF has been shown to possess antiangiogenic effects (51) and to inhibit

VEGF-induced endothelial cell migration and proliferation (56) as well as PDGF-BB-induced proliferation and migration of vascular SMCs (186). However, our group recently reported an increased proliferation of vascular SMCs upon PEDF treatment as well as the activation of inflammatory signaling pathways (64). In clinical studies higher PEDF levels in type 2 diabetic patients (114, 316) have been found, and it was shown that PEDF levels are strongly associated with the Metabolic Syndrome (316), vascular inflammation, and carotid IMT (268).

In Fig. 2 we provide an additional schematic overview of the above-described adipokines summarizing their impact on central features of cardiovascular diseases such as proliferation, migration, NO production, and induction of inflammatory cytokines.

#### Adipokines with Less Strong Evidence for Involvement in Cardiovascular Disease

The following adipokines are also important factors for the development of cardiovascular disease, but these adipokines have diminished evidence linking NF- $\kappa$ B inflammation and cardiovascular diseases, since KO or transgenic studies have not been reported.

*Chemerin, resistin, visfatin, and vaspin: novel adipokines.* Chemerin is a newly described adipokine that is highly expressed in adipose tissue and liver and effects adipocyte metabolism (223). In humans, its plasma levels have been shown to be associated with inflammation and various components of the Metabolic Syndrome such as BMI, triglycerides, and hypertension (151, 259), whereas no differences were observed between subjects who are nondiabetic and those who have type 2 diabetes (25). Studies in rodents that investigated the role of chemerin with regard to obesity and diabetes have revealed controversial data. In obese *db/db* mice, the expression in

adipose tissue and serum levels of chemerin are reduced compared with lean controls (269), whereas its expression is higher adipose tissue of obese diabetic *Psammomys obesus* compared with lean normoglycemic *P. obesus* (25). Experimental data have shown that chemerin promotes proliferation and migration of endothelial cells (125). However, until now the effects of chemerin on vascular SMCs have not been investigated, and future studies have to determine the impact of chemerin on cells of the vessel wall in relation to cardiovascular diseases.

Resistin was identified as adipokine in 2001 and shown to be increased in diet-induced and genetic forms of obesity in rodents (261) as well as in morbidly obese subjects compared with lean controls (235). In humans, resistin is expressed in and secreted by monocytes and macrophages in addition to adipocytes (140, 170). It promotes vascular SMC migration (120) and proliferation (33), induces monocyte-endothelial cell adhesion, and increases the expression of VCAM-1, ICAM-1, and MCP-1 in endothelial cells (286). Clinical studies have revealed that plasma resistin levels correlate with markers of inflammation and are predictive of coronary atherosclerosis in humans (218). Resistin may therefore represent a potential link between obesity and cardiovascular diseases.

Visfatin is an adipokine that was suggested to act as a proinflammatory since it induces cytokine production (185). In clinical studies it has been shown that visfatin expression was increased in plaques from patients with unstable carotid and coronary atherosclerosis (48). In addition, adipose tissue levels of visfatin were significantly higher in CAD patients relative to control subjects (42). Visfatin induces endothelial cell proliferation and migration, induces eNOS and iNOS, and has antiapoptotic effects in both vascular cell types (1, 161, 225, 284). These data indicate that visfatin may be implicated in the

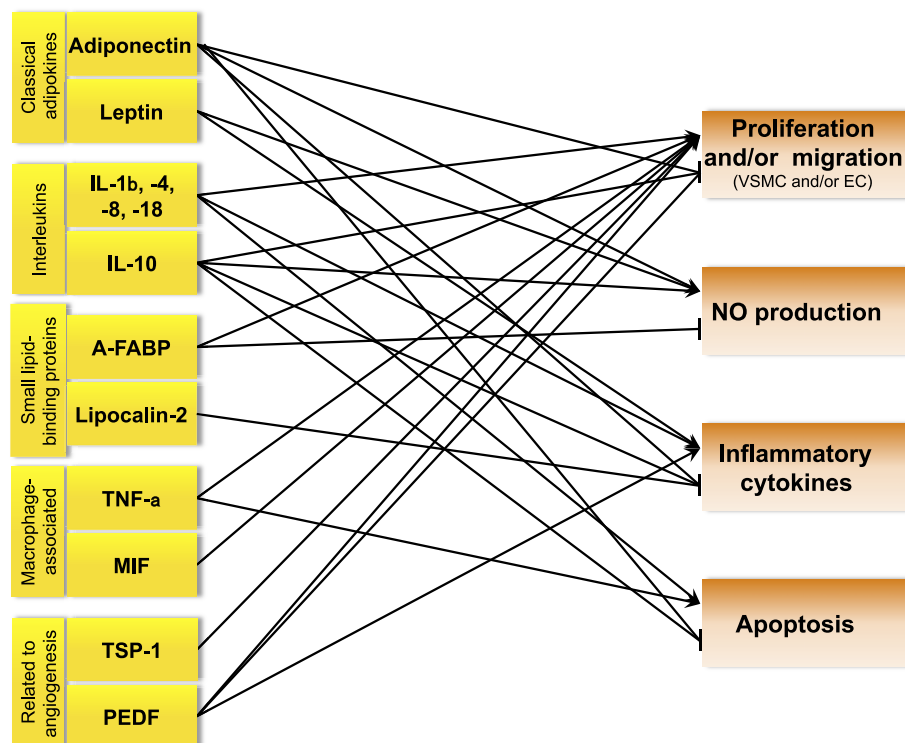


Fig. 2. Effects of selected adipokines. Arrow indicates stimulation. Line indicates inhibition of the respective process. A-FABP, adipocyte fatty acid-binding protein; MIF, macrophage migration inhibitory factor; TSP-1, thrombospondin-1; PEDF, pigment epithelium-derived factor; VSMC, vascular smooth muscle cells; EC, endothelial cells; NO, nitric oxide.



pathogenesis of atherosclerosis and cardiovascular disease (77). Further studies will be needed to clearly define the impact of visfatin in this context.

Vaspin, a serine protease inhibitor, was originally identified as an adipokine predominantly secreted from visceral adipose tissue in Otsuka Long-Evans Tokushima fatty, an animal model of obesity and type 2 diabetes (101). In humans, vaspin mRNA expression in adipose tissue is regulated in a fat depot-specific manner and is associated with parameters of obesity, insulin resistance, and glucose metabolism (135). Plasma levels of vaspin are associated with age, sex, BMI, and insulin resistance (44, 321, 325). Studies investigating an association between circulating vaspin and cardiovascular diseases reported controversial results. One study have reported an association between plasma level and atherosclerosis in women (44), whereas another study have found no association between vaspin concentration and parameters of atherosclerosis severity (11). However, the expression of vaspin has been described in vascular SMCs and foam cells in atherosclerotic lesions (257). In addition, antiapoptotic effects of vaspin in endothelial cells have been reported (119). In vascular SMCs, vaspin have been shown to inhibit TNF- $\alpha$ -induced reactive oxygen species generation, NF- $\kappa$ B activation, and expression of ICAM-1, pointing toward a protective role of vaspin (208). Clearly, further investigations have to be conducted to clarify the impact of vaspin for cardiovascular disease.

*Angiotensin II and VEGF: proliferative proteins.* Angiotensin II, the major effector of the renin angiotensin system, has many functions including vasoconstriction, cell growth, generation of oxidative stress, and inflammation. Angiotensin II has proinflammatory effects in the vascular wall by inducing gene expression of inflammatory cytokines and cell adhesion molecules (173). In vitro studies have revealed that angiotensin II induces proliferation and migration in human vascular SMCs (323) and endothelial cells (336). These experimental findings point to the important cardiovascular actions of angiotensin II. However, the precise mechanisms of angiotensin II in the context of cardiovascular diseases need further investigations.

VEGF is a well-characterized growth factor, which is involved in the regulation and differentiation of the vascular system. However, the results of studies investigating the role of VEGF in the context of cardiovascular diseases are controversial until now. On the one hand, studies predict a beneficial role VEGF for cardiovascular health by enhancing protective vascular functions (255). But on the other hand data obtained in some studies in mouse models of atherosclerosis seem to promote a proatherogenic role of angiogenesis (127, 255). Although VEGF has an important role in various physiological processes, the very same qualities cause it to play a part in the origin and maintenance of various pathological processes, including atherosclerosis (103). To date, our understanding of the mechanisms and the precise role of VEGF in the context of cardiovascular disease in humans remains unclear and is an important question to be addressed in future studies.

#### *Perivascular Adipose Tissue and Cardiovascular Diseases*

Perivascular adipose tissue (PVAT) is defined as adipose tissue around blood vessels that occurs in a way that no fascial layer separates this fat depot from the vascular wall. In addition to this barrier-free connection between PVAT, an infiltration of

adipocytes into the outer region of the adventitia has been observed (38). In obesity, PVAT is increased in humans and rodents (126, 150). The amount of PVAT was highly associated with visceral obesity and moderately correlated with subcutaneous adipose tissue and body mass index (237). Adipocytes in PVAT have been compared with subcutaneous and visceral adipocytes in humans and rodents in various studies, and there is still controversy as for the classification of PVAT as a depot of white adipose tissue or brown adipose tissue. It appears that PVAT surrounding abdominal and thoracic aortas might be multifaceted as for its adipocyte phenotype (38, 211).

In the obese state, adipose tissue is characterized by infiltration of various immune cells including macrophages and T lymphocytes (301) and low-grade chronic inflammation. Adipocytes of patients who are obese are characterized by increased release of various proinflammatory adipokines such as IL-6 and MCP-1. Most of the studies analyzing differences in adipokine expression and release of adipocytes in the obese state work with subcutaneous or visceral adipocytes. In contrast, little is known on adipokine expression and release in PVAT compared with other fat depots and in pathological states. In vitro differentiated human PVAT adipocytes are characterized by lower adiponectin release and higher secretion of MCP-1 (38). In rodents, PVAT expression of adiponectin and FABP4 (A-FABP) was lower and expression of leptin and MCP-1 was higher in high-fat diet-fed mice compared with those of controls (38, 211). Data on the presence of immune cells in PVAT are controversial. A very recent publication analyzed macrophage content in thoracic PVAT compared with white and brown adipose tissue and found PVAT to be resistant to high-fat diet-induced immune cell infiltration similar to brown adipose tissue (303). In contrast, adipose tissue inflammation in PVAT and macrophage infiltration could be described to be responsible for a loss of anticontractile function of this fat depot in the mesenteric bed (69). Furthermore, adipokines released from PVAT strongly induced the chemotaxis of peripheral blood leukocytes to the interface between PVAT and the adventitia in human atherosclerotic arteries (99). These chemotactic effects have been ascribed to IL-8, and MCP-1 and have been proposed to underlie the accumulation of macrophages and T cells in atherosclerosis.

Vascular relaxation factors, proatherogenic and proinflammatory adipokines, and growth factors secreted from PVAT were found to directly regulate vascular function through paracrine and endocrine effects on the vascular wall. Adipokines that are increased in the obese state such as leptin have been described to affect vascular function on the level of endothelial dysfunction (206). Endothelial dysfunction preceding atherosclerosis is characterized by deregulation of vasoreactivity, increased inflammatory and oxidative stress, and impaired barrier function (12). Oxidative stress has been shown to be increased in PVAT in obesity (231). Vasorelaxing effects of PVAT are lost in human obesity, which is associated with expansion of PVAT (86). The addition of TNF- $\alpha$  and inhibition of adiponectin inhibit the vasodilator activity of PVAT around healthy blood vessels, whereas the blocking of TNF- $\alpha$  by specific antibodies could reverse the obesity-induced defects in vasodilatation. In addition to TNF- $\alpha$  and adiponectin, leptin and resistin could be described to be mediators of endothelial dysfunction (27).

Migration of vascular SMCs from the media to the intima and their proliferation in the synthetic state are crucial steps in arterial wall thickening in atherosclerosis. In vitro, PVAT explants induce proliferation of vascular SMCs (13, 144). Secretory products of PVAT from diet-induced obese rats significantly induced human SMC proliferation compared with that from lean controls (13). Increased neointima formation diet-induced obese mice was accompanied by a decreased expression of adiponectin and induction of inflammatory markers such as MCP-1, TNF- $\alpha$ , IL-6, and PAI-1 in PVAT (270). Importantly, adiponectin-deficient mice display increased neointima formation when compared with wild-type mice, and this effect could be reversed by a local administration of adiponectin to the periadventitial area (138). In line with this, adiponectin has been found to abrogate adipokine-induced SMC proliferation (144). It is currently unknown which factors secreted from PVAT contribute to vascular SMC migration and proliferation. Potential candidates include leptin, resistin, and visfatin, which have been found to directly affect SMCs (33, 153, 292).

For the comprehensiveness of this review, a short paragraph should be devoted to epicardial adipose tissue that is a major fat depot associated with obesity and cardiovascular diseases. As epicardial adipose tissue is directly lying on the surface of the myocardium and also in direct contact with coronary vessels, this depot can also be seen as a special PVAT. While an association between epicardial adipose tissue thickness and the prevalence of cardiovascular diseases and the metabolic syndrome is widely accepted [recently reviewed in Ouwens et al. (201)], few studies analyze a cross talk between epicardial adipose tissue and cardiomyocytes. Secretory products from epicardial adipose tissue induces insulin resistance and defects in contractility and calcium influx in cardiomyocytes with the responsible adipokines still to be identified (87). Cardioprotective effects have been assigned to adiponectin (290), whereas resistin has been characterized as cardiodepressant (132).

