Multilevel systems biology modeling characterized the atheroprotective efficiencies of modified dairy fats in a hamster model

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In fact, several strategies have aimed to manipulate the composition of dairy fats to improve their cardiovascular features further (21, 30, 41, 45, 51). Targeting both saturated fatty acids and cholesterol in milk fat has appeared advantageous in humans with regard to cardiovascular disease risk (21, 41, 51). To target these components, zootechnical means, such as modifying the cattle diet by providing polyunsaturated fatty acids, have been used (41, 51), but there is a physiological limitation on the extent to which the profiles of fatty acids can be modified, and the cholesterol content cannot be changed. Physicochemical techniques allow for more important modifications, such as physical fractionation, interesterification, mixing with other fats, and the removal of cholesterol.

We thus designed a study that examined the atherosclerotic impacts of various technologically transformed dairy fats in a hamster model that we and others have found suitable for addressing this issue (23, 33, 69). The occurrence of atherosclerosis, which constitutes one of the main pathological foundations of cardiovascular disease, is highly multifactorial. Hence, evaluating and comparing dietary strategies to improve atherosclerosis also require the examination of a comprehen-
sive set of disease components in addition to measuring the severity of vascular lesions (34). We thus designed a systems biology approach for identifying the diet-related components of the disease and assessing how they could be differentially modified by each dietary treatment. To achieve these goals, we implemented a multiphase analysis combining conventional blood clinical chemistry and fatty acid analyses, broad atherosclerosis-related gene expression analysis of blood and liver samples, and untargeted metabolite profiling of biofluids. The complex biological information obtained was condensed into a reduced number of biological sets, which were modeled into scores to facilitate interpretation. The quantitative contribution of each biological score to atherogenesis across the dietary conditions was calculated from a multivariate predictive equation. Interaction graphs allowed us to report the cross talk among the biological functions in modulating atherosclerosis, as well as the differential responses of these functions to the various dietary fats.

MATERIALS AND METHODS

Animals and Diets

All of the experiments were approved by the referee for animal care at our institution and were conducted in accordance with the French Regulations for Animal Experimentation (Art. 19, Oct., 1987, Ministry of Agriculture). All of the animal procedures conformed to the guidelines from Directive 2010/63/EU of the European Parliament on the protection of animals used for scientific purposes. Forty male Golden Syrian hamsters (9 wk old, Janvier strain weighing 65–70 g) were purchased from the Janvier Breeding Center (Le Genest-St. Isle, France). Each hamster was individually caged and fed a commercial chow diet (25/18 standard diet from Mucedola, Settimo, Milan, Italy) for a 1-wk adaptation period. Eight hamsters per group were then fed experimental diets (200 g/kg) for 12 wk. The various fats were: regular butter (RB); RB diluted to 50% (by weight) by a vegetable oil mixture made with 50% canola oil, 30% high oleic sunflower oil, and 20% sunflower oil (VRB); a low cholesterol butter diluted with the above plant oil mix (VLCB); a low cholesterol and a low saturated fatty acid butter (LCSB); and a plant oil mixture of 30% canola oil and 70% palm oil (VM). Diet and water were available ad libitum. An anti-oxidant mixture (Oxy‘Block; Naturex, Avignon, France), containing carnosic acid and carnosol (8%) and soya tocopherols (35%) completed to 100% with sunflower oil, was added at 300 ppm to the LCSB fat. The dietary fats were provided by Corman. The compositions of the various fat diets are listed in Table 1.

Blood Sampling and Chemical Assays

The hamsters were anesthetized 5–8 h after their last meal by an intramuscular injection of Zoletil 50 at a dose of 250 mg/kg. A cardiac puncture was used to collect 3–4 ml of blood, and plasma aliquots were stored at −80°C until ready for analysis.

Plasma lipids were measured by enzymatic procedures using commercial kits (Biomerieux, Lyon, France), as follows: total cholesterol (RTU method) and triglycerides and phospholipids (PAP 150 method) (59).

