A model of preeclampsia in rats: the reduced uterine perfusion pressure (RUPP) model

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Submitted 9 February 2012; accepted in final form 12 April 2012

Li J, LaMarca B, Reckelhoff JF. A model of preeclampsia in rats: the reduced uterine perfusion pressure (RUPP) model. Am J Physiol Heart Circ Physiol 303: H1–H8, 2012.—Preeclampsia is defined as new-onset hypertension with proteinuria after 20 wk gestation and is hypothesized to be due to shallow trophoblast invasion in the spiral arteries thus resulting in progressive placental ischemia as the fetus grows. Many animal models have been developed that mimic changes in maternal circulation or immune function associated with preeclampsia. The model of reduced uterine perfusion pressure in pregnant rats closely mimics the hypertension, immune system abnormalities, systemic and renal vasoconstriction, and oxidative stress in the mother, and intrauterine growth restriction found in the offspring. The model has been successfully used in many species; however, rat and primate are the most consistent in comparison of characteristics with human preeclampsia. The model suffers, however, from lack of the ability to study the mechanisms responsible for abnormal placentation that ultimately leads to placental ischemia. Despite this limitation, the model is excellent for studying the consequences of reduced uterine blood flow as it mimics many of the salient features of preeclampsia during the last weeks of gestation in humans. This review discusses these features.

Preeclampsia is Associated with Abnormal Uteroplacental Blood Flow

It has been hypothesized that preeclampsia arises from reductions in uteroplacental perfusion, leading to fetoplacental ischemia (82, 83). The mechanisms leading to reduced placental perfusion in preeclampsia are not clear, but studies in humans implicate impaired first-trimester cytotrophoblast invasion of spiral arterioles as an important factor. During weeks 8–18 of normal pregnancy, the villous cytotrophoblasts originating from the fetus invade the uterine wall through the entire depth of the endometrium and the inner third of the myometrium (42, 82) (see Fig. 1). As a result, the spiral arteries are extensively remodeled, replacing their smooth muscle and endothelium by the invading cytotrophoblasts. These arteries become widely dilated, with low resistance, and are less responsive, or even refractory, to vasoconstrictor substances (45). This normal invasive vascular remodeling occurring early in placentation is abnormal in pregnant women that develop preeclampsia (45). Thus preeclampsia is thought to be caused by a reduction in uterine blood flow due to abnormal trophoblast invasion of the spiral arteries, a concept first suggested by Young in 1914 (107). In support of this hypothesis, preeclamptic human placent al bed biopsies indicate shallow trophoblast invasion, compared with normal pregnancy. As a result, the corresponding arterial segments remain undilated, leading to insufficient blood flow to the uteroplacental unit, and the
Fig. 1. The three-stage model of preeclampsia. Stages 1 and 2 of preeclampsia are well accepted, whereas stage 0 is hypothetical. Based on this model, preeclampsia is proposed to arise from abnormal maternal immune response to fetopaternal antigens in the early stage of pregnancy (stage 0), leading to poorly perfused placenta in stage 1, which further causes multiple stresses and the final clinical manifestations in stage 2 of preeclampsia (82, 83).

Maternal circulation remains responsive to vasoconstrictors, such as angiotensin II (ANG II) (18) and endothelin (ET-1) (51). The ischemic placenta releases cytokines, reactive oxygen species, and anti-angiogenic factors, such as soluble fms-like tyrosine kinase-1 (sFlt-1) (35), soluble endoglin (69, 70), and ANG II type 1 receptor autoantibodies (AT1-AA) (100), into the circulation, making this an aggravated interaction between maternal and fetal circulation. It should be mentioned that studies in double-knockout Rag2-/-Il2rg-/- mice, in which there is no spiral artery remodeling during pregnancy, placental ischemia is present, but this does not result in IUGR or hypertension (25). These data suggest that other mechanisms are involved in the development of preeclampsia in addition to poor spiral artery remodeling at least in the mouse.

Animal Models for the Study of Preeclampsia

An ideal animal model to study preeclampsia should duplicate the pathogenesis of preeclampsia in women, including failure of early immune mechanisms, impaired first-trimester trophoblast invasion and placentation, reduced uteroplacental perfusion, and fetoplacental ischemia, thus resulting in systemic inflammation, proteinuria, and endothelial dysfunction in the mother and restricted growth in the fetus. However, preeclampsia is thought to occur spontaneously only in women, and only very few cases have been reported in great apes, including chimpanzees (49, 91) and gorillas (15, 95). The use of nonhuman primates and other animal species as models for preeclampsia are discussed at length in a review by Podjarny et al. (80) and McCarthy et al. (72).