### Conclusion

Enlarged adipose tissue releases a host of adipokines that play a key role in the interorgan cross talk between adipose tissue and the vasculature. Here we present a novel strategy to narrow down the huge number of potential biomarkers and drug targets by analyzing molecules that activate NF- $\kappa$ B pro-inflammatory signaling pathways. Comprehensive literature analysis resulted in a final list of known and novel adipokines that we suggest as a major candidates for future work. This should address the role of these molecules in linking obesity, chronic inflammation, and cardiovascular disease and their use as potential targets for diagnosis and treatment.

### GRANTS

This work was supported by the Ministerium für Wissenschaft und Forschung des Landes Nordrhein-Westfalen (Ministry of Science and Research of the State of North Rhine-Westphalia), the Bundesministerium für Gesundheit (Federal Ministry of Health), the Commission of the European Communities (Collaborative Project ADAPT, Contract No. HEALTH-F2-2008-201100), European Union COST Action BM0602, and the Jühling Foundation.

### DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the author(s).

### AUTHOR CONTRIBUTIONS

A.T., R.S., K.E., and J.E. analyzed data; A.T. and K.E. prepared figures; A.T., R.S., H.S., and J.E. drafted manuscript; A.T., R.S., H.S., K.E., and J.E.

edited and revised manuscript; K.E. and J.E. conception and design of research; K.E. and J.E. approved final version of manuscript.

### REFERENCES

- Adya R, Tan BK, Punj A, Chen J, Randeve HS. Visfatin induces human endothelial VEGF and MMP-2/9 production via MAPK and PI3K/Akt signalling pathways: novel insights into visfatin-induced angiogenesis. *Cardiovasc Res* 78: 356–365, 2008.
- Alan S, Ulgen MS, Ozturk O, Alan B, Ozdemir L, Toprak N. Relation between coronary artery disease, risk factors and intima-media thickness of carotid artery, arterial distensibility, and stiffness index. *Angiology* 54: 261–267, 2003.
- Alessi MC, Bastelica D, Morange P, Berthet B, Leduc I, Verdier M, Geel O, Juhan-Vague I. Plasminogen activator inhibitor 1, transforming growth factor-beta1, and BMI are closely associated in human adipose tissue during morbid obesity. *Diabetes* 49: 1374–1380, 2000.
- Alvarez-Llamas G, Szalowska E, de Vries MP, Weening D, Landman K, Hoek A, Wolffebuttel BH, Roelofsen H, Vonk RJ. Characterization of the human visceral adipose tissue secretome. *Mol Cell Proteomics* 6: 589–600, 2007.
- Alvehus M, Buren J, Sjoström M, Goedecke J, Olsson T. The human visceral fat depot has a unique inflammatory profile. *Obesity (Silver Spring)* 18: 879–883, 2010.
- Apostolakis S, Vogiatzi K, Amanatidou V, Spandidos DA. Interleukin 8 and cardiovascular disease. *Cardiovasc Res* 84: 353–360, 2009.
- Aragones G, Ferre R, Lazaro I, Cabre A, Plana N, Merino J, Heras M, Girona J, Masana L. Fatty acid-binding protein 4 is associated with endothelial dysfunction in patients with type 2 diabetes. *Atherosclerosis* 213: 329–331, 2010.
- Arita Y, Kihara S, Ouchi N, Maeda K, Kuriyama H, Okamoto Y, Kumada M, Hotta K, Nishida M, Takahashi M, Nakamura T, Shimomura I, Muraguchi M, Ohmoto Y, Funahashi T, Matsuzawa Y. Adipocyte-derived plasma protein adiponectin acts as a platelet-derived growth factor-BB-binding protein and regulates growth factor-induced common postreceptor signal in vascular smooth muscle cell. *Circulation* 105: 2893–2898, 2002.
- Arner P. Regional adiposity in man. *J Endocrinol* 155: 191–192, 1997.
- Arner P. Regional differences in protein production by human adipose tissue. *Biochem Soc Trans* 29: 72–75, 2001.
- Aust G, Richter O, Rohm S, Kerner C, Hauss J, Kloting N, Ruschke K, Kovacs P, Youn BS, Bluher M. Vaspin serum concentrations in patients with carotid stenosis. *Atherosclerosis* 204: 262–266, 2009.
- Bakker W, Eringa EC, Sipkema P, van Hinsbergh VW. Endothelial dysfunction and diabetes: roles of hyperglycemia, impaired insulin signaling and obesity. *Cell Tissue Res* 335: 165–189, 2009.
- Barandier C, Montani JP, Yang Z. Mature adipocytes and perivascular adipose tissue stimulate vascular smooth muscle cell proliferation: effects of aging and obesity. *Am J Physiol Heart Circ Physiol* 289: H1807–H1813, 2005.
- Barnes PJ, Karin M. Nuclear factor-kappaB: a pivotal transcription factor in chronic inflammatory diseases. *N Engl J Med* 336: 1066–1071, 1997.
- Bauche IB, El Mkaed SA, Pottier AM, Senou M, Many MC, Rezhohazy R, Penicaud L, Maeda N, Funahashi T, Brichard SM. Overexpression of adiponectin targeted to adipose tissue in transgenic mice: impaired adipocyte differentiation. *Endocrinology* 148: 1539–1549, 2007.
- Baudry A, Yang ZZ, Hemmings BA. PKBalpha is required for adipose differentiation of mouse embryonic fibroblasts. *J Cell Sci* 119: 889–897, 2006.
- Berndt J, Kloting N, Kralisch S, Kovacs P, Fasshauer M, Schon MR, Stumvoll M, Bluher M. Plasma visfatin concentrations and fat depot-specific mRNA expression in humans. *Diabetes* 54: 2911–2916, 2005.
- Bernhagen J, Krohn R, Lue H, Gregory JL, Zerneck A, Koenen RR, Dewor M, Georgiev I, Schober A, Leng L, Kooistra T, Fingerle-Rowson G, Ghezzi P, Kleemann R, McColl SR, Bucala R, Hickey MJ, Weber C. MIF is a noncognate ligand of CXC chemokine receptors in inflammatory and atherogenic cell recruitment. *Nat Med* 13: 587–596, 2007.
- Bigalke B, Stellos K, Geisler T, Seizer P, Mozes V, Gawaz M. High plasma levels of adipocytokines are associated with platelet activation in patients with coronary artery disease. *Platelets* 21: 11–19, 2010.

20. Blankenberg S, Tiret L, Bickel C, Peetz D, Cambien F, Meyer J, Rupprecht HJ. Interleukin-18 is a strong predictor of cardiovascular death in stable and unstable angina. *Circulation* 106: 24–30, 2002.
21. Boisvert WA, Curtiss LK, Terkeltaub RA. Interleukin-8 and its receptor CXCR2 in atherosclerosis. *Immunol Res* 21: 129–137, 2000.
22. Bokarewa M, Nagaev I, Dahlberg L, Smith U, Tarkowski A. Resistin, an adipokine with potent proinflammatory properties. *J Immunol* 174: 5789–5795, 2005.
23. Bosello O, Zamboni M. Visceral obesity and metabolic syndrome. *Obes Rev* 1: 47–56, 2000.
24. Bots ML, Dijk JM, Oren A, Grobbee DE. Carotid intima-media thickness, arterial stiffness and risk of cardiovascular disease: current evidence. *J Hypertens* 20: 2317–2325, 2002.
25. Bozaoglu K, Bolton K, McMillan J, Zimmet P, Jowett J, Collier G, Walder K, Segal D. Chemerin is a novel adipokine associated with obesity and metabolic syndrome. *Endocrinology* 148: 4687–4694, 2007.
26. Brennan AM, Li TY, Kelesidis I, Gavrilu A, Hu FB, Mantzoros CS. Circulating leptin levels are not associated with cardiovascular morbidity and mortality in women with diabetes: a prospective cohort study. *Diabetologia* 50: 1178–1185, 2007.
27. Britton KA, Fox CS. Perivascular adipose tissue and vascular disease. *Clin Lipidol* 6: 79–91, 2011.
28. Bruun JM, Lihn AS, Madan AK, Pedersen SB, Schiott KM, Fain JN, Richelsen B. Higher production of IL-8 in visceral vs. subcutaneous adipose tissue. Implication of nonadipose cells in adipose tissue. *Am J Physiol Endocrinol Metab* 286: E8–E13, 2004.
29. Bruun JM, Lihn AS, Pedersen SB, Richelsen B. Monocyte chemoattractant protein-1 release is higher in visceral than subcutaneous human adipose tissue (AT): implication of macrophages resident in the AT. *J Clin Endocrinol Metab* 90: 2282–2289, 2005.
30. Bu DX, Hemdahl AL, Gabrielsen A, Fuxe J, Zhu C, Eriksson P, Yan ZQ. Induction of neutrophil gelatinase-associated lipocalin in vascular injury via activation of nuclear factor-kappaB. *Am J Pathol* 169: 2245–2253, 2006.
31. Burger-Kentischer A, Goebel H, Seiler R, Fraedrich G, Schaefer HE, Dimmeler S, Kleemann R, Bernhagen J, Ihling C. Expression of macrophage migration inhibitory factor in different stages of human atherosclerosis. *Circulation* 105: 1561–1566, 2002.
32. Byrne AM, Bouchier-Hayes DJ, Harmey JH. Angiogenic and cell survival functions of vascular endothelial growth factor (VEGF). *J Cell Mol Med* 9: 777–794, 2005.
33. Calabro P, Samudio I, Willerson JT, Yeh ET. Resistin promotes smooth muscle cell proliferation through activation of extracellular signal-regulated kinase 1/2 and phosphatidylinositol 3-kinase pathways. *Circulation* 110: 3335–3340, 2004.
34. Catalan V, Gomez-Ambrosi J, Rodriguez A, Ramirez B, Silva C, Rotellar F, Gil MJ, Cienfuegos JA, Salvador J, Fruhbeck G. Increased adipose tissue expression of lipocalin-2 in obesity is related to inflammation and matrix metalloproteinase-2 and metalloproteinase-9 activities in humans. *J Mol Med (Berl)* 87: 803–813, 2009.
35. Cattaruzza M, Slodowski W, Stojakovic M, Krzesz R, Hecker M. Interleukin-10 induction of nitric-oxide synthase expression attenuates CD40-mediated interleukin-12 synthesis in human endothelial cells. *J Biol Chem* 278: 37874–37880, 2003.
36. Cavusoglu E, Marmur JD, Hojjati MR, Chopra V, Butala M, Subnani R, Huda MS, Yanamadala S, Ruwende C, Eng C, Pinsky DJ. Plasma interleukin-10 levels and adverse outcomes in acute coronary syndrome. *Am J Med* 124: 724–730, 2011.
37. Chandrasekar B, Mummidi S, Mahimainathan L, Patel DN, Bailey SR, Imam SZ, Greene WC, Valente AJ. Interleukin-18-induced human coronary artery smooth muscle cell migration is dependent on NF-kappaB- and AP-1-mediated matrix metalloproteinase-9 expression and is inhibited by atorvastatin. *J Biol Chem* 281: 15099–15109, 2006.
38. Chatterjee TK, Stoll LL, Denning GM, Harrelson A, Blomkalns AL, Idelman G, Rothenberg FG, Neltner B, Romig-Martin SA, Dickson EW, Rudich S, Weintraub NL. Proinflammatory phenotype of perivascular adipocytes: influence of high-fat feeding. *Circ Res* 104: 541–549, 2009.
39. Chen CC, Manning AM. TGF-beta 1, IL-10 and IL-4 differentially modulate the cytokine-induced expression of IL-6 and IL-8 in human endothelial cells. *Cytokine* 8: 58–65, 1996.
40. Chen X, Kelemen SE, Autieri MV. AIF-1 expression modulates proliferation of human vascular smooth muscle cells by autocrine expression of G-CSF. *Arterioscler Thromb Vasc Biol* 24: 1217–1222, 2004.
41. Chen Y, Budd RC, Kelm RJ Jr, Sobel BE, Schneider DJ. Augmentation of proliferation of vascular smooth muscle cells by plasminogen activator inhibitor type 1. *Arterioscler Thromb Vasc Biol* 26: 1777–1783, 2006.
42. Cheng KH, Chu CS, Lee KT, Lin TH, Hsieh CC, Chiu CC, Voon WC, Sheu SH, Lai WT. Adipocytokines and proinflammatory mediators from abdominal and epicardial adipose tissue in patients with coronary artery disease. *Int J Obes (Lond)* 32: 268–274, 2008.
43. Choi KM, Lee JS, Kim EJ, Baik SH, Seo HS, Choi DS, Oh DJ, Park CG. Implication of lipocalin-2 and visfatin levels in patients with coronary heart disease. *Eur J Endocrinol* 158: 203–207, 2008.
44. Choi SH, Kwak SH, Lee Y, Moon MK, Lim S, Park YJ, Jang HC, Kim MS. Plasma vaspin concentrations are elevated in metabolic syndrome in men and are correlated with coronary atherosclerosis in women. *Clin Endocrinol (Oxf)* 75: 628–635, 2011.
45. Csizsar A, Labinskyy N, Smith K, Rivera A, Orosz Z, Ungvari Z. Vasculoprotective effects of anti-tumor necrosis factor-alpha treatment in aging. *Am J Pathol* 170: 388–398, 2007.
46. Csizsar A, Ungvari Z, Koller A, Edwards JG, Kaley G. Proinflammatory phenotype of coronary arteries promotes endothelial apoptosis in aging. *Physiol Genomics* 17: 21–30, 2004.
47. Daemen MJ, Lombardi DM, Bosman FT, Schwartz SM. Angiotensin II induces smooth muscle cell proliferation in the normal and injured rat arterial wall. *Circ Res* 68: 450–456, 1991.
48. Dahl TB, Yndestad A, Skjelland M, Oie E, Dahl A, Michelsen A, Damas JK, Tunheim SH, Ueland T, Smith C, Bendz B, Tonstad S, Gullestad L, Froland SS, Krohg-Sorensen K, Russell D, Aukrust P, Halvorsen B. Increased expression of visfatin in macrophages of human unstable carotid and coronary atherosclerosis: possible role in inflammation and plaque destabilization. *Circulation* 115: 972–980, 2007.
49. Dahlof B. Cardiovascular disease risk factors: epidemiology and risk assessment. *Am J Cardiol* 105: 3A–9A, 2010.
50. Davenport P, Tipping PG. The role of interleukin-4 and interleukin-12 in the progression of atherosclerosis in apolipoprotein E-deficient mice. *Am J Pathol* 163: 1117–1125, 2003.
51. Dawson DW, Volpert OV, Gillis P, Crawford SE, Xu H, Benedict W, Bouck NP. Pigment epithelium-derived factor: a potent inhibitor of angiogenesis. *Science* 285: 245–248, 1999.
52. Deng G, Long Y, Yu YR, Li MR. Adiponectin directly improves endothelial dysfunction in obese rats through the AMPK-eNOS pathway. *Int J Obes (Lond)* 34: 165–171, 2010.
53. Deng Y, Scherer PE. Adipokines as novel biomarkers and regulators of the metabolic syndrome. *Ann NY Acad Sci* 1212: E1–E19, 2010.
54. Diebold I, Djordjevic T, Hess J, Gorlach A. Rac-1 promotes pulmonary artery smooth muscle cell proliferation by upregulation of plasminogen activator inhibitor-1: role of NFkappaB-dependent hypoxia-inducible factor-1alpha transcription. *Thromb Haemost* 100: 1021–1028, 2008.
55. Doll JA, Stellmach VM, Bouck NP, Bergh AR, Lee C, Abramson LP, Cornwell ML, Pins MR, Borensztajn J, Crawford SE. Pigment epithelium-derived factor regulates the vasculature and mass of the prostate and pancreas. *Nat Med* 9: 774–780, 2003.
56. Duh EJ, Yang HS, Suzuma I, Miyagi M, Youngman E, Mori K, Katai M, Yan L, Suzuma K, West K, Davarya S, Tong P, Gehlbach P, Pearlman J, Crabb JW, Aiello LP, Campochiaro PA, Zack DJ. Pigment epithelium-derived factor suppresses ischemia-induced retinal neovascularization and VEGF-induced migration and growth. *Invest Ophthalmol Vis Sci* 43: 821–829, 2002.
57. Elhage R, Jawien J, Rudling M, Ljunggren HG, Takeda K, Akira S, Bayard F, Hansson GK. Reduced atherosclerosis in interleukin-18 deficient apolipoprotein E-knockout mice. *Cardiovasc Res* 59: 234–240, 2003.
58. Elmasri H, Karaaslan C, Teper Y, Ghelfi E, Weng M, Ince TA, Kozakewich H, Bischoff J, Cataltepe S. Fatty acid binding protein 4 is a target of VEGF and a regulator of cell proliferation in endothelial cells. *FASEB J* 23: 3865–3873, 2009.
59. Eriksson P, van H, V, Hoffstedt J, Lundquist P, Vidal H, Stemme V, Hamsten A, Arner P, Reynisdottir S. Regional variation in plasminogen activator inhibitor-1 expression in adipose tissue from obese individuals. *Thromb Haemost* 83: 545–548, 2000.
60. Esteve E, Ricart W, Fernandez-Real JM. Adipocytokines and insulin resistance: the possible role of lipocalin-2, retinol binding protein-4, and adiponectin. *Diabetes Care* 32, Suppl 2: S362–S367, 2009.