HDL-cholesterol (HDL-Chol) was measured after dextran precipitation (Abcam ab65390 assay kit, Cambridge, UK), and the non-HDL-Chol (VLDL + LDL) was calculated by the difference between total cholesterol and HDL-Chol.

Two milliliters of whole blood were also collected in PAXgene blood RNA tubes (Qiagen, Courtaboeuf, France) and were stored at −80°C for further quantitative PCR analysis of gene expression.

Aorta Sampling

For each hamster, the aorta was rinsed by perfusing cold saline from the left ventricle and was excised from the aortic arch up to the inferior iliac bifurcation. Whole aortic lipid classes were extracted and analyzed by gas liquid chromatography (LC), using a high-temperature nonpolar capillary column (33, 60). Whole aortic cholesteryl esters were considered to indicate early atherogenesis (fatty streaks) and were expressed as percentages of cholesteryl-esters of tissue phospholipids (33, 39). Such biochemical analysis was preferred over staining methods due to practical considerations and better accuracy than staining evaluations.

Metabolomics

Sample preparation. Urine samples were collected with a needle inserted directly into the bladder after anesthesia and were stored at −80°C before processing, thus preventing any oxidation. After thawing, the samples were processed by centrifugation at 17,500 g for 10 min at 4°C and then were diluted with ultra-pure water (1:4 vol/vol) and centrifuged again at 8,000 g for 5 min. The supernatant was collected and kept at −80°C until analysis.

Blood was collected from the left ventricle in heparinized tubes and left for no more than 1 h in a bed of ice. Plasma was separated by centrifugation (2,000 g 10 min) and was stored immediately at −80°C. Plasma was thawed before extraction. Metabolite extraction has been validated elsewhere (31, 46). Polar compounds in plasma were extracted by mixing 400 μl of cold methanol with 200 μl of plasma. After a minimum of 30 min of incubation at −20°C, the samples were thoroughly shaken and then centrifuged for 15 min at 11,000 rpm and 4°C. The supernatants were transferred to fresh microtubes and centrifuged again for 15 min under the same conditions. The supernatants were transferred again into fresh tubes, dried under nitrogen flow, and stored at −80°C before analysis. To assess the data quality, a blank sample (deionized water) and a pooled sample (a mixture of all of the samples) were extracted/diluted and analyzed repeatedly along with the sample series (49).

For the purpose of further metabolite identification, MS/MS analyses were performed on selected ions in the urine and plasma samples and completed by real standard analysis whenever possible. When identification was made with standard molecules, both retention times and MS/MS fragmentation spectra were used to confirm metabolite annotation. For MS/MS, the MRM parameters were set at 8.0 for isolation width, 2.0 for acquisition factor, and 10.0, 20.0, and 30.0 eV for collision energies. Solutions of 1 mg/ml were prepared in different solvent mixtures according to each molecule’s LogP value and were injected at a 10 μg/ml concentration. MS/MS spectra analyses and metabolite assignments are reported in the Supplemental Table S1.

Gas-Chromatography Fatty Acid Analysis

Five-hundred microliters of plasma and 100 μg of liver tissues were extracted according to the methods of Bligh and Dyer (7) and of Folch et al. (12). Plasma lipid extracts were fractionated by thin-layer chromatography, and the fatty acid moieties of the fractionated lipid classes were transmethylated using boron trifluoride 5% in methanol (Sigma-Aldrich, L’Isle d’Abeau Chesnes, France) (32). Fatty acids in total liver lipid extracts were transmethylated using acetyl-chloride in methanol, as described earlier (11). Fatty acid methyl esters were
Quantitative PCR for Gene Expression Analysis