There is evidence that hypertensive inbred BPH-5 mice, derived from brother-sister mating of borderline hypertensive BPH/2 mice, exhibit further increases in blood pressure (BP) in late gestation (from ~130 to 160 mmHg, compared with ~105 mmHg in control C57BL-6) that resolves within 2 days of delivery and is accompanied by increases in proteinuria and IUGR of the pups (26, 28). The mechanisms responsible for the increased BP during pregnancy in this model have not been elucidated, but adenoviral vector-mediated vascular endothelial growth factor (VEGF) 121 (46) and tempol (103) prevented the increase in BP (103), suggesting that placental ischemia and oxidative stress may contribute to the further elevated BP.

Models to Evaluate a Single Pathway in the Progression of Preeclampsia

There are several genetically manipulated mouse models of preeclampsia that affect a single gene, such as sFlt-1, the renin-angiotensin system, VEGF, endothelin, endothelial nitric oxide synthase, etc. (47). These models have varying levels of similarity with human preeclampsia, although, interestingly, few develop new-onset hypertension close to term. Readers are referred to a recent review by Ishida and colleagues (47) for more information on mouse models of preeclampsia. Below we discuss studies performed in models other than mice to learn more about the role of specific pathways hypothesized to contribute to the pathogenesis of preeclampsia.

Nitric Oxide Inhibition

Nitric oxide (NO) is an important mediator in controlling vascular tone, endothelial function, and oxidative stress. Increased NO production plays a critical role in the maternal BP and glomerular hemodynamic adaptation to pregnancy. NO production is reduced in preeclampsia (22, 27). Chronic NO synthase (NOS) inhibition in pregnant rats causes a dose-dependent increase in BP, renal vasoconstriction, proteinuria, maternal morbidity and mortality, and IUGR of pups (19, 74, 106). However, plasma from women with preeclampsia increases, rather than decreases, endothelial cell NO (16), and the effect of pregnancy on BP in endothelial NOS knockout mice is controversial, with one study reporting typical pregnancy-induced decreases in BP (89) while another reported a further increase of BP during pregnancy (43).

Anti-Angiogenic Factors

Women with preeclampsia have elevated circulating sFlt-1, a soluble VEGF receptor binding to and inactivating VEGF and placental growth factor (PIGF), critical players in angiogenesis and placentation. Placental and amniotic sFlt-1 levels are elevated while plasma levels of free VEGF and PIGF are decreased (70). Adenoviral vector-mediated sFlt-1 in pregnant rats resulted in increased BP, proteinuria, glomerular endotheliosis, and decreases in plasma free VEGF and PIGF (70). Similar studies performed in mice resulted in similar vascular effects with significantly lower pup and placental weight (66). In later studies performed by Murphy et al. (17, 76), sFlt-1 infusion in rats increased vascular/placental oxidative stress, decreased vasodilatory responses to agonists, reduced placenta and fetal weight, decreased maternal circulating VEGF and NO, and increased renal preproendothelin (prepro-ET-1). Importantly, the effects of sFlt-1 also occurred in a dose-dependent manner in nonpregnant controls, indicating these results are not specific for pregnancy, suggesting that increases in sFlt-1 under any circumstances compromise the cardiovascular system.

Models Based on Inflammatory Mediators

Women with preeclampsia exhibit increased systemic inflammation, heightened oxidative stress, and circulating auto-
Intramurine sFlt-1, soluble fms-like tyrosine kinase-1; IUGR, intrauterine growth restriction.

Sheep Occluder around internal iliac artery or abdominal

Guinea pig Banding uterine arteries, ovarian arteries Inc BP, UPrV, creatinine 39

models (e.g., sheep) and humans may alter the cardiovas-

cular responses to the RUPP maneuvers. Furthermore, in
dogs (104, 105), rabbits (3, 65), sheep (23, 61), guinea pigs (39), primates (4, 20) (see Table 1), and rats (1, 11, 29). It should be noted that differences in placentaion between animal models (e.g., sheep) and humans may alter the cardiovascular responses to the RUPP maneuvers. Furthermore, in sheep, increased BP only occurs with RUPP when dams are given a high-salt diet (61). The most well-characterized and utilized is the RUPP pregnant rat model (see Table 2). To our knowledge, there have been no studies using the RUPP model in mice.