61. Faggiotto A, Ross R, Harker L. Studies of hypercholesterolemia in the nonhuman primate. I. Changes that lead to fatty streak formation. *Arteriosclerosis* 4: 323–340, 1984.
62. Fain JN, Madan AK, Hiler ML, Cheema P, Bahouth SW. Comparison of the release of adipokines by adipose tissue, adipose tissue matrix, and adipocytes from visceral and subcutaneous abdominal adipose tissues of obese humans. *Endocrinology* 145: 2273–2282, 2004.
63. Fain JN, Tagele BM, Cheema P, Madan AK, Tichansky DS. Release of 12 adipokines by adipose tissue, nonfat cells, and fat cells from obese women. *Obesity (Silver Spring)* 18: 890–896, 2010.
64. Famulla S, Lamers D, Hartwig S, Passlack W, Horrhigs A, Cramer A, Lehr S, Sell H, Eckel J. Pigment epithelium-derived factor is one of the most abundant proteins secreted by human adipocytes and induces insulin resistance and inflammatory signaling in muscle and fat cells. *Int J Obes (Lond)* 35: 762–772, 2011.
65. Fargnoli JL, Sun Q, Olenczuk D, Qi L, Zhu Y, Hu FB, Mantzoros CS. Resistin is associated with biomarkers of inflammation whereas total and high-molecular weight adiponectin are associated with biomarkers of inflammation, insulin resistance, and endothelial function. *Eur J Endocrinol* 162: 281–288, 2010.
66. Ferrara N, Davis-Smyth T. The biology of vascular endothelial growth factor. *Endocr Rev* 18: 4–25, 1997.
67. Festa A, D'Agostino R Jr, Williams K, Karter AJ, Mayer-Davis EJ, Tracy RP, Haffner SM. The relation of body fat mass and distribution to markers of chronic inflammation. *Int J Obes Relat Metab Disord* 25: 1407–1415, 2001.
68. Fisher FM, McTernan PG, Valsamakis G, Chetty R, Harte AL, Anwar AJ, Starcynski J, Crocker J, Barnett AH, McTernan CL, Kumar S. Differences in adiponectin protein expression: effect of fat depots and type 2 diabetic status. *Horm Metab Res* 34: 650–654, 2002.
69. Fitzgibbons TP, Kogan S, Aouadi M, Hendricks GM, Straubhaar J, Czech MP. Similarity of mouse perivascular and brown adipose tissues and their resistance to diet-induced inflammation. *Am J Physiol Heart Circ Physiol* 301: H1425–H1437, 2011.
70. Flaster H, Bernhagen J, Calandra T, Bucala R. The macrophage migration inhibitory factor-glucocorticoid dyad: regulation of inflammation and immunity. *Mol Endocrinol* 21: 1267–1280, 2007.
71. Flower DR. The lipocalin protein family: structure and function. *Biochem J* 318: 1–14, 1996.
72. Frangogiannis NG, Ren G, Dewald O, Zymek P, Haudek S, Koerting A, Winkelmann K, Michael LH, Lawler J, Entman ML. Critical role of endogenous thrombospondin-1 in preventing expansion of healing myocardial infarcts. *Circulation* 111: 2935–2942, 2005.
73. Fried SK, Bunkin DA, Greenberg AS. Omental and subcutaneous adipose tissues of obese subjects release interleukin-6: depot difference and regulation by glucocorticoid. *J Clin Endocrinol Metab* 83: 847–850, 1998.
74. Frostegard J, Ulfgren AK, Nyberg P, Hedin U, Swedenborg J, Andersson U, Hansson GK. Cytokine expression in advanced human atherosclerotic plaques: dominance of pro-inflammatory (Th1) and macrophage-stimulating cytokines. *Atherosclerosis* 145: 33–43, 1999.
75. Funaki M. Saturated fatty acids and insulin resistance. *J Med Invest* 56: 88–92, 2009.
76. Galea J, Armstrong J, Gadsdon P, Holden H, Francis SE, Holt CM. Interleukin-1 beta in coronary arteries of patients with ischemic heart disease. *Arterioscler Thromb Vasc Biol* 16: 1000–1006, 1996.
77. Garten A, Petzold S, Korner A, Imai S, Kiess W. Nampt: linking NAD biology, metabolism and cancer. *Trends Endocrinol Metab* 20: 130–138, 2009.
78. Gealekman O, Guseva N, Hartigan C, Apotheker S, Gorgoglione M, Gurav K, Tran KV, Straubhaar J, Nicoloso S, Czech MP, Thompson M, Perugini RA, Corvera S. Depot-specific differences and insufficient subcutaneous adipose tissue angiogenesis in human obesity. *Circulation* 123: 186–194, 2011.
79. Geisel J, Jodden V, Obeid R, Knapp JP, Bodis M, Herrmann W. Stimulatory effect of homocysteine on interleukin-8 expression in human endothelial cells. *Clin Chem Lab Med* 41: 1045–1048, 2003.
80. George J, Mulkins M, Shaish A, Casey S, Schatzman R, Sigal E, Harats D. Interleukin (IL)-4 deficiency does not influence fatty streak formation in C57BL/6 mice. *Atherosclerosis* 153: 403–411, 2000.
81. Gerszten RE, Garcia-Zepeda EA, Lim YC, Yoshida M, Ding HA, Gimbrone MA Jr, Luster AD, Luscinskas FW, Rosenzweig A. MCP-1 and IL-8 trigger firm adhesion of monocytes to vascular endothelium under flow conditions. *Nature* 398: 718–723, 1999.
82. Goetze S, Kintscher U, Kaneshiro K, Meehan WP, Collins A, Fleck E, Hsueh WA, Law RE. TNF $\alpha$  induces expression of transcription factors c-fos, Egr-1, and Ets-1 in vascular lesions through extracellular signal-regulated kinases 1/2. *Atherosclerosis* 159: 93–101, 2001.
83. Goetze S, Xi XP, Kawano Y, Kawano H, Fleck E, Hsueh WA, Law RE. TNF- $\alpha$ -induced migration of vascular smooth muscle cells is MAPK dependent. *Hypertension* 33: 183–189, 1999.
84. Goralski KB, McCarthy TC, Hanniman EA, Zabel BA, Butcher EC, Parlee SD, Muruganandan S, Sinal CJ. Chemerin, a novel adipokine that regulates adipogenesis and adipocyte metabolism. *J Biol Chem* 282: 28175–28188, 2007.
85. Gossli M, Modder UI, Gulati R, Rihal CS, Prasad A, Loeffler D, Lerman LO, Khosla S, Lerman A. Coronary endothelial dysfunction in humans is associated with coronary retention of osteogenic endothelial progenitor cells. *Eur Heart J* 31: 2909–2914, 2010.
86. Greenstein AS, Khavandi K, Withers SB, Sonoyama K, Clancy O, Jeziorska M, Laing I, Yates AP, Pemberton PW, Malik RA, Heagerty AM. Local inflammation and hypoxia abolish the protective anticontractile properties of perivascular fat in obese patients. *Circulation* 119: 1661–1670, 2009.
87. Greulich S, de Wiza DH, Preilowski S, Ding Z, Mueller H, Langin D, Jaquet K, Ouwens DM, Eckel J. Secretory products of guinea pig epicardial fat induce insulin resistance and impair primary adult rat cardiomyocyte function. *J Cell Mol Med* 15: 2399–2410, 2011.
88. Gualillo O, Gonzalez-Juanatey JR, Lago F. The emerging role of adipokines as mediators of cardiovascular function: physiologic and clinical perspectives. *Trends Cardiovasc Med* 17: 275–283, 2007.
89. Hajer GR, van Haeften TW, Visseren FL. Adipose tissue dysfunction in obesity, diabetes, and vascular diseases. *Eur Heart J* 29: 2959–2971, 2008.
90. Halaas JL, Gajiwala KS, Maffei M, Cohen SL, Chait BT, Rabinowitz D, Lallone RL, Burley SK, Friedman JM. Weight-reducing effects of the plasma protein encoded by the obese gene. *Science* 269: 543–546, 1995.
91. Han X, Kitamoto S, Lian Q, Boisvert WA. Interleukin-10 facilitates both cholesterol uptake and efflux in macrophages. *J Biol Chem* 284: 32950–32958, 2009.
92. Hansson GK, Hermansson A. The immune system in atherosclerosis. *Nat Immunol* 12: 204–212, 2011.
93. Hart R, Greaves DR. Chemerin contributes to inflammation by promoting macrophage adhesion to VCAM-1 and fibronectin through clustering of VLA-4 and VLA-5. *J Immunol* 185: 3728–3739, 2010.
94. Hastings NE, Feaver RE, Lee MY, Wamhoff BR, Blackman BR. Human IL-8 regulates smooth muscle cell VCAM-1 expression in response to endothelial cells exposed to atheroprone flow. *Arterioscler Thromb Vasc Biol* 29: 725–731, 2009.
95. Hasty AH, Shimano H, Osuga J, Namatame I, Takahashi A, Yahagi N, Perrey S, Iizuka Y, Tamura Y, Amemiya-Kudo M, Yoshikawa T, Okazaki H, Ohashi K, Harada K, Matsuzaka T, Sone H, Gotoda T, Nagai R, Ishibashi S, Yamada N. Severe hypercholesterolemia, hypertriglyceridemia, and atherosclerosis in mice lacking both leptin and the low density lipoprotein receptor. *J Biol Chem* 276: 37402–37408, 2001.
96. Hattori Y, Matsumura M, Kasai K. Vascular smooth muscle cell activation by C-reactive protein. *Cardiovasc Res* 58: 186–195, 2003.
97. Hattori Y, Suzuki M, Hattori S, Kasai K. Globular adiponectin upregulates nitric oxide production in vascular endothelial cells. *Diabetologia* 46: 1543–1549, 2003.
98. Hayes IM, Jordan NJ, Towers S, Smith G, Paterson JR, Earnshaw JJ, Roach AG, Westwick J, Williams RJ. Human vascular smooth muscle cells express receptors for CC chemokines. *Arterioscler Thromb Vasc Biol* 18: 397–403, 1998.
99. Henrichot E, Juge-Aubry CE, Pernin A, Pache JC, Velebit V, Dayer JM, Meda P, Chizzolini C, Meier CA. Production of chemokines by perivascular adipose tissue: a role in the pathogenesis of atherosclerosis? *Arterioscler Thromb Vasc Biol* 25: 2594–2599, 2005.
100. Herder C, Baumert J, Thorand B, Martin S, Lowel H, Kolb H, Koenig W. Chemokines and incident coronary heart disease: results from the MONICA/KORA Augsburg case-cohort study, 1984–2002. *Arterioscler Thromb Vasc Biol* 26: 2147–2152, 2006.
101. Hida K, Wada J, Eguchi J, Zhang H, Baba M, Seida A, Hashimoto I, Okada T, Yasuhara A, Nakatsuka A, Shikata K, Hourai S, Futami J, Watanabe E, Matsuki Y, Hiramatsu R, Akagi S, Makino H, Kanwar YS. Visceral adipose tissue-derived serine protease inhibitor: a