Total RNA was isolated from liver samples using Nucleopin RNA II purification columns (Macherey-Nagel, Horedt, France) according to the suppliers’ instructions. Blood RNA preserved in the PAXgene RNA tubes was extracted according to the instructions of the PAX-Gene RNA kit. The integrity of the RNA was assessed using an Agilent 2100 Bioanalyser; RNA samples scoring an RIN greater than 8 were considered of good quality and were used for further analysis. From 1.5 g/9262 g of RNA extracted from liver samples or 0.4 g/9262 g of RNA extracted from blood samples, cDNA was synthesized using SuperScript-II Reverse Transcription kit (Invitrogen, Les Ulis, France) and was diluted in sterile water to final volumes of 340 and 100 μl, respectively. Gene expression levels were determined using real-time PCR (on a Stratagene Mx 3005, La Jolla, CA) with SYBR Green as a fluorescent dye. 18S was used as a reference gene because it showed stable expression for all of the diet groups (data not shown). All of the pairs of primers were previously validated (36), and the stability of the Tm products was verified after each run. All of the biological samples were run in duplicate. Gene expression was calculated using the mean normalized expression. The expression of 44 genes implicated in the metabolic control was examined in each hamster (35 in liver and 9 in whole blood; Supplemental Fig. S1).

Statistics

All partial least squares (PLS) models and analyses were performed using SIMCA P+11 (Umetrics, Umea, Sweden). Interaction networks were calculated using Cytoscape (52), hierarchical analyses with Permutmatrix (8), receiver operator characteristic (ROC) curve and random forest analyses with Metaboanalyst (66), naïve Bayes analyses with Tanagra (48) (http://eric.univ-lyon2.fr/~ricco/tanagra/), univariate statistics with Statview (SAS Institute, Cary, NC), and partial correlations with the R package GeneNet. The statistical workflow in connection with the analytical analyses is shown is Fig. 1.

Table 1. Diet composition, including the fatty acids, cholesterol, phytosterol, and tocopherol contents of the lipid moiety