Method to Induce the RUPP Model in Pregnant Rats

The RUPP rat model of preeclampsia was adapted by Crews and colleagues (24), and the clipping procedure is tightly controlled, in terms of gestation time and silver clip position and size. Pregnant rats weighing ~200–250 g undergo clipping at day 14 of gestation, in which a silver clip (0.203 mm ID) is placed around the aorta above the iliac bifurcation (see Fig. 2). Because compensatory blood flow to the placenta occurs via an adaptive increase in ovarian blood flow, both right and left uterine arcades are also clipped (silver clip, 0.100 mm ID) (11). These procedures reduce uterine blood flow in the gravid rat by ~40% (24).

Characteristics of the RUPP Model

The RUPP rat model of preeclampsia mimics numerous physiological features of preeclampsia in women (see Table 3), including hypertension (−20–30 mmHg increase in mean arterial pressure), proteinuria (~5-fold increase in urinary protein excretion), impaired renal function, as indicated by reduction in glomerular filtration rate (GFR) (<40%) and renal plasma flow (–23%), an increase in vascular reactivity, a reduction in cardiac index (90), and increases in leptin (14) and blood lactate (36). Fetal IUGR also occurs in RUPP rats, with decreased litter size and pup weight (10, 40).

RUPP rats have reductions in NO (24) and increases in vasoconstrictor substances or response (84). Vascular smooth muscle cells isolated from renal interlobar arteries displayed increased contractility to ANG II via enhanced calcium entry (75). Inhibition of ANG II synthesis with a converting-enzyme inhibitor had no effect on the BP in RUPP rats (8), but, in contrast, treatment of RUPP rats with losartan, the AT1R antagonist, significantly reduced their BP (58). These findings lead to the discovery that, just as in women with preeclampsia, RUPP rats have increased AT1-AA that causes activation of AT1R and is a contributor to hypertension in the model (100). RUPP rats also exhibit increased tissue ET-1, and ETA receptor antagonists attenuate the BP increases (93). Furthermore, as in women with preeclampsia, sera from RUPP rats causes endothelial cell activation that is attenuated by AT1R blockade.

Table 1. Models of reduced uterine perfusion pressure in animals other than rat

<table>
<thead>
<tr>
<th>Species</th>
<th>Method</th>
<th>Characteristics</th>
<th>Ref. No.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dog</td>
<td>Occlusion of abdominal aorta below the renal arteries</td>
<td>Inc BP, UPRV, glomerular lesions, fluid retention diffuse hemorrhagic infarction of the placenta</td>
<td>3, 5, 77, 104, 105</td>
</tr>
<tr>
<td>Rabbit</td>
<td>Clamp or ligation of aorta proximal to the ovarian and distal to the renal arteries</td>
<td>Inc BP, UPRV, PVR, IUGR; endothelial swelling, inc glomerular fibrinogen RVR, placential lesions</td>
<td>2, 3, 65</td>
</tr>
<tr>
<td>Nonhuman primate</td>
<td>Red, abdominal aortic blood flow below kidneys</td>
<td>Inc BP, UPRV, RVR, serum uric acid, dec PRA, endotheliosis of glomeruli, inc sFlt-1</td>
<td>4, 20, 68, 91</td>
</tr>
<tr>
<td>Guinea pig</td>
<td>Banding uterine arteries, ovarian arteries</td>
<td>Inc BP, UPRV, creatinine</td>
<td>39</td>
</tr>
<tr>
<td>Sheep</td>
<td>Occlude around internal iliac artery or abdominal aorta distal to renal arteries</td>
<td>Inc BP only with high-salt diet, IUGR</td>
<td>23, 61, 94</td>
</tr>
</tbody>
</table>

Inc, increased; dec, decreased; BP, blood pressure; UPRV, proteinuria; PVR, peripheral vascular resistance; RVR, renal vascular resistance; PRA, plasma renal activity; sFlt-1, soluble fms-like tyrosine kinase-1; IUGR, intrauterine growth restriction.
suggested a role for the AT1-AA in the circulation of RUPP rats to activate the AT1R on the vascular endothelium (84). As discussed previously, women with preeclampsia have increases in the levels of inflammatory cytokines (52). In RUPP rats, serum levels of TNF-α and IL-6 are increased, and infusion of TNF-α or IL-6 increases BP in normal pregnant rats, suggesting that inflammation could contribute to the hypertension of RUPP rats (30, 59). Supporting this notion, administration of a TNF-α soluble receptor, etanercept, attenuated the hypertension and reduced ET-1 transcript expression and endothelial cell activation from RUPP rats (55), Sunderland and colleagues (92) recently reported similar findings with infusion of TNF-α in pregnant baboons that also resulted in increases in sFlt-1 and soluble endoglin.