- unique insulin-sensitizing adipocytokine in obesity. *Proc Natl Acad Sci USA* 102: 10610–10615, 2005.
102. **Hirsch J, Batchelor B.** Adipose tissue cellularity in human obesity. *Clin Endocrinol Metab* 5: 299–311, 1976.
  103. **Holm PW, Slart RH, Zeebregts CJ, Hillebrands JL, Tio RA.** Atherosclerotic plaque development and instability: a dual role for VEGF. *Ann Med* 41: 257–264, 2009.
  104. **Hotamisligil GS, Arner P, Caro JF, Atkinson RL, Spiegelman BM.** Increased adipose tissue expression of tumor necrosis factor- $\alpha$  in human obesity and insulin resistance. *J Clin Invest* 95: 2409–2415, 1995.
  105. **Hotamisligil GS, Johnson RS, Distel RJ, Ellis R, Papaioannou VE, Spiegelman BM.** Uncoupling of obesity from insulin resistance through a targeted mutation in aP2, the adipocyte fatty acid binding protein. *Science* 274: 1377–1379, 1996.
  106. **Hsu BG, Chen YC, Lee RP, Lee CC, Lee CJ, Wang JH.** Fasting serum level of fatty-acid-binding protein 4 positively correlates with metabolic syndrome in patients with coronary artery disease. *Circ J* 74: 327–331, 2010.
  107. **Hu E, Liang P, Spiegelman BM.** AdipoQ is a novel adipose-specific gene dysregulated in obesity. *J Biol Chem* 271: 10697–10703, 1996.
  108. **Huang F, Xiong X, Wang H, You S, Zeng H.** Leptin-induced vascular smooth muscle cell proliferation via regulating cell cycle, activating ERK1/2 and NF- $\kappa$ B. *Acta Biochim Biophys Sin (Shanghai)* 42: 325–331, 2010.
  109. **Hubert HB, Feinleib M, McNamara PM, Castelli WP.** Obesity as an independent risk factor for cardiovascular disease: a 26-year follow-up of participants in the Framingham Heart Study. *Circulation* 67: 968–977, 1983.
  110. **Huo Y, Ley K.** Adhesion molecules and atherogenesis. *Acta Physiol Scand* 173: 35–43, 2001.
  111. **Ingelsson E, Lind L.** Circulating retinol-binding protein 4 and subclinical cardiovascular disease in the elderly. *Diabetes Care* 32: 733–735, 2009.
  112. **Inoue T, Komoda H, Nonaka M, Kameda M, Uchida T, Node K.** Interleukin-8 as an independent predictor of long-term clinical outcome in patients with coronary artery disease. *Int J Cardiol* 124: 319–325, 2008.
  113. **Janssen I, Katzmarzyk PT, Ross R.** Body mass index, waist circumference, and health risk: evidence in support of current National Institutes of Health guidelines. *Arch Intern Med* 162: 2074–2079, 2002.
  114. **Jenkins A, Zhang SX, Gosmanova A, Aston C, Dashti A, Baker MZ, Lyons T, Ma JX.** Increased serum pigment epithelium derived factor levels in Type 2 diabetes patients. *Diabetes Res Clin Pract* 82: e5–e7, 2008.
  115. **Jha HC, Divya A, Prasad J, Mittal A.** Plasma circulatory markers in male and female patients with coronary artery disease. *Heart Lung* 39: 296–303, 2010.
  116. **Jha HC, Srivastava P, Sarkar R, Prasad J, Mittal AS.** Association of plasma circulatory markers, Chlamydia pneumoniae, and high sensitive C-reactive protein in coronary artery disease patients of India. *Mediators Inflamm* 2009: 561532, 2009.
  117. **Jovinge S, Hamsten A, Tornvall P, Proudler A, Bavenholm P, Ericsson CG, Godstrand I, de FU, Nilsson J.** Evidence for a role of tumor necrosis factor  $\alpha$  in disturbances of triglyceride and glucose metabolism predisposing to coronary heart disease. *Metabolism* 47: 113–118, 1998.
  118. **Jovinge S, Hultgardh-Nilsson A, Regnstrom J, Nilsson J.** Tumor necrosis factor- $\alpha$  activates smooth muscle cell migration in culture and is expressed in the balloon-injured rat aorta. *Arterioscler Thromb Vasc Biol* 17: 490–497, 1997.
  119. **Jung CH, Lee WJ, Hwang JY, Seol SM, Kim YM, Lee YL, Park JY.** Vaspin protects vascular endothelial cells against free fatty acid-induced apoptosis through a phosphatidylinositol 3-kinase/Akt pathway. *Biochem Biophys Res Commun* 413: 264–269, 2011.
  120. **Jung HS, Park KH, Cho YM, Chung SS, Cho HJ, Cho SY, Kim SJ, Kim SY, Lee HK, Park KS.** Resistin is secreted from macrophages in atheromas and promotes atherosclerosis. *Cardiovasc Res* 69: 76–85, 2006.
  121. **Kamari Y, Werman-Venkert R, Shaish A, Werman A, Harari A, Gonen A, Voronov E, Grosskopf I, Sharabi Y, Grossman E, Iwakura Y, Dinarello CA, Apte RN, Harats D.** Differential role and tissue specificity of interleukin-1 $\alpha$  gene expression in atherogenesis and lipid metabolism. *Atherosclerosis* 195: 31–38, 2007.
  122. **Karakas M, Zierer A, Herder C, Baumert J, Meisinger C, Koenig W, Thorand B.** Leptin, adiponectin, their ratio and risk of Coronary Heart Disease: results from the MONICA/KORA Augsburg Study 1984–2002. *Atherosclerosis* 209: 220–225, 2010.
  123. **Karalis KP, Giannogonas P, Kodela E, Koutmani Y, Zoumakis M, Teli T.** Mechanisms of obesity and related pathology: linking immune responses to metabolic stress. *FEBS J* 276: 5747–5754, 2009.
  124. **Kato K, Matsubara T, Iida K, Suzuki O, Sato Y.** Elevated levels of pro-inflammatory cytokines in coronary artery thrombi. *Int J Cardiol* 70: 267–273, 1999.
  125. **Kaur J, Adya R, Tan BK, Chen J, Randeve HS.** Identification of chemerin receptor (ChemR23) in human endothelial cells: chemerin-induced endothelial angiogenesis. *Biochem Biophys Res Commun* 391: 1762–1768, 2010.
  126. **Ketonen J, Shi J, Martonen E, Mervaala E.** Periadventitial adipose tissue promotes endothelial dysfunction via oxidative stress in diet-induced obese C57Bl/6 mice. *Circ J* 74: 1479–1487, 2010.
  127. **Khurana R, Simons M, Martin JF, Zachary IC.** Role of angiogenesis in cardiovascular disease: a critical appraisal. *Circulation* 112: 1813–1824, 2005.
  128. **Kim HJ, Kim MY, Hwang JS, Kim HJ, Lee JH, Chang KC, Kim JH, Han CW, Kim JH, Seo HG.** PPAR $\delta$  inhibits IL-1 $\beta$ -stimulated proliferation and migration of vascular smooth muscle cells via up-regulation of IL-1Ra. *Cell Mol Life Sci* 67: 2119–2130, 2010.
  129. **Kim I, Moon SO, Park SK, Chae SW, Koh GY.** Angiopoietin-1 reduces VEGF-stimulated leukocyte adhesion to endothelial cells by reducing ICAM-1, VCAM-1, and E-selectin expression. *Circ Res* 89: 477–479, 2001.
  130. **Kim J, Choi YS, Lim S, Yea K, Yoon JH, Jun DJ, Ha SH, Kim JW, Kim JH, Suh PG, Ryu SH, Lee TG.** Comparative analysis of the secretory proteome of human adipose stromal vascular fraction cells during adipogenesis. *Proteomics* 10: 394–405, 2010.
  131. **Kim JE, Song SE, Kim YW, Kim JY, Park SC, Park YK, Baek SH, Lee IK, Park SY.** Adiponectin inhibits palmitate-induced apoptosis through suppression of reactive oxygen species in endothelial cells: involvement of cAMP/protein kinase A and AMP-activated protein kinase. *J Endocrinol* 207: 35–44, 2010.
  132. **Kim M, Oh JK, Sakata S, Liang I, Park W, Hajjar RJ, Lebeche D.** Role of resistin in cardiac contractility and hypertrophy. *J Mol Cell Cardiol* 45: 270–280, 2008.
  133. **Kirbis S, Breskvar UD, Sabovic M, Zupan I, Sinkovic A.** Inflammation markers in patients with coronary artery disease—comparison of intracoronary and systemic levels. *Wien Klin Wochenschr* 122, Suppl 2: 31–34, 2010.
  134. **Kirih H, Niwa T, Yamada Y, Wada H, Saito K, Iwakura Y, Asano M, Moriwaki H, Seishima M.** Lack of interleukin-1 $\beta$  decreases the severity of atherosclerosis in ApoE-deficient mice. *Arterioscler Thromb Vasc Biol* 23: 656–660, 2003.
  135. **Kloting N, Berndt J, Kralisch S, Kovacs P, Fasshauer M, Schon MR, Stumvoll M, Bluher M.** Vaspin gene expression in human adipose tissue: association with obesity and type 2 diabetes. *Biochem Biophys Res Commun* 339: 430–436, 2006.
  136. **Korshunov VA, Nikonenko TA, Tkachuk VA, Brooks A, Berk BC.** Interleukin-18 and macrophage migration inhibitory factor are associated with increased carotid intima-media thickening. *Arterioscler Thromb Vasc Biol* 26: 295–300, 2006.
  137. **Kouri FM, Queisser MA, Konigshoff M, Chrobak I, Preissner KT, Seeger W, Eickelberg O.** Plasminogen activator inhibitor type 1 inhibits smooth muscle cell proliferation in pulmonary arterial hypertension. *Int J Biochem Cell Biol* 40: 1872–1882, 2008.
  138. **Kubota N, Terauchi Y, Yamauchi T, Kubota T, Moroi M, Matsui J, Eto K, Yamashita T, Kamon J, Satoh H, Yano W, Froguel P, Nagai R, Kimura S, Kadowaki T, Noda T.** Disruption of adiponectin causes insulin resistance and neointimal formation. *J Biol Chem* 277: 25863–25866, 2002.
  139. **Kumada M, Kihara S, Sumitsuji S, Kawamoto T, Matsumoto S, Ouchi N, Arita Y, Okamoto Y, Shimomura I, Hiraoka H, Nakamura T, Funahashi T, Matsuzawa Y.** Association of hypo-adiponectinemia with coronary artery disease in men. *Arterioscler Thromb Vasc Biol* 23: 85–89, 2003.
  140. **Kunnari AM, Savolainen ER, Ukkola OH, Kesaniemi YA, Jokela MA.** The expression of human resistin in different leucocyte lineages is modulated by LPS and TNF $\alpha$ . *Regul Pept* 157: 57–63, 2009.