<table>
<thead>
<tr>
<th>Diet Components</th>
<th>RB</th>
<th>VRB*</th>
<th>VLCB†</th>
<th>LCSB</th>
<th>VM†</th>
</tr>
</thead>
<tbody>
<tr>
<td>Nutrients, g/100 g</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>dl-Methionine</td>
<td>0.16</td>
<td>0.16</td>
<td>0.16</td>
<td>0.16</td>
<td>0.16</td>
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<tr>
<td>Starch</td>
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<td>39.84</td>
<td>39.84</td>
<td>39.84</td>
<td>39.84</td>
</tr>
<tr>
<td>Sucrose</td>
<td>8</td>
<td>8</td>
<td>8</td>
<td>8</td>
<td>8</td>
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<tr>
<td>Casein</td>
<td>22</td>
<td>22</td>
<td>22</td>
<td>22</td>
<td>22</td>
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<tr>
<td>Lipids</td>
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</tr>
<tr>
<td>Fiber</td>
<td>5</td>
<td>5</td>
<td>5</td>
<td>5</td>
<td>5</td>
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<tr>
<td>Minerals</td>
<td>4</td>
<td>4</td>
<td>4</td>
<td>4</td>
<td>4</td>
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<tr>
<td>Vitamins mix</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>Energy, MJ/kg</td>
<td>19.5</td>
<td>19.5</td>
<td>19.5</td>
<td>19.5</td>
<td>19.5</td>
</tr>
<tr>
<td>Fatty acids, g/100 g</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>C4:0</td>
<td>3.4</td>
<td>1.7</td>
<td>1.7</td>
<td>0.7</td>
<td>0</td>
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<tr>
<td>C6:0</td>
<td>2.2</td>
<td>1.1</td>
<td>1.1</td>
<td>0.8</td>
<td>0</td>
</tr>
<tr>
<td>C8:0</td>
<td>1.3</td>
<td>0.6</td>
<td>0.6</td>
<td>0.6</td>
<td>0</td>
</tr>
<tr>
<td>C10:0</td>
<td>2.6</td>
<td>1.3</td>
<td>1.4</td>
<td>1.8</td>
<td>0.1</td>
</tr>
<tr>
<td>C10:1</td>
<td>0.2</td>
<td>0.1</td>
<td>0.1</td>
<td>0.2</td>
<td>0</td>
</tr>
<tr>
<td>C12:0</td>
<td>3.2</td>
<td>1.6</td>
<td>1.7</td>
<td>2.3</td>
<td>0.2</td>
</tr>
<tr>
<td>C14:0</td>
<td>10.8</td>
<td>5.3</td>
<td>5.4</td>
<td>6.8</td>
<td>0.8</td>
</tr>
<tr>
<td>C14:1</td>
<td>1.7</td>
<td>0.8</td>
<td>0.9</td>
<td>2.3</td>
<td>0</td>
</tr>
<tr>
<td>C15:0</td>
<td>1.1</td>
<td>0.5</td>
<td>0.6</td>
<td>0.7</td>
<td>0.1</td>
</tr>
<tr>
<td>C16:0</td>
<td>30.2</td>
<td>17.2</td>
<td>17.1</td>
<td>17.6</td>
<td>32.4</td>
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<tr>
<td>C16:1 (+t)</td>
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<td>1.3</td>
<td>1.4</td>
<td>3.7</td>
<td>0.4</td>
</tr>
<tr>
<td>C17:0</td>
<td>0.7</td>
<td>0.4</td>
<td>0.3</td>
<td>0.5</td>
<td>0.1</td>
</tr>
<tr>
<td>C17:1</td>
<td>0.3</td>
<td>0.2</td>
<td>0.2</td>
<td>0.5</td>
<td>0</td>
</tr>
<tr>
<td>C18:0</td>
<td>11.1</td>
<td>6.8</td>
<td>6.8</td>
<td>7.9</td>
<td>3.6</td>
</tr>
<tr>
<td>C18:1 (+t)</td>
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<td>42.6</td>
<td>42.3</td>
<td>43.7</td>
<td>45.5</td>
</tr>
<tr>
<td>C18:2n-6</td>
<td>2.4</td>
<td>14.0</td>
<td>13.9</td>
<td>4</td>
<td>12.7</td>
</tr>
<tr>
<td>C18:3n-3</td>
<td>0.6</td>
<td>0.3</td>
<td>0.3</td>
<td>1.5</td>
<td>0</td>
</tr>
<tr>
<td>n-6/n-3</td>
<td>0.4</td>
<td>2.7</td>
<td>2.6</td>
<td>0.9</td>
<td>2.9</td>
</tr>
<tr>
<td>Total saturated</td>
<td>66.6</td>
<td>36.5</td>
<td>36.7</td>
<td>39.7</td>
<td>37.3</td>
</tr>
<tr>
<td>Total mono</td>
<td>28.6</td>
<td>45</td>
<td>44.9</td>
<td>50.4</td>
<td>45.9</td>
</tr>
<tr>
<td>Total poly</td>
<td>3.4</td>
<td>17</td>
<td>16.8</td>
<td>6.4</td>
<td>15.6</td>
</tr>
<tr>
<td>Polyunsaturated/saturated</td>
<td>0.05</td>
<td>0.47</td>
<td>0.46</td>
<td>0.16</td>
<td>0.42</td>
</tr>
<tr>
<td>Unidentified</td>
<td>1.4</td>
<td>1.5</td>
<td>1.6</td>
<td>3.5</td>
<td>1.2</td>
</tr>
<tr>
<td>Cholesterol mg/100 g fat (% in the diet)</td>
<td>240 (0.05)</td>
<td>120 (0.025)</td>
<td>&gt;26 (0.005)</td>
<td>20 (0.004)</td>
<td>0</td>
</tr>
<tr>
<td>Total phytosterols, ppm</td>
<td>212</td>
<td>55,425</td>
<td>57,220</td>
<td>129</td>
<td>62,000</td>
</tr>
<tr>
<td>Stigmastanol</td>
<td>25</td>
<td>24</td>
<td>Not detected</td>
<td>3</td>
<td>13</td>
</tr>
<tr>
<td>β-Sitosterol</td>
<td>6</td>
<td>1,400</td>
<td>1,200</td>
<td>3</td>
<td>960</td>
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<tr>
<td>Campesterol</td>
<td>200</td>
<td>54,000</td>
<td>56,000</td>
<td>126</td>
<td>61,000</td>
</tr>
<tr>
<td>Total tocopherol, μg/g fat</td>
<td>75.4</td>
<td>655.7</td>
<td>408.7</td>
<td>530.9</td>
<td>528.4</td>
</tr>
<tr>
<td>Alpha, %total</td>
<td>37.7 (50.0)</td>
<td>516.6 (78.8)</td>
<td>308.6 (75.5)</td>
<td>51.7 (9.7)</td>
<td>316.9 (60.0)</td>
</tr>
<tr>
<td>Beta, %total</td>
<td>16.0 (21.2)</td>
<td>14.3 (20.8)</td>
<td>9.9 (2.4)</td>
<td>17.0 (3.2)</td>
<td>10.0 (1.9)</td>
</tr>
<tr>
<td>Gamma, %total</td>
<td>19.7 (26.1)</td>
<td>123.2 (18.8)</td>
<td>89.4 (21.9)</td>
<td>372.8 (70.2)</td>
<td>194.3 (36.8)</td>
</tr>
<tr>
<td>Delta, %total</td>
<td>2.0 (2.7)</td>
<td>1.0 (0.2)</td>
<td>0.7 (0.2)</td>
<td>89.4 (16.8)</td>
<td>7.2 (1.4)</td>
</tr>
</tbody>
</table>