Angiostatic factors, such as sFlt-1 and soluble endoglin, may be important in development and progression of preeclampsia in women (69, 98). RUPP rats exhibit increased plasma and placental sFlt-1 and decreased plasma VEGF and PIGF (35). A recent study demonstrated that chronic infusion of recombinant VEGF-121 during late gestation restored GFR and endothelial function and reduced BP in RUPP rats (38). Serum and placental soluble endoglin are increased along with placental hypoxia-inducible factor-1α expression, whereas placental heme oxygenase-1 (HO-1) is decreased in the RUPP rats (37). An HO-1 inducer and IL-6 are increased, and renal superoxide dismutase activity is decreased in RUPP rats. Chronic treatment with tempol attenuates the hypertension associated with RUPP (88). George and colleagues (32, 34) have recently shown that the reduction in sFlt-1 with heme oxygenase induction is mediated via production of antioxidants, biliverdin and carbon monoxide.

**Advantages of the RUPP Model**

The RUPP model of preeclampsia resembles numerous features exhibited by women with preeclampsia (Table 3) and...
serves as a good tool for investigating potential mechanisms underlying hypertension induced by placental ischemia in pregnancy. Importantly, RUPP models also successfully develop fetal IUGR, which is not always seen in other animal models of preeclampsia, and is a very useful model for studying fetal programming of cardiovascular disease. The RUPP model has also been exploited in nonhuman primates by Makris and colleagues (68), who find similar changes such as hypertension and elevated sFlt-1. The RUPP model provides the opportunity to study potential therapeutic approaches for management of preeclampsia that may not be appropriate for study in women due to the consideration of the health of the fetus.

**Limitations of the RUPP Model**

Despite the utility of the RUPP model of preeclampsia, the model does have several limitations. First, an increase in BP with RUPP does not always occur in all species or even within the same species (5). The RUPP model developed by Granger (40), LaMarca (54–57), Alexander (7), and colleagues in rats requires the clipping of both the lower abdominal aorta and the uterine arteries on day 14 of gestation in ~250 g female rats (78). Thus the conditions to produce the model must be rigorous to have a successful outcome of hypertension that recedes after delivery of the pups. The use of rats as a RUPP model is cost effective, but somewhat limited, since the clipping is performed at day 14 of gestation, which only lasts 21–22 days. Thus the window for study is narrow in the rat. The use of nonhuman primates for the RUPP model, as characterized by Makris and colleagues (68), provides a longer gestation time, placentation, and trophoblast invasion closer to humans than rats but is significantly more costly to maintain the animals.

A second important limitation of the RUPP model is that it only mimics the pathogenesis responsible for the hypertension in preeclampsia downstream of midgestational placental ischemia. Thus the RUPP model is not useful to investigate the early immune mechanisms, trophoblast invasion, and vascular remodeling abnormalities. Furthermore, aortic clipping may have a hemodynamic impact on sympathetic nervous system and cardiac output, since Sholook and colleagues (90) reported that the RUPP procedure results in reductions in blood flow to the heart, stomach, intestine, and skeletal muscle, without differences in blood flow to the brain, liver, kidney, or spleen compared with the normal pregnant rats.

A third limitation is that the RUPP model in the rat does not mimic the glomerular endotheliosis that is one of the hallmarks of preeclampsia (62). This may be important because several investigators find that podocyte shedding occurs in women with preeclampsia and may be an early marker for preeclampsia (6, 31).

Finally, the RUPP model is not successful in mimicking severe preeclampsia, which is characterized by the HELLP (hemolysis, elevated liver enzymes, and low platelets) syndrome. RUPP rats do not have changes in hemoglobin, platelets, or liver function (48).

**Summary**

In conclusion, pregnancy in women involves unique fetal villous trophoblast invasion and placental vascular remodeling, namely placentation. This process is only seen in women and