141. **Lago F, Dieguez C, Gomez-Reino J, Gualillo O.** Adipokines as emerging mediators of immune response and inflammation. *Nat Clin Pract Rheumatol* 3: 716–724, 2007.
142. **Lai Y, Shen Y, Liu XH, Zhang Y, Zeng Y, Liu YF.** Interleukin-8 induces the endothelial cell migration through the activation of phosphoinositide 3-kinase-Rac1/RhoA pathway. *Int J Biol Sci* 7: 782–791, 2011.
143. **Lamers D, Famulla S, Wronkowitz N, Hartwig S, Lehr S, Ouwers DM, Eckardt K, Kaufman JM, Ryden M, Muller S, Hanisch FG, Ruige J, Arner P, Sell H, Eckel J.** Dipeptidyl peptidase 4 is a novel adipokine potentially linking obesity to the metabolic syndrome. *Diabetes* 60:1917–1925, 2011.
144. **Lamers D, Schlich R, Greulich S, Sasson S, Sell H, Eckel J.** Oleic acid and adipokines synergize in inducing proliferation and inflammatory signaling in human vascular smooth muscle cells. *J Cell Mol Med* 15: 1177–1188, 2011.
145. **Larsson B, Bjorntorp P, Tibblin G.** The health consequences of moderate obesity. *Int J Obes* 5: 97–116, 1981.
146. **Lee MY, Li H, Xiao Y, Zhou Z, Xu A, Vanhoutte PM.** Chronic administration of BMS309403 improves endothelial function in apolipoprotein E-deficient mice and in cultured human endothelial cells. *Br J Pharmacol* 162: 1564–1576, 2011.
147. **Lee T, Nesselroth SM, Olson ET, Esemuede N, Lawler J, Sumpio BE, Gahtan V.** Thrombospondin-1-induced vascular smooth muscle cell chemotaxis: the role of the type 3 repeat and carboxyl terminal domains. *J Cell Biochem* 89: 500–506, 2003.
148. **Lee YW, Kuhn H, Hennig B, Toborek M.** IL-4 induces apoptosis of endothelial cells through the caspase-3-dependent pathway. *FEBS Lett* 485: 122–126, 2000.
149. **Lee YW, Lee WH, Kim PH.** Oxidative mechanisms of IL-4-induced IL-6 expression in vascular endothelium. *Cytokine* 49: 73–79, 2010.
150. **Lehman SJ, Massaro JM, Schlett CL, O'Donnell CJ, Hoffmann U, Fox CS.** Peri-aortic fat, cardiovascular disease risk factors, and aortic calcification: the Framingham Heart Study. *Atherosclerosis* 210: 656–661, 2010.
151. **Lehrke M, Becker A, Greif M, Stark R, Laubender RP, von ZF, Leberherz C, Tittus J, Reiser M, Becker C, Goke B, Leber AW, Parhofer KG, Broedl UC.** Chemerin is associated with markers of inflammation and components of the metabolic syndrome but does not predict coronary atherosclerosis. *Eur J Endocrinol* 161: 339–344, 2009.
152. **Li FY, Cheng KK, Lam KS, Vanhoutte PM, Xu A.** Cross-talk between adipose tissue and vasculature: role of adiponectin. *Acta Physiol (Oxf)* 203: 167–180, 2011.
153. **Li L, Mamputu JC, Wiernsperger N, Renier G.** Signaling pathways involved in human vascular smooth muscle cell proliferation and matrix metalloproteinase-2 expression induced by leptin: inhibitory effect of metformin. *Diabetes* 54: 2227–2234, 2005.
154. **Libby P, Okamoto Y, Rocha VZ, Folco E.** Inflammation in atherosclerosis: transition from theory to practice. *Circ J* 74: 213–220, 2010.
155. **Lihn AS, Bruun JM, He G, Pedersen SB, Jensen PF, Richelsen B.** Lower expression of adiponectin mRNA in visceral adipose tissue in lean and obese subjects. *Mol Cell Endocrinol* 219: 9–15, 2004.
156. **Lim JH, Um HJ, Park JW, Lee IK, Kwon TK.** Interleukin-1 $\beta$  promotes the expression of monocyte chemoattractant protein-1 in human aorta smooth muscle cells via multiple signaling pathways. *Exp Mol Med* 41: 757–764, 2009.
157. **Lin Y, Rajala MW, Berger JP, Moller DE, Barzilay N, Scherer PE.** Hyperglycemia-induced production of acute phase reactants in adipose tissue. *J Biol Chem* 276: 42077–42083, 2001.
158. **Liu JT, Song E, Xu A, Berger T, Mak TW, Tse HF, Law IK, Huang B, Liang Y, Vanhoutte PM, Wang Y.** Lipocalin-2 deficiency prevents endothelial dysfunction associated with dietary obesity: role of cytochrome P450 2C inhibition. *Br J Pharmacol* 165: 520–531, 2012.
159. **Lloyd-Jones DM.** Cardiovascular risk prediction: basic concepts, current status, and future directions. *Circulation* 121: 1768–1777, 2010.
160. **Lofgren I, Herron K, Zern T, West K, Patalay M, Schachter NS, Koo SI, Fernandez ML.** Waist circumference is a better predictor than body mass index of coronary heart disease risk in overweight premenopausal women. *J Nutr* 134: 1071–1076, 2004.
161. **Lovren F, Pan Y, Shukla PC, Quan A, Teoh H, Szmítko PE, Peterson MD, Gupta M, Al-Omran M, Verma S.** Visfatin activates eNOS via Akt and MAP kinases and improves endothelial cell function and angiogenesis in vitro and in vivo: translational implications for atherosclerosis. *Am J Physiol Endocrinol Metab* 296: E1440–E1449, 2009.
162. **Luo Y, D'Amore PA, Dorf ME.** Beta-chemokine TCA3 binds to and activates rat vascular smooth muscle cells. *J Immunol* 157: 2143–2148, 1996.
163. **Madani R, Karastergiou K, Ogston NC, Miheisi N, Bhome R, Haloob N, Tan GD, Karpe F, Malone-Lee J, Hashemi M, Jahangiri M, Mohamed-Ali V.** RANTES release by human adipose tissue in vivo and evidence for depot-specific differences. *Am J Physiol Endocrinol Metab* 296: E1262–E1268, 2009.
164. **Maffei M, Halaas J, Ravussin E, Pratley RE, Lee GH, Zhang Y, Fei H, Kim S, Lallone R, Ranganathan S.** Leptin levels in human and rodent: measurement of plasma leptin and ob RNA in obese and weight-reduced subjects. *Nat Med* 1: 1155–1161, 1995.
165. **Mahadev K, Wu X, Donnelly S, Ouedraogo R, Eckhart AD, Goldstein BJ.** Adiponectin inhibits vascular endothelial growth factor-induced migration of human coronary artery endothelial cells. *Cardiovasc Res* 78: 376–384, 2008.
166. **Makino A, Nakamura T, Hirano M, Kitta Y, Sano K, Kobayashi T, Fujioka D, Saito Y, Watanabe K, Watanabe Y, Kawabata K, Obata JE, Kugiyama K.** High plasma levels of macrophage migration inhibitory factor are associated with adverse long-term outcome in patients with stable coronary artery disease and impaired glucose tolerance or type 2 diabetes mellitus. *Atherosclerosis* 213: 573–578, 2010.
167. **Makowski L, Hotamisligil GS.** Fatty acid binding proteins—the evolutionary crossroads of inflammatory and metabolic responses. *J Nutr* 134: 2464S–2468S, 2004.
168. **Mallat Z, Besnard S, Duriez M, Deleuze V, Emmanuel F, Bureau MF, Soubrier F, Esposito B, Duez H, Fievet C, Staels B, Duverger N, Scherman D, Tedgui A.** Protective role of interleukin-10 in atherosclerosis. *Circ Res* 85: e17–e24, 1999.
169. **Mallat Z, Corbaz A, Scoazec A, Besnard S, Leseche G, Chvatchko Y, Tedgui A.** Expression of interleukin-18 in human atherosclerotic plaques and relation to plaque instability. *Circulation* 104: 1598–1603, 2001.
170. **Manduteanu I, Pirvulescu M, Gan AM, Stan D, Simion V, Dragomir E, Calin M, Manea A, Simionescu M.** Similar effects of resistin and high glucose on P-selectin and fractalkine expression and monocyte adhesion in human endothelial cells. *Biochem Biophys Res Commun* 391: 1443–1448, 2010.
171. **Mange H, Schauenstein K, Stroedter L, Griesl A, Maerz W, Borkenstein M.** Low grade inflammation in juvenile obesity and type 1 diabetes associated with early signs of atherosclerosis. *Exp Clin Endocrinol Diabetes* 112: 378–382, 2004.
172. **Manna SK, Ramesh GT.** Interleukin-8 induces nuclear transcription factor-kappaB through a TRAF6-dependent pathway. *J Biol Chem* 280: 7010–7021, 2005.
173. **Marchesi C, Paradis P, Schiffrin EL.** Role of the renin-angiotensin system in vascular inflammation. *Trends Pharmacol Sci* 29: 367–374, 2008.
174. **Martin T, Cardarelli PM, Parry GC, Felts KA, Cobb RR.** Cytokine induction of monocyte chemoattractant protein-1 gene expression in human endothelial cells depends on the cooperative action of NF-kappa B and AP-1. *Eur J Immunol* 27: 1091–1097, 1997.
175. **Marumo T, Schini-Kerth VB, Busse R.** Vascular endothelial growth factor activates nuclear factor-kappaB and induces monocyte chemoattractant protein-1 in bovine retinal endothelial cells. *Diabetes* 48: 1131–1137, 1999.
176. **Matsuzawa Y, Funahashi T, Kihara S, Shimomura I.** Adiponectin and metabolic syndrome. *Arterioscler Thromb Vasc Biol* 24: 29–33, 2004.
177. **Matter CM, Handschin C.** RANTES (regulated on activation, normal T cell expressed and secreted), inflammation, obesity, and the metabolic syndrome. *Circulation* 115: 946–948, 2007.
178. **Mazighi M, Pelle A, Gonzalez W, Mtairag eM, Philippe M, Henin D, Michel JB, Feldman LJ.** IL-10 inhibits vascular smooth muscle cell activation in vitro and in vivo. *Am J Physiol Heart Circ Physiol* 287: H866–H871, 2004.
179. **McTernan PG, McTernan CL, Chetty R, Jenner K, Fisher FM, Lauer MN, Crocker J, Barnett AH, Kumar S.** Increased resistin gene and protein expression in human abdominal adipose tissue. *J Clin Endocrinol Metab* 87: 2407, 2002.
180. **Miao CY, Li ZY.** The role of perivascular adipose tissue in vascular smooth muscle cell growth. *Br J Pharmacol* 165: 643–658, 2012.
181. **Milner KL, van der Poorten D, Xu A, Bugianesi E, Kench JG, Lam KS, Chisholm DJ, George J.** Adipocyte fatty acid binding protein levels relate to inflammation and fibrosis in nonalcoholic fatty liver disease. *Hepatology* 49: 1926–1934, 2009.