RB, regular butter; VRB, regular butter and 50% vegetable oil mix 1; VLCB, low cholesterol butter and 50% vegetable mix 1; LCSB, low cholesterol and low saturated fat butter; VM, vegetable mix 2. *Vegetable oil mix 1, made with 50% canola oil, 30% high oleic sunflower oil, 20% sunflower oil. †Vegetable oil mix 2, made with 30% canola oil and 70% palm oil.

analyzed by fast-GC (Perkin-Elmer, model Clarus 600, Les Ulis, France) (11).
purpose was to select the most relevant variables relative to atherogenic status (step 1, PLS methods), to aggregate these variables into biological clusters according to statistical or ontological considerations (step 2), to evaluate the impact of the dietary treatment on each biological cluster (step 3), and to display the concerted action and the contribution of each biological cluster to the modulation of atherosclerosis (step 4).

RESULTS

Diets and Atherogenic Status

There were no differences in weight gain among the hamsters fed the various diets at the end of the nutritional trial (Table 2). The liver weights were slightly lower in the hamsters deprived of cholesterol in their diet (Fig. 2A). Conversely, the atherogenic outcomes were dramatically affected by the lipid diets given to the hamsters (Fig. 2B). This atherogenic outcome was defined as the ratio of total cholesteryl-ester to total phospholipids in the aorta (see MATERIALS AND METHODS). In fact, the atherogenic status obtained with the experimental fat diets (VRB, VLCB, and LCSB with athero indexes of 4.47, 4.26, and 3.35, respectively) were framed by the highest values found in the RB hamster group (athero index of 6.34) and the lowest values found in the VM hamster group (athero index of 1.86). It should be noted that all of the hamsters were atherosclerotic compared with hamsters fed a low-fat chow diet for the same period and used as “no-atherosclerotic” reference individuals (athero index of 0.99, not shown). Atherogenicity was unrelated to the current intake of phytosterols in our study (Table 1 and Fig. 2, A and B). Conversely, a lower atherogenic status was obtained by decreasing by half the total saturated fatty acids and cholesterol content in the diet (VRB diet), which was accomplished by diluting the RB with canola oil. Further lowering cholesterol intake to 80% (VLCB) by removing the cholesterol from dairy fat while keeping the other nutrients, comparable to the VRB diet did not significantly improve the atherogenic outcome. However, using a technological process to decrease both the saturated fatty acids and cholesterol (LCSB) similarly was more efficient in improving the atherogenic status (−12.5%; P < 0.05; Fig. 2B).