### Table 3. Comparison of characteristics in women with preeclampsia and the model of RUPP in the rat

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Women with Preeclampsia</th>
<th>RUPP in Rats</th>
<th>Ref. No.</th>
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<tbody>
<tr>
<td>New-onset hypertension</td>
<td>Yes</td>
<td>Yes</td>
<td>11</td>
</tr>
<tr>
<td>Abnormal placentation</td>
<td>Yes</td>
<td>No</td>
<td>42, 45, 82, 83</td>
</tr>
<tr>
<td>Proteinuria</td>
<td>Yes</td>
<td>Yes</td>
<td>11</td>
</tr>
<tr>
<td>Renal plasma flow</td>
<td>Dec</td>
<td>Dec</td>
<td>90</td>
</tr>
<tr>
<td>GFR</td>
<td>Dec</td>
<td>Dec</td>
<td>11</td>
</tr>
<tr>
<td>Inflammation (TNF-α, IL-6, CD4+ T cells)</td>
<td>Inc</td>
<td>Inc</td>
<td>30, 59, 99</td>
</tr>
<tr>
<td>Oxidative stress</td>
<td>Inc</td>
<td>Inc</td>
<td>88</td>
</tr>
<tr>
<td>Endothelin-1</td>
<td>Inc</td>
<td>Inc</td>
<td>33</td>
</tr>
<tr>
<td>sFlt-1</td>
<td>Inc</td>
<td>Inc</td>
<td>35</td>
</tr>
<tr>
<td>VEGF, PIGF</td>
<td>Dec</td>
<td>Dec</td>
<td>35</td>
</tr>
<tr>
<td>Soluble endoglin</td>
<td>Inc</td>
<td>Inc</td>
<td>37</td>
</tr>
<tr>
<td>Placental HIF-1α</td>
<td>Inc</td>
<td>Inc</td>
<td>37</td>
</tr>
<tr>
<td>AT1-AA</td>
<td>Inc</td>
<td>Inc</td>
<td>58</td>
</tr>
<tr>
<td>Glomerular endotheliosis</td>
<td>Yes</td>
<td>No</td>
<td>62</td>
</tr>
<tr>
<td>Podocyte shedding</td>
<td>Yes</td>
<td>Yes</td>
<td>6, 31</td>
</tr>
<tr>
<td>HELLP syndrome</td>
<td>In severe cases</td>
<td>No</td>
<td>48</td>
</tr>
<tr>
<td>Fetal IUGR</td>
<td>Yes</td>
<td>Yes</td>
<td>7, 52</td>
</tr>
</tbody>
</table>

*TNF-α, tumor necrosis factor-α; IL-6, interleukin-6; PIGF, placental growth factor; HIF-1α, hypoxia-inducible factor-1α; AT1-AA, agonistic autoantibodies against angiotensin type 1 receptor; HELLP syndrome, hemolysis, elevated liver enzymes, low platelets.*
great apes. Thus to our knowledge, preeclampsia occurs spontaneously only in women and great apes due to abnormal placentation and the resulting placental ischemia. Thus no animal model of preeclampsia can mimic the entire pathogenesis of the disease as seen in women. A variety of animal models, including genetically manipulated mouse models, have been developed based on a single mediator of preeclampsia that only reflects limited aspects of the mechanisms underlying the disease; however, these are useful to identify mechanisms that could potentially contribute to the pathogenesis of preeclampsia. The RUPP model has been performed in various species (see Table 1) and involves induction of placental ischemia by reducing uterine blood flow by some maneuver, such as placement of clips on the uterine arteries and abdominal aorta, that results in a preeclamptic state. The RUPP model in the rat has been the most extensively studied (see Table 2) and exhibits many of the manifestations of preeclampsia (see Table 3). Unfortunately, the greatest limitation of the RUPP model is its lack of suitability for the study of early placentation and the subsequent pathogenesis responsible for placental ischemia, such as early occurring abnormal immune mechanisms or trophoblast invasion and vascular remodeling. However, the RUPP model is excellent for the study of many of the consequences of placental ischemia, including hypertension, vascular dysfunction, and immune dysfunction. The ability to develop the RUPP model in both rats and nonhuman primates makes it advantageous for many future studies. Whether the RUPP model could be developed in mice is not clear.

GRANTS

This work was supported by National Institutes of Health (NIH) RO1s HL-66072, HL-69194, and PO1 HL-51971 (to J. F. Reckelhoff), American Heart Association Frant SDG 0835472N, and NIH Grant HD-067541. (to B. B. and J. F. Reckelhoff), American Heart Association Frant SDG 0835472N, and NIH Grant HD-067541. (to B. B. and J. F. Reckelhoff).

DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

AUTHOR CONTRIBUTIONS

Author contributions: J. L. and B. L. prepared figures; J. L. drafted manuscript; B. L. and J. F. R. edited and revised manuscript; J. F. R. approved final version of manuscript.

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AJP-Heart Circ Physiol • doi:10.1152/ajpheart.00117.2012 • www.ajpheart.org

RAT MODEL OF PREECLAMPSIA H7
H8 RAT MODEL OF PREECLAMPSIA