182. Minocci A, Savia G, Lucantoni R, Berselli ME, Tagliaferri M, Calo G, Petroni ML, de Medici C, Viberti GC, Liuzzi A. Leptin plasma concentrations are dependent on body fat distribution in obese patients. *Int J Obes Relat Metab Disord* 24: 1139–1144, 2000.
183. Mizia-Stec K, Gasior Z, Zahorska-Markiewicz B, Janowska J, Szulc A, Jastrzebska-Maj E, Kobielski-Gembala I. Serum tumour necrosis factor- $\alpha$ , interleukin-2 and interleukin-10 activation in stable angina and acute coronary syndromes. *Coron Artery Dis* 14: 431–438, 2003.
184. Morand EF, Leech M, Bernhagen J. MIF: a new cytokine link between rheumatoid arthritis and atherosclerosis. *Nat Rev Drug Discov* 5: 399–410, 2006.
185. Moschen AR, Kaser A, Enrich B, Mosheimer B, Theurl M, Niederegger H, Tilg H. Visfatin, an adipocytokine with proinflammatory and immunomodulating properties. *J Immunol* 178: 1748–1758, 2007.
186. Nakamura K, Yamagishi S, Matsui T, Yoshida T, Takenaka K, Jinnouchi Y, Yoshida Y, Ueda S, Adachi H, Imaizumi T. Pigment epithelium-derived factor inhibits neointimal hyperplasia after vascular injury by blocking NADPH oxidase-mediated reactive oxygen species generation. *Am J Pathol* 170: 2159–2170, 2007.
187. Nakamura Y, Shimada K, Fukuda D, Shimada Y, Ehara S, Hirose M, Kataoka T, Kamimori K, Shimodozono S, Kobayashi Y, Yoshiyama M, Takeuchi K, Yoshikawa J. Implications of plasma concentrations of adiponectin in patients with coronary artery disease. *Heart* 90: 528–533, 2004.
188. Narizhneva NV, Byers-Ward VJ, Quinn MJ, Zidar FJ, Plow EF, Topol EJ, Byzova TV. Molecular and functional differences induced in thrombospondin-1 by the single nucleotide polymorphism associated with the risk of premature, familial myocardial infarction. *J Biol Chem* 279: 21651–21657, 2004.
189. Narizhneva NV, Razorenova OV, Podrez EA, Chen J, Chandrasekharan UM, DiCorleto PE, Plow EF, Topol EJ, Byzova TV. Thrombospondin-1 up-regulates expression of cell adhesion molecules and promotes monocyte binding to endothelium. *FASEB J* 19: 1158–1160, 2005.
190. Natarajan R, Rosdahl J, Gonzales N, Bai W. Regulation of 12-lipoxygenase by cytokines in vascular smooth muscle cells. *Hypertension* 30: 873–879, 1997.
191. Nesselroth SM, Willis AI, Fuse S, Olson ET, Lawler J, Sumpio BE, Gahtan V. The C-terminal domain of thrombospondin-1 induces vascular smooth muscle cell chemotaxis. *J Vasc Surg* 33: 595–600, 2001.
192. Odamaki M, Furuya R, Kinumura Y, Ikegaya N, Kumagai H. Association between plasma adiponectin concentration and visceral fat accumulation in hemodialysis patients. *Nephron Clin Pract* 102: c8–c13, 2006.
193. Okada T, Tsukano H, Endo M, Tabata M, Miyata K, Kadomatsu T, Miyashita K, Semba K, Nakamura E, Tsukano M, Mizuta H, Oike Y. Synovocyte-derived angiopoietin-like protein 2 contributes to synovial chronic inflammation in rheumatoid arthritis. *Am J Pathol* 176: 2309–2319, 2010.
194. Otabe S, Yuan X, Fukutani T, Wada N, Hashinaga T, Nakayama H, Hirota N, Kojima M, Yamada K. Overexpression of human adiponectin in transgenic mice results in suppression of fat accumulation and prevention of premature death by high-calorie diet. *Am J Physiol Endocrinol Metab* 293: E210–E218, 2007.
195. Ouchi N, Higuchi A, Ohashi K, Oshima Y, Gokce N, Shibata R, Akasaki Y, Shimono A, Walsh K. Sfrp5 is an anti-inflammatory adipokine that modulates metabolic dysfunction in obesity. *Science* 329: 454–457, 2010.
196. Ouchi N, Kihara S, Arita Y, Maeda K, Kuriyama H, Okamoto Y, Hotta K, Nishida M, Takahashi M, Nakamura T, Yamashita S, Funahashi T, Matsuzawa Y. Novel modulator for endothelial adhesion molecules: adipocyte-derived plasma protein adiponectin. *Circulation* 100: 2473–2476, 1999.
197. Ouchi N, Kihara S, Arita Y, Okamoto Y, Maeda K, Kuriyama H, Hotta K, Nishida M, Takahashi M, Muraguchi M, Ohmoto Y, Nakamura T, Yamashita S, Funahashi T, Matsuzawa Y. Adiponectin, an adipocyte-derived plasma protein, inhibits endothelial NF- $\kappa$ B signaling through a cAMP-dependent pathway. *Circulation* 102: 1296–1301, 2000.
198. Ouchi N, Kihara S, Funahashi T, Matsuzawa Y, Walsh K. Obesity, adiponectin and vascular inflammatory disease. *Curr Opin Lipidol* 14: 561–566, 2003.
199. Ouchi N, Parker JL, Lugus JJ, Walsh K. Adipokines in inflammation and metabolic disease. *Nat Rev Immunol* 11: 85–97, 2011.
200. Ouedraogo R, Gong Y, Berzins B, Wu X, Mahadev K, Hough K, Chan L, Goldstein BJ, Scalia R. Adiponectin deficiency increases leukocyte-endothelium interactions via upregulation of endothelial cell adhesion molecules in vivo. *J Clin Invest* 117: 1718–1726, 2007.
201. Ouwens DM, Sell H, Greulich S, Eckel J. The role of epicardial and perivascular adipose tissue in the pathophysiology of cardiovascular disease. *J Cell Mol Med* 14: 2223–2234, 2010.
202. Ozeren A, Aydin M, Tokac M, Demircan N, Unalacak M, Gurel A, Yazici M. Levels of serum IL-1 $\beta$ , IL-2, IL-8 and tumor necrosis factor- $\alpha$  in patients with unstable angina pectoris. *Mediators Inflamm* 12: 361–365, 2003.
203. Packard RR, Libby P. Inflammation in atherosclerosis: from vascular biology to biomarker discovery and risk prediction. *Clin Chem* 54: 24–38, 2008.
204. Pakala R, Watanabe T, Benedict CR. Induction of endothelial cell proliferation by angiogenic factors released by activated monocytes. *Cardiovasc Radiat Med* 3: 95–101, 2002.
205. Pan JH, Sukhova GK, Yang JT, Wang B, Xie T, Fu H, Zhang Y, Satoskar AR, David JR, Metz CN, Bucala R, Fang K, Simon DI, Chapman HA, Libby P, Shi GP. Macrophage migration inhibitory factor deficiency impairs atherosclerosis in low-density lipoprotein receptor-deficient mice. *Circulation* 109: 3149–3153, 2004.
206. Payne GA, Borbouse L, Kumar S, Neeb Z, Alloosh M, Sturek M, Tune JD. Epicardial perivascular adipose-derived leptin exacerbates coronary endothelial dysfunction in metabolic syndrome via a protein kinase C- $\beta$  pathway. *Arterioscler Thromb Vasc Biol* 30: 1711–1717, 2010.
207. Pena AS, Belobrajdic DP, Wiltshire E, Gent R, Hirte C, Couper J. Adiponectin relates to smooth muscle function and folate in obese children. *Int J Pediatr Obes* 5: 185–191, 2010.
208. Phalitakul S, Okada M, Hara Y, Yamawaki H. Vaspin prevents TNF- $\alpha$ -induced intracellular adhesion molecule-1 via inhibiting reactive oxygen species-dependent NF- $\kappa$ B and PKC $\theta$  activation in cultured rat vascular smooth muscle cells. *Pharmacol Res* 64: 493–500, 2011.
209. Pinderski Oslund LJ, Hedrick CC, Olvera T, Hagenbaugh A, Territo M, Berliner JA, Fyfe AI. Interleukin-10 blocks atherosclerotic events in vitro and in vivo. *Arterioscler Thromb Vasc Biol* 19: 2847–2853, 1999.
210. Pischon T, Girman CJ, Hotamisligil GS, Rifai N, Hu FB, Rimm EB. Plasma adiponectin levels and risk of myocardial infarction in men. *JAMA* 291: 1730–1737, 2004.
211. Police SB, Thatcher SE, Charnigo R, Daugherty A, Cassis LA. Obesity promotes inflammation in periaortic adipose tissue and angiotensin II-induced abdominal aortic aneurysm formation. *Arterioscler Thromb Vasc Biol* 29: 1458–1464, 2009.
212. Qian HS, Gu JM, Liu P, Kauser K, Halks-Miller M, Vergona R, Sullivan ME, Dole WP, Deng GG. Overexpression of PAI-1 prevents the development of abdominal aortic aneurysm in mice. *Gene Ther* 15: 224–232, 2008.
213. Ramanjaneya M, Chen J, Brown JE, Tripathi G, Hallschmid M, Patel S, Kern W, Hillhouse EW, Lehnert H, Tan BK, Randeve HS. Identification of nesfatin-1 in human and murine adipose tissue: a novel depot-specific adipokine with increased levels in obesity. *Endocrinology* 151: 3169–3180, 2010.
214. Ramis JM, Franssen-van Hal NL, Kramer E, Llado I, Bouillaud F, Palou A, Keijer J. Carboxypeptidase E and thrombospondin-1 are differentially expressed in subcutaneous and visceral fat of obese subjects. *Cell Mol Life Sci* 59: 1960–1971, 2002.
215. Rastogi S, Rizwani W, Joshi B, Kunigal S, Chellappan SP. TNF- $\alpha$  response of vascular endothelial and vascular smooth muscle cells involve differential utilization of ASK1 kinase and p73. *Cell Death Differ* 19: 274–283, 2012.
216. Rattarasarn C, Leelawattana R, Soonthornpun S, Setasuban W, Thamprasit A, Lim A, Chayanunnukul W, Thamkumpee N, Daendurongsub T. Regional abdominal fat distribution in lean and obese Thai type 2 diabetic women: relationships with insulin sensitivity and cardiovascular risk factors. *Metabolism* 52: 1444–1447, 2003.
217. Reddy VS, Valente AJ, Delafontaine P, Chandrasekar B. Interleukin-18/WNT1-inducible signaling pathway protein-1 signaling mediates human saphenous vein smooth muscle cell proliferation. *J Cell Physiol* 226: 3303–3315, 2011.
218. Reilly MP, Lehrke M, Wolfe ML, Rohatgi A, Lazar MA, Rader DJ. Resistin is an inflammatory marker of atherosclerosis in humans. *Circulation* 111: 932–939, 2005.