Biological Analyses and Atherogenic Status

We also performed a multiplatform analysis of hamster biofluids and tissues to obtain better insights into the underlying mechanisms that might be associated with the severity of atherosclerosis. We performed untargeted metabolomics on urine and plasma samples, along with fatty acid analysis of plasma lipid classes and total liver lipids, analysis of total lipid classes in the whole aorta, analysis of the expression of 44 target genes in the liver and in whole blood involved in the metabolic control associated to atherogenic status (Supplemental Fig. S1), and analysis of conventional blood chemistry to assess the blood lipoprotein particles and lipid contents (Fig. 1). These analyses generated a set of 1,666 variables per hamster. Our statistical model retained 87 variables strongly correlated with atherosclerosis (Supplemental Table S2). LC-ESI-HRMS, liquid chromatography-electrospray ionization-high resolution mass spectrometry; GC-FID, gas-chromatography flame ionization detection; HCA, hierarchical clustering analysis; QPCR, quantitative polymerase chain reaction; PLS; projection on latent structure; OPLS-DA, orthogonal projection on latent structure discriminant analysis.

Fig. 1. Analytical and statistical workflow. To facilitate data processing and interpretation, the hamsters were stratified according to the median of their atherogenic index values (total cholesteryl-ester content relative to structural phospholipids in the whole aorta), into low or high atherogenic status groups. An orthogonal partial least square discriminate analysis (PLS-DA), using low or high atherogenic status as a class variable, was applied to each variable category (blocks), e.g., either liquid chromatography-mass spectrometry features in urine or in plasma, fatty acids both in liver and plasma, plasma biochemistry, or liver and blood gene expression. The combined model obtained after applying selected variables to individual variable categories was highly significant, with elevated explained variance of both observations and classes (Supplemental Table S3). From the initial 1,666 variables, this model, after curation for redundancies and artifacts, finally retained 87 variables significantly correlated with atherosclerosis (Supplemental Table S2). LC-ESI-HRMS, liquid chromatography-electrospray ionization-high resolution mass spectrometry; GC-FID, gas-chromatography flame ionization detection; HCA, hierarchical clustering analysis; QPCR, quantitative polymerase chain reaction, PLS; projection on latent structure; OPLS-DA, orthogonal projection on latent structure discriminant analysis.
Physiological outcomes of hamsters fed different diets after 12 wk of nutritional challenge

<table>
<thead>
<tr>
<th>Physiological Outcome</th>
<th>RB</th>
<th>VRB</th>
<th>VLCB</th>
<th>LCSB</th>
<th>VM</th>
</tr>
</thead>
<tbody>
<tr>
<td>Initial weight, g</td>
<td>66 ± 9</td>
<td>66 ± 10</td>
<td>67 ± 7</td>
<td>67 ± 9</td>
<td>67 ± 9</td>
</tr>
<tr>
<td>Weight after 10 wk of dietary challenge, g</td>
<td>133 ± 12</td>
<td>131 ± 10.2</td>
<td>131.8 ± 10.4</td>
<td>133 ± 7.7</td>
<td>137.6 ± 8.1</td>
</tr>
<tr>
<td>Liver weight, g</td>
<td>7.1 ± 0.7b</td>
<td>7.1 ± 0.3b</td>
<td>6.2 ± 0.7a,b</td>
<td>6.5 ± 1.0a,b</td>
<td>5.8 ± 0.4a</td>
</tr>
<tr>
<td>HDL cholesterol, mmol/l</td>
<td>1.4 ± 0.3c</td>
<td>2 ± 0.2a</td>
<td>1.7 ± 0.2a</td>
<td>3.2 ± 0.2a</td>
<td>1.5 ± 0.2a</td>
</tr>
<tr>
<td>Plasma cholesterol, mmol/l</td>
<td>2.5 ± 0.5</td>
<td>1.5 ± 0.2</td>
<td>1.9 ± 0.3</td>
<td>1.8 ± 0.3</td>
<td>1.4 ± 0.4</td>
</tr>
<tr>
<td>Nonfasted (chow 1.4)</td>
<td>8.1 ± 0.2c</td>
<td>5.6 ± 0.3b</td>
<td>5.8 ± 0.2b</td>
<td>5.2 ± 0.2a,b</td>
<td>4.5 ± 0.3a</td>
</tr>
<tr>
<td>Fasted</td>
<td>8.2 ± 0.5b</td>
<td>6.2 ± 0.4a</td>
<td>5.8 ± 0.2a</td>
<td>5.3 ± 0.2a</td>
<td>5.3 ± 0.2a</td>
</tr>
<tr>
<td>HDL cholesterol, mmol/l</td>
<td>5.1 ± 0.4b</td>
<td>3.3 ± 0.3a</td>
<td>3.8 ± 0.3a</td>
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<td>3.9 ± 0.1a</td>
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<td>Nonfasted (chow 2.5)</td>
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<td>3.9 ± 0.4a</td>
<td>4.6 ± 0.2a,b</td>
<td>3.5 ± 0.2a</td>
<td>3.5 ± 0.3a</td>
</tr>
<tr>
<td>Fasted</td>
<td>2.9 ± 0.3b</td>
<td>2.1 ± 0.2b</td>
<td>2.1 ± 0.1b</td>
<td>2.2 ± 0.2b</td>
<td>0.8 ± 0.1a</td>
</tr>
<tr>
<td>Non-HDL cholesterol, mmol/l</td>
<td>2.6 ± 0.3b</td>
<td>2.1 ± 0.5a,b</td>
<td>1.2 ± 0.1a</td>
<td>1.8 ± 0.2a,b</td>
<td>1.8 ± 0.2a,b</td>
</tr>
</tbody>
</table>

Values are means ± SD; n = 8 hamsters per group. Different letters on the same line indicate a statistical significance (Bonferroni-Dunn test). Chow values correspond to plasma values measured in 2 chow-fed hamsters for the same period.

Table 2.

Physiological outcomes of hamsters fed different diets after 12 wk of nutritional challenge

Associated with the atherogenic index, including some metabolomics features that remained unidentified (Supplemental Table S2).

Statistical Modeling of Diatological Clusters to Predict the Atherogenic Status

To consider the complex dimensions of atherosclerosis, we visualized the relationships among the 87 variables that found to be associated with the disease, using an interaction network. The network self-organized around the atherogenic index surrounded by different regions of variables. These regions of variables, in turn, defined 10 biological clusters, as described in Supplemental Fig. S2 and Supplemental Table S2.

To facilitate data interpretation while keeping all of the significant information, we modeled the 10 biological clusters into numerical biological scores as we already reported else-

Fig. 2. Prediction of atherosclerosis with selected biological clusters. Observed atherosclerosis is expressed as a percentage of relative total cholesteryl-ester deposition in the aorta (to total phospholipids) in the hamsters with low or high atherogenic status (A) or in the hamsters fed the various diets (B). Corresponding predicted atherosclerosis is shown in C and D. Predicted atherosclerosis is calculated from the PLS equation: pred athero = 0.108091 × [dairy fat derived fatty acids] + 0.152669 × [endogenous derived fatty acids] - 0.0340185 × [mitochondron function] + 0.173429 × [regul. lipid metabol. transport] - 0.0751566 × [vitamin E metabolism] - 0.153142 × [hemostasis] - 0.110269 × [amino acid metabolism] + 0.110269 × [blood cholesterol related] + 0.231903 × [inflammation] - 0.00456344 × [miscellaneous and unidentified] + 0.46731. E: heat map of the quantitative contribution of each biological score to the atherogenic index. The results shown are from the hamsters with either low or high atherogenic status or in the hamsters fed the various diets. Red color indicates increased values, and green color indicates decreased values. See Table 3 for corresponding numerical values; n = 8 hamsters per group. Dietary fat correspondence: RB, regular butter; VRB, regular butter and 50% vegetable oil mix; VLCB, low cholesterol butter and 50% vegetable mix; LCSB, low cholesterol and saturated butter; VM, vegetable mix. Groups sharing a different lowercase letter (a, b, c, e) are significantly different (P < 0.05, Tukey post hoc test after ANOVA).
where (56), using a validated mathematical method (64) that is
detailed in the supporting material. Each biological score can
be observed as a multiplex variable computed from the original
variables, defining the biological clusters. Such a strategy,
using multi-omics data assembled into biological modules, was
recently published when this article was under revision (63).