219. **Rexrode KM, Hennekens CH, Willett WC, Colditz GA, Stampfer MJ, Rich-Edwards JW, Speizer FE, Manson JE.** A prospective study of body mass index, weight change, and risk of stroke in women. *JAMA* 277: 1539–1545, 1997.
220. **Rhee EJ, Lee WY, Park CY, Oh KW, Kim BJ, Sung KC, Kim BS.** The association of serum adipocyte fatty acid-binding protein with coronary artery disease in Korean adults. *Eur J Endocrinol* 160: 165–172, 2009.
221. **Rocha VZ, Libby P.** Obesity, inflammation, and atherosclerosis. *Nat Rev Cardiol* 6: 399–409, 2009.
222. **Rodriguez A, Fortuno A, Gomez-Ambrosi J, Zalba G, Diez J, Fruhbeck G.** The inhibitory effect of leptin on angiotensin II-induced vasoconstriction in vascular smooth muscle cells is mediated via a nitric oxide-dependent mechanism. *Endocrinology* 148: 324–331, 2007.
223. **Roh SG, Song SH, Choi KC, Katoh K, Wittamer V, Parmentier M, Sasaki S.** Chemerin—a new adipokine that modulates adipogenesis via its own receptor. *Biochem Biophys Res Commun* 362: 1013–1018, 2007.
224. **Rollins BJ, Pober JS.** Interleukin-4 induces the synthesis and secretion of MCP-1/JE by human endothelial cells. *Am J Pathol* 138: 1315–1319, 1991.
225. **Romacho T, Azcutia V, Vazquez-Bella M, Matesanz N, Cercas E, Nevado J, Carraro R, Rodriguez-Manas L, Sanchez-Ferrer CF, Peiro C.** Extracellular PBEF/NAMPT/visfatin activates pro-inflammatory signalling in human vascular smooth muscle cells through nicotinamide phosphoribosyltransferase activity. *Diabetologia* 52: 2455–2463, 2009.
226. **Rosenow A, Arrey TN, Bouwman FG, Noben JP, Wabitsch M, Mariman EC, Karas M, Renes J.** Identification of novel human adipocyte secreted proteins by using SGBS cells. *J Proteome Res* 9: 5389–5401, 2010.
227. **Ross R.** The pathogenesis of atherosclerosis—an update. *N Engl J Med* 314: 488–500, 1986.
228. **Ross R.** The pathogenesis of atherosclerosis: a perspective for the 1990s. *Nature* 362: 801–809, 1993.
229. **Ross R, Dagnone D, Jones PJ, Smith H, Paddags A, Hudson R, Janssen I.** Reduction in obesity and related comorbid conditions after diet-induced weight loss or exercise-induced weight loss in men. A randomized, controlled trial. *Ann Intern Med* 133: 92–103, 2000.
230. **Sahar S, Dwarakanath RS, Reddy MA, Lanting L, Todorov I, Natarajan R.** Angiotensin II enhances interleukin-18 mediated inflammatory gene expression in vascular smooth muscle cells: a novel cross-talk in the pathogenesis of atherosclerosis. *Circ Res* 96: 1064–1071, 2005.
231. **Salgado-Somoza A, Teixeira-Fernandez E, Fernandez AL, Gonzalez-Juanatey JR, Eiras S.** Proteomic analysis of epicardial and subcutaneous adipose tissue reveals differences in proteins involved in oxidative stress. *Am J Physiol Heart Circ Physiol* 299: H202–H209, 2010.
232. **Salminen A, Kaarniranta K.** Control of p53 and NF- $\kappa$ B signaling by WIP1 and MIF: role in cellular senescence and organismal aging. *Cell Signal* 23: 747–752, 2011.
233. **Samaras K, Botelho NK, Chisholm DJ, Lord RV.** Subcutaneous and visceral adipose tissue gene expression of serum adipokines that predict type 2 diabetes. *Obesity (Silver Spring)* 18: 884–889, 2010.
234. **Sasu S, Beasley D.** Essential roles of I $\kappa$ B kinase  $\alpha$  and  $\beta$  in serum- and IL-1-induced human VSMC proliferation. *Am J Physiol Heart Circ Physiol* 278: H1823–H1831, 2000.
235. **Savage DB, Sewter CP, Klenk ES, Segal DG, Vidal-Puig A, Considine RV, O'Rahilly S.** Resistin / Fizz3 expression in relation to obesity and peroxisome proliferator-activated receptor- $\gamma$  action in humans. *Diabetes* 50: 2199–2202, 2001.
236. **Schiffrin EL, Touyz RM.** Inflammation and vascular hypertrophy induced by angiotensin II: role of NADPH oxidase-derived reactive oxygen species independently of blood pressure elevation? *Arterioscler Thromb Vasc Biol* 23: 707–709, 2003.
237. **Schlett CL, Massaro JM, Lehman SJ, Bamberg F, O'Donnell CJ, Fox CS, Hoffmann U.** Novel measurements of periaortic adipose tissue in comparison to anthropometric measures of obesity, and abdominal adipose tissue. *Int J Obes (Lond)* 33: 226–232, 2009.
238. **Schmeisser A, Marquetant R, Illmer T, Graffy C, Garlich CD, Bockler D, Menschikowski D, Braun-Dullaeus R, Daniel WG, Strasser RH.** The expression of macrophage migration inhibitory factor 1 $\alpha$  (MIF 1 $\alpha$ ) in human atherosclerotic plaques is induced by different proatherogenic stimuli and associated with plaque instability. *Atherosclerosis* 178: 83–94, 2005.
239. **Schmidt MD, Dwyer T, Magnussen CG, Venn AJ.** Predictive associations between alternative measures of childhood adiposity and adult cardio-metabolic health. *Int J Obes (Lond)* 35: 38–45, 2011.
240. **Schrans-Stassen BH, Lue H, Sonnemans DG, Bernhagen J, Post MJ.** Stimulation of vascular smooth muscle cell migration by macrophage migration inhibitory factor. *Antioxid Redox Signal* 7: 1211–1216, 2005.
241. **Schwartz SM, Campbell GR, Campbell JH.** Replication of smooth muscle cells in vascular disease. *Circ Res* 58: 427–444, 1986.
242. **Sell H, Laurencikienė J, Taube A, Eckardt K, Cramer A, Horrigs A, Arner P, Eckel J.** Chemerin is a novel adipocyte-derived factor inducing insulin resistance in primary human skeletal muscle cells. *Diabetes* 58: 2731–2740, 2009.
243. **Selzman CH, McIntyre RC Jr, Shames BD, Whitehill TA, Banerjee A, Harken AH.** Interleukin-10 inhibits human vascular smooth muscle proliferation. *J Mol Cell Cardiol* 30: 889–896, 1998.
244. **Shimizu K, Minami M, Shubiki R, Lopez-Illasaca M, MacFarlane L, Asami Y, Li Y, Mitchell RN, Libby P.** CC chemokine receptor-1 activates intimal smooth muscle-like cells in graft arterial disease. *Circulation* 120: 1800–1813, 2009.
245. **Shimomura I, Funahashi T, Takahashi M, Maeda K, Kotani K, Nakamura T, Yamashita S, Miura M, Fukuda Y, Takemura K, Tokunaga K, Matsuzawa Y.** Enhanced expression of PAI-1 in visceral fat: possible contributor to vascular disease in obesity. *Nat Med* 2: 800–803, 1996.
246. **Sierra-Johnson J, Romero-Corral A, Lopez-Jimenez F, Gami AS, Sert Kuniyoshi FH, Wolk R, Somers VK.** Relation of increased leptin concentrations to history of myocardial infarction and stroke in the United States population. *Am J Cardiol* 100: 234–239, 2007.
247. **Silswal N, Singh AK, Aruna B, Mukhopadhyay S, Ghosh S, Ehtesham NZ.** Human resistin stimulates the pro-inflammatory cytokines TNF- $\alpha$  and IL-12 in macrophages by NF- $\kappa$ B-dependent pathway. *Biochem Biophys Res Commun* 334: 1092–1101, 2005.
248. **Simonini A, Moscucci M, Muller DW, Bates ER, Pagani FD, Burdick MD, Strieter RM.** IL-8 is an angiogenic factor in human coronary atherosclerotic tissue. *Circulation* 101: 1519–1526, 2000.
249. **Singh P, Hoffmann M, Wolk R, Shamsuzzaman AS, Somers VK.** Leptin induces C-reactive protein expression in vascular endothelial cells. *Arterioscler Thromb Vasc Biol* 27: e302–e307, 2007.
250. **Singh P, Peterson TE, Barber KR, Kuniyoshi FS, Jensen A, Hoffmann M, Shamsuzzaman AS, Somers VK.** Leptin upregulates the expression of plasminogen activator inhibitor-1 in human vascular endothelial cells. *Biochem Biophys Res Commun* 392: 47–52, 2010.
251. **Singh RJ, Mason JC, Lidington EA, Edwards DR, Nuttall RK, Khokha R, Knauper V, Murphy G, Gavrilovic J.** Cytokine stimulated vascular cell adhesion molecule-1 (VCAM-1) ectodomain release is regulated by TIMP-3. *Cardiovasc Res* 67: 39–49, 2005.
252. **Skoog T, Dichtl W, Boquist S, Skoglund-Andersson C, Karpe F, Tang R, Bond MG, de FU, Nilsson J, Eriksson P, Hamsten A.** Plasma tumour necrosis factor- $\alpha$  and early carotid atherosclerosis in healthy middle-aged men. *Eur Heart J* 23: 376–383, 2002.
253. **Skurk T, Alberti-Huber C, Herder C, Hauner H.** Relationship between adipocyte size and adipokine expression and secretion. *J Clin Endocrinol Metab* 92: 1023–1033, 2007.
254. **Skurk T, Herder C, Kraft I, Muller-Scholz S, Hauner H, Kolb H.** Production and release of macrophage migration inhibitory factor from human adipocytes. *Endocrinology* 146: 1006–1011, 2005.
255. **Sluijter JC, Daemen MJ.** Novel concepts in atherogenesis: angiogenesis and hypoxia in atherosclerosis. *J Pathol* 218: 7–29, 2009.
256. **Smith DA, Irving SD, Sheldon J, Cole D, Kaski JC.** Serum levels of the anti-inflammatory cytokine interleukin-10 are decreased in patients with unstable angina. *Circulation* 104: 746–749, 2001.
257. **Spiroglou SG, Kostopoulos CG, Varakis JN, Papadaki HH.** Adipokines in periaortic and epicardial adipose tissue: differential expression and relation to atherosclerosis. *J Atheroscler Thromb* 17: 115–130, 2010.
258. **Staiger H, Tschritter O, Machann J, Thamer C, Fritsche A, Maerker E, Schick F, Haring HU, Stumvoll M.** Relationship of serum adiponectin and leptin concentrations with body fat distribution in humans. *Obes Res* 11: 368–372, 2003.
259. **Stejskal D, Karpisek M, Hanulova Z, Svestak M.** Chemerin is an independent marker of the metabolic syndrome in a Caucasian population—a pilot study. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub* 152: 217–221, 2008.
260. **Stenina OI, Plow EF.** Counterbalancing forces: what is thrombospondin-1 doing in atherosclerotic lesions? *Circ Res* 103: 1053–1055, 2008.



261. Steppan CM, Bailey ST, Bhat S, Brown EJ, Banerjee RR, Wright CM, Patel HR, Ahima RS, Lazar MA. The hormone resistin links obesity to diabetes. *Nature* 409: 307–312, 2001.
262. Steppich BA, Demetz G, Schulz S, von Wedel J, Pogatsa-Murray G, Braun SL, Stein A, Kastrati A, Schomig A, Ott I. Effects of G-CSF on systemic inflammation, coagulation and platelet activation in patients with acute myocardial infarction. *Thromb Res* 127: 119–121, 2011.
263. Stintzing S, Ocker M, Hartner A, Amann K, Barbera L, Neureiter D. Differentiation patterning of vascular smooth muscle cells (VSMC) in atherosclerosis. *Virchows Arch* 455: 171–185, 2009.
264. Stofkova A. Leptin and adiponectin: from energy and metabolic dysbalance to inflammation and autoimmunity. *Endocr Regul* 43: 157–168, 2009.
265. Streit M, Velasco P, Riccardi L, Spencer L, Brown LF, Janes L, Lange-Asschenfeldt B, Yano K, Hawighorst T, Iruela-Arispe L, Detmar M. Thrombospondin-1 suppresses wound healing and granulation tissue formation in the skin of transgenic mice. *EMBO J* 19: 3272–3282, 2000.
266. Sweeney G. Cardiovascular effects of leptin. *Nat Rev Cardiol* 7: 22–29, 2010.
267. Szekecz Z, Shah MR, Harlow LA, Pearce WH, Koch AE. Interleukin-8 and tumor necrosis factor- $\alpha$  are involved in human aortic endothelial cell migration. The possible role of these cytokines in human aortic aneurysmal blood vessel growth. *Pathobiology* 62: 134–139, 1994.
268. Tahara N, Yamagishi S, Tahara A, Nitta Y, Kodama N, Mizoguchi M, Mohar D, Ishibashi M, Hayabuchi N, Imaizumi T. Serum level of pigment epithelium-derived factor is a marker of atherosclerosis in humans. *Atherosclerosis* 219: 311–315, 2011.
269. Takahashi M, Takahashi Y, Takahashi K, Zolotaryov FN, Hong KS, Kitazawa R, Iida K, Okimura Y, Kaji H, Kitazawa S, Kasuga M, Chihara K. Chemerin enhances insulin signaling and potentiates insulin-stimulated glucose uptake in 3T3-L1 adipocytes. *FEBS Lett* 582: 573–578, 2008.
270. Takaoka M, Nagata D, Kihara S, Shimomura I, Kimura Y, Tabata Y, Saito Y, Nagai R, Sata M. Periadventitial adipose tissue plays a critical role in vascular remodeling. *Circ Res* 105: 906–911, 2009.
271. Tan BK, Adya R, Chen J, Farhatullah S, Heutling D, Mitchell D, Lehnert H, Randeve HS. Metformin decreases angiogenesis via NF- $\kappa$ B and Erk1/2/Erk5 pathways by increasing the antiangiogenic thrombospondin-1. *Cardiovasc Res* 83: 566–574, 2009.
272. Tan BK, Chen J, Farhatullah S, Adya R, Kaur J, Heutling D, Lewandowski KC, O'Hare JP, Lehnert H, Randeve HS. Insulin and metformin regulate circulating and adipose tissue chemerin. *Diabetes* 58: 1971–1977, 2009.
273. te Boekhorst BC, Bovens SM, Hellings WE, van der Kraak PH, van de Kolk KW, Vink A, Moll FL, van Oosterhout MF, de Vries JP, Doevendans PA, Goumans MJ, de Kleijn DP, van Echteld CJ, Pasterkamp G, Sluijter JP. Molecular MRI of murine atherosclerotic plaque targeting NGAL: a protein associated with unstable human plaque characteristics. *Cardiovasc Res* 89: 680–688, 2011.
274. Toi M, Harris AL, Bicknell R. Interleukin-4 is a potent mitogen for capillary endothelium. *Biochem Biophys Res Commun* 174: 1287–1293, 1991.
275. Trayhurn P, Beattie JH. Physiological role of adipose tissue: white adipose tissue as an endocrine and secretory organ. *Proc Nutr Soc* 60: 329–339, 2001.
276. Trayhurn P, Wood IS. Signalling role of adipose tissue: adipokines and inflammation in obesity. *Biochem Soc Trans* 33: 1078–1081, 2005.
277. Tsai S, Hollenbeck ST, Ryer EJ, Edlin R, Yamanouchi D, Kundi R, Wang C, Liu B, Kent KC. TGF- $\beta$  through Smad3 signaling stimulates vascular smooth muscle cell proliferation and neointimal formation. *Am J Physiol Heart Circ Physiol* 297: H540–H549, 2009.
278. Tso AW, Xu A, Chow WS, Lam KS. Adipose tissue and the metabolic syndrome: focusing on adiponectin and several novel adipokines. *Biomark Med* 2: 239–252, 2008.
279. Tummala PE, Chen XL, Sundell CL, Laursen JB, Hammes CP, Alexander RW, Harrison DG, Medford RM. Angiotensin II induces vascular cell adhesion molecule-1 expression in rat vasculature: A potential link between the renin-angiotensin system and atherosclerosis. *Circulation* 100: 1223–1229, 1999.
280. Uyemura K, Demer LL, Castle SC, Jullien D, Berliner JA, Gately MK, Warriar RR, Pham N, Fogelman AM, Modlin RL. Cross-regulatory roles of interleukin (IL)-12 and IL-10 in atherosclerosis. *J Clin Invest* 97: 2130–2138, 1996.
281. Uysal KT, Scheja L, Wiesbrock SM, Bonner-Weir S, Hotamisligil GS. Improved glucose and lipid metabolism in genetically obese mice lacking ap2. *Endocrinology* 141: 3388–3396, 2000.
282. Vadiveloo PK, Stanton HR, Cochran FW, Hamilton JA. Interleukin-4 inhibits human smooth muscle cell proliferation. *Artery* 21: 161–181, 1994.
283. Van der Heiden K, Cuhlmann S, Luong IA, Zakkar M, Evans PC. Role of nuclear factor kappaB in cardiovascular health and disease. *Clin Sci (Lond)* 118: 593–605, 2010.
284. van d V, Nong Z, O'Neil C, Urquhart B, Freeman D, Pickering JG. Pre-B-cell colony-enhancing factor regulates NAD<sup>+</sup>-dependent protein deacetylase activity and promotes vascular smooth muscle cell maturation. *Circ Res* 97: 25–34, 2005.
285. Vecchione C, Maffei A, Colella S, Aretini A, Poulet R, Frati G, Gentile MT, Fratta L, Trimarco V, Trimarco B, Lembo G. Leptin effect on endothelial nitric oxide is mediated through Akt-endothelial nitric oxide synthase phosphorylation pathway. *Diabetes* 51: 168–173, 2002.
286. Verma S, Li SH, Wang CH, Fedak PW, Li RK, Weisel RD, Mickle DA. Resistin promotes endothelial cell activation: further evidence of adipokine-endothelial interaction. *Circulation* 108: 736–740, 2003.
287. Vidal H. Gene expression in visceral and subcutaneous adipose tissues. *Ann Med* 33: 547–555, 2001.
288. Wajchenberg BL. Subcutaneous and visceral adipose tissue: their relation to the metabolic syndrome. *Endocr Rev* 21: 697–738, 2000.
289. Wallace AM, McMahon AD, Packard CJ, Kelly A, Shepherd J, Gaw A, Sattar N. Plasma leptin and the risk of cardiovascular disease in the west of Scotland coronary prevention study (WOSCOPS). *Circulation* 104: 3052–3056, 2001.
290. Walsh K. Adipokines, myokines and cardiovascular disease. *Circ J* 73: 13–18, 2009.
291. Wang P, Smit E, Brouwers MC, Goossens GH, van der Kallen CJ, van Greevenbroek MM, Mariman EC. Plasma pigment epithelium-derived factor is positively associated with obesity in Caucasian subjects, in particular with the visceral fat depot. *Eur J Endocrinol* 159: 713–718, 2008.
292. Wang P, Xu TY, Guan YF, Su DF, Fan GR, Miao CY. Perivascular adipose tissue-derived visfatin is a vascular smooth muscle cell growth factor: role of nicotinamide mononucleotide. *Cardiovasc Res* 81: 370–380, 2009.
293. Wang SH, Liang CJ, Wu JC, Huang JJ, Chien HF, Tsai JS, Yen YS, Tseng YC, Lue JH, Chen YL. Pigment epithelium-derived factor reduces the PDGF-induced migration and proliferation of human aortic smooth muscle cells through PPAR $\gamma$  activation. *Int J Biochem Cell Biol* 44: 280–289, 2012.
294. Wang W, Chen HJ, Giedd KN, Schwartz A, Cannon PJ, Rabbani LE. T-cell lymphokines, interleukin-4 and gamma interferon, modulate the induction of vascular smooth muscle cell tissue plasminogen activator and migration by serum and platelet-derived growth factor. *Circ Res* 77: 1095–1106, 1995.
295. Wang X, Feuerstein GZ, Clark RK, Yue TL. Enhanced leucocyte adhesion to interleukin-1 beta stimulated vascular smooth muscle cells is mainly through intercellular adhesion molecule-1. *Cardiovasc Res* 28: 1808–1814, 1994.
296. Wang X, Feuerstein GZ, Gu JL, Lysko PG, Yue TL. Interleukin-1 beta induces expression of adhesion molecules in human vascular smooth muscle cells and enhances adhesion of leukocytes to smooth muscle cells. *Atherosclerosis* 115: 89–98, 1995.
297. Wang Y, Lam KS, Kraegen EW, Sweeney G, Zhang J, Tso AW, Chow WS, Wat NM, Xu JY, Hoo RL, Xu A. Lipocalin-2 is an inflammatory marker closely associated with obesity, insulin resistance, and hyperglycemia in humans. *Clin Chem* 53: 34–41, 2007.
298. Wang Z, Castresana MR, Newman WH. NF- $\kappa$ B is required for TNF- $\alpha$ -directed smooth muscle cell migration. *FEBS Lett* 508: 360–364, 2001.
299. Wang Z, Kong L, Kang J, Vaughn DM, Bush GD, Walding AL, Grigorian AA, Robinson JS Jr, Nakayama DK. Interleukin-1 $\beta$  induces migration of rat arterial smooth muscle cells through a mechanism involving increased matrix metalloproteinase-2 activity. *J Surg Res* 169: 328–336, 2011.
300. Wang Z, Rao PJ, Castresana MR, Newman WH. TNF- $\alpha$  induces proliferation or apoptosis in human saphenous vein smooth muscle cells depending on phenotype. *Am J Physiol Heart Circ Physiol* 288: H293–H301, 2005.

301. Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL, Ferrante AW Jr. Obesity is associated with macrophage accumulation in adipose tissue. *J Clin Invest* 112: 1796–1808, 2003.
302. Welsh P, Murray HM, Buckley BM, de Craen AJ, Ford I, Jukema JW, Macfarlane PW, Packard CJ, Stott DJ, Westendorp RG, Shepherd J, Sattar N. Leptin predicts diabetes but not cardiovascular disease: results from a large prospective study in an elderly population. *Diabetes Care* 32: 308–310, 2009.
303. Withers SB, Agabiti-Rosei C, Livingstone DM, Little MC, Aslam R, Malik RA, Heagerty AM. Macrophage activation is responsible for loss of anticontractile function in inflamed perivascular fat. *Arterioscler Thromb Vasc Biol* 31: 908–913, 2011.
304. Wolf G, Hamann A, Han DC, Helmchen U, Thaiss F, Ziyadeh FN, Stahl RA. Leptin stimulates proliferation and TGF-beta expression in renal glomerular endothelial cells: potential role in glomerulosclerosis. *Kidney Int* 56: 860–872, 1999.
305. Wolk R, Berger P, Lennon RJ, Brilakis ES, Johnson BD, Somers VK. Plasma leptin and prognosis in patients with established coronary atherosclerosis. *J Am Coll Cardiol* 44: 1819–1824, 2004.
306. Wood IS, de Heredia FP, Wang B, Trayhurn P. Cellular hypoxia and adipose tissue dysfunction in obesity. *Proc Nutr Soc* 68: 370–377, 2009.
307. Wright PS, Cooper JR, Kropp KE, Busch SJ. Induction of vascular cell adhesion molecule-1 expression by IL-4 in human aortic smooth muscle cells is not associated with increased nuclear NF-kappaB levels. *J Cell Physiol* 180: 381–389, 1999.
308. Wulster-Radcliffe MC, Ajuwon KM, Wang J, Christian JA, Spurlock ME. Adiponectin differentially regulates cytokines in porcine macrophages. *Biochem Biophys Res Commun* 316: 924–929, 2004.
309. Xiao N, Yin M, Zhang L, Qu X, Du H, Sun X, Mao L, Ren G, Zhang C, Geng Y, An L, Pan J. Tumor necrosis factor-alpha deficiency retards early fatty-streak lesion by influencing the expression of inflammatory factors in apoE-null mice. *Mol Genet Metab* 96: 239–244, 2009.
310. Xiao X, Dong Y, Zhong J, Cao R, Zhao X, Wen G, Liu J. Adiponectin protects endothelial cells from the damages induced by the intermittent high level of glucose. *Endocrine* 40: 386–393, 2011.
311. Xu A, Tso AW, Cheung BM, Wang Y, Wat NM, Fong CH, Yeung DC, Janus ED, Sham PC, Lam KS. Circulating adipocyte-fatty acid binding protein levels predict the development of the metabolic syndrome: a 5-year prospective study. *Circulation* 115: 1537–1543, 2007.
312. Xu A, Wang Y, Xu JY, Stejskal D, Tam S, Zhang J, Wat NM, Wong WK, Lam KS. Adipocyte fatty acid-binding protein is a plasma biomarker closely associated with obesity and metabolic syndrome. *Clin Chem* 52: 405–413, 2006.
313. Xu H, Barnes GT, Yang Q, Tan G, Yang D, Chou CJ, Sole J, Nichols A, Ross JS, Tartaglia LA, Chen H. Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. *J Clin Invest* 112: 1821–1830, 2003.
314. Xu S, Zhi H, Hou X, Jiang B. Angiotensin II modulates interleukin-1beta-induced inflammatory gene expression in vascular smooth muscle cells via interfering with ERK-NF-kappaB crosstalk. *Biochem Biophys Res Commun* 410: 543–548, 2011.
315. Yamagami H, Kitagawa K, Hoshi T, Furukado S, Hougaku H, Nagai Y, Hori M. Associations of serum IL-18 levels with carotid intima-media thickness. *Arterioscler Thromb Vasc Biol* 25: 1458–1462, 2005.
316. Yamagishi S, Adachi H, Abe A, Yashiro T, Enomoto M, Furuki K, Hino A, Jinnouchi Y, Takenaka K, Matsui T, Nakamura K, Imai-zumi T. Elevated serum levels of pigment epithelium-derived factor in the metabolic syndrome. *J Clin Endocrinol Metab* 91: 2447–2450, 2006.
317. Yamaoka-Tojo M, Tojo T, Masuda T, Machida Y, Kitano Y, Kurosawa T, Izumi T. C-reactive protein-induced production of interleukin-18 in human endothelial cells: a mechanism of orchestrating cytokine cascade in acute coronary syndrome. *Heart Vessels* 18: 183–187, 2003.
318. Yamawaki H. Vascular effects of novel adipocytokines: focus on vascular contractility and inflammatory responses. *Biol Pharm Bull* 34: 307–310, 2011.
319. Yamawaki H, Kuramoto J, Kameshima S, Usui T, Okada M, Hara Y. Omentin, a novel adipocytokine inhibits TNF-induced vascular inflammation in human endothelial cells. *Biochem Biophys Res Commun* 408: 339–343, 2011.
320. Yan QW, Yang Q, Mody N, Graham TE, Hsu CH, Xu Z, Houstis NE, Kahn BB, Rosen ED. The adipokine lipocalin 2 is regulated by obesity and promotes insulin resistance. *Diabetes* 56: 2533–2540, 2007.
321. Yang RZ, Lee MJ, Hu H, Pray J, Wu HB, Hansen BC, Shuldiner AR, Fried SK, McLenithan JC, Gong DW. Identification of omentin as a novel depot-specific adipokine in human adipose tissue: possible role in modulating insulin action. *Am J Physiol Endocrinol Metab* 290: E1253–E1261, 2006.
322. Yang YY, Hu CJ, Chang SM, Tai TY, Leu SJ. Aspirin inhibits monocyte chemoattractant protein-1 and interleukin-8 expression in TNF-alpha stimulated human umbilical vein endothelial cells. *Atherosclerosis* 174: 207–213, 2004.
323. Yasunari K, Maeda K, Nakamura M, Yoshikawa J. Pressure promotes angiotensin II-mediated migration of human coronary smooth muscle cells through increase in oxidative stress. *Hypertension* 39: 433–437, 2002.
324. Yeung DC, Xu A, Cheung CW, Wat NM, Yau MH, Fong CH, Chau MT, Lam KS. Serum adipocyte fatty acid-binding protein levels were independently associated with carotid atherosclerosis. *Arterioscler Thromb Vasc Biol* 27: 1796–1802, 2007.
325. Youn BS, Kloting N, Kratzsch J, Lee N, Park JW, Song ES, Ruschke K, Oberbach A, Fasshauer M, Stumvoll M, Bluher M. Serum vaspin concentrations in human obesity and type 2 diabetes. *Diabetes* 57: 372–377, 2008.
326. Yudkin JS. Adipose tissue, insulin action and vascular disease: inflammatory signals. *Int J Obes Relat Metab Disord* 27, Suppl 3: S25–S28, 2003.
327. Yudkin JS, Stehouwer CD, Emeis JJ, Coppack SW. C-reactive protein in healthy subjects: associations with obesity, insulin resistance, and endothelial dysfunction: a potential role for cytokines originating from adipose tissue? *Arterioscler Thromb Vasc Biol* 19: 972–978, 1999.
328. Yue TL, Mckenna PJ, Gu JL, Feuerstein GZ. Interleukin-8 is chemotactic for vascular smooth muscle cells. *Eur J Pharmacol* 240: 81–84, 1993.
329. Zakyntinos E, Pappa N. Inflammatory biomarkers in coronary artery disease. *J Cardiol* 53: 317–333, 2009.
330. Zhang J, Wu Y, Zhang Y, LeRoith D, Bernlohr DA, Chen X. The role of lipocalin 2 in the regulation of inflammation in adipocytes and macrophages. *Mol Endocrinol* 22: 1416–1426, 2008.
331. Zhang Z, Chu G, Wu HX, Zou N, Sun BG, Dai QY. IL-8 reduces VCAM-1 secretion of smooth muscle cells by increasing p-ERK expression when 3-D co-cultured with vascular endothelial cells. *Clin Invest Med* 34: E138–E146, 2011.
332. Zhao HY, Zhao M, Yi TN, Zhang J. Globular adiponectin protects human umbilical vein endothelial cells against apoptosis through adiponectin receptor 1/adenosine monophosphate-activated protein kinase pathway. *Chin Med J (Engl)* 124: 2540–2547, 2011.
333. Zhao Y, Biswas SK, McNulty PH, Kozak M, Jun JY, Segar L. PDGF-induced vascular smooth muscle cell proliferation is associated with dysregulation of insulin receptor substrates. *Am J Physiol Cell Physiol* 300: C1375–C1385, 2011.
334. Zhong J, Krawczyk SA, Chaerkady R, Huang H, Goel R, Bader JS, Wong GW, Corkey BE, Pandey A. Temporal profiling of the secretome during adipogenesis in humans. *J Proteome Res* 9: 5228–5238, 2010.
335. Zhou G, Zhou Z, Ge S, Liu D, Zhang R, Xu G, Zhu W, Yin Q, Chen AF, Liu X. IL-18 accelerates the cell apoptosis by up-regulating cysteinyl leukotriene 2 receptor expression in human umbilical vein endothelial cells at the early stage of administration. *Vascul Pharmacol* 50: 171–177, 2009.
336. Zhu N, Zhang D, Chen S, Liu X, Lin L, Huang X, Guo Z, Liu J, Wang Y, Yuan W, Qin Y. Endothelial enriched microRNAs regulate angiotensin II-induced endothelial inflammation and migration. *Atherosclerosis* 215: 286–293, 2011.
337. Zimmerman MA, Reznikov LL, Raeburn CD, Selzman CH. Interleukin-10 attenuates the response to vascular injury. *J Surg Res* 121: 206–213, 2004.
338. Zou M, Liu X, Li Y, Lai Y. Experimental study on the migration of vascular endothelial cells stimulated by IL-8. [In Chinese.] *Sheng Wu Yi Xue Gong Cheng Xue Za Zhi* 23: 1013–1016, 2006.