With the use of the scores of the biological clusters, it was
possible using the PLS algorithm to calculate a predictive
equation of the atherogenic index for each hamster, as follows:

\[
\text{predicted atherogenicity} = 0.108091 \times \text{[dairy fat derived fatty acids]} + 0.152669 \times \text{[endogenous derived fatty acids]} - 0.0340185 \times \text{[mitochondrion function]} + 0.173429 \times \text{[regul. lipid metabol. trans.]} - 0.0751566 \times \text{[vitamin E metabolism]} - 0.153142 \times \text{[hemostasis]} - 0.110269 \times \text{[amino acid metabolism]} + 0.110269 \times \text{[blood cholesterol related]} + 0.231903 \times \text{[inflammation]} - 0.00456344 \times \text{[miscellaneous and unidentified]} + 4.06731. \]

We previously applied this type of predictive equation to cardiovascular risk using gene single
nucleotide polymorphism in humans (10). When considering
a hamster with a low or a high atherogenic status, the
predicted atherosclerosis level calculated from the equation
above was similar to the observed atherosclerosis level (Fig.
2, A and B). The quantitative contribution of each biological
cluster to the atherogenic index around this value and across
the experimental diets thus could be easily calculated using
the equation (Fig. 2E and Table 3 for corresponding numeri-
cal values).

Integration of the Dietary Challenges, the Interplay Between
the Biological Clusters, and the Atherogenic Status Using an
Interaction Network

To gain further insight into the concerted interactions of the
biological clusters to explain atherosclerosis, we calculated the
partial correlations matrix among all of the cluster score
values, together with the atherogenic index. The matrix was
mapped as a network, and we focused on the most relevant
interactions using \( q \) values \( \leq 0.10 \) after false discovery rate
filtering, to account for multiple comparisons (Fig. 3). This
process yielded a very sparse network that was topologically
divided in two parts, in which one could identify three impor-
tant nodes having a hub position in the network, namely
regulation of lipid transport and metabolism, inflammation, and
amino acid metabolism. The node most statistically connected
to atherosclerosis was regulation of lipid transport and metab-
olism, including SREBP1c as a regulating nuclear receptor and
some of its target genes (ACC, SDC1, MTP, and LDL-r). The
inflammation node connected the two parts of the network
together. Amino acid metabolism also appeared as the main
crossroad in the graph. In addition, we constructed a heat map
to show how the individual biological cluster components were
affected by the dietary treatments and which individual com-
ponent was the most related to atherosclerosis (determined by
their PLS-discriminant analysis-variable importance value).

DISCUSSION

The main goal of our study was to investigate the potential
of various dairy fat design strategies to modulate early ather-
sclerosis. To examine the biological impact of the diets,
we implemented a specific systems biology workflow that allowed
for identifying and quantifying the contribution of biological
sets associated with the disease and sensitive to dairy fat
quality. For these purposes, we used the atherosclerosis-prone
hamster model. This model shares similarity with humans in
terms of cholesterol metabolism and has thus been frequently
used in studies examining cholesterol-related cardiovascular
diseases (18, 33, 40, 55, 59, 60).

Justification of the Dietary Design and Atherogenic Impact
of Diets

Three test fats were evaluated, bounded by a reference
dairy fat (regular butter) and a plant oil mixture, both of
which were designed not to diverge too much from the test
fats. The test fats were formulated to maintain the total
saturated fat content at \(<40\%\) and the n-6/n-3 polyunsatu-
rated fatty acid ratio at \(\sim 5\). It must be emphasized that our
protocol was not designed, nor was it able, to examine the
atherogenic activity of individual saturated fatty acids of
various chain lengths but rather of total saturated fat. To
determine possible interactions with dietary cholesterol, the
fats were free of cholesterol or contained gradual amounts of
cholesterol up to natural level (Table 1). It should be
noted that the maximum amount of cholesterol in the test
diets (0.05\%) was usually less than what has commonly
been used in similar hamster studies [0.12\% (39) up to 3%
(24)], making difficult strict comparisons with such studies.
We chose these cholesterol amounts because they were
similar to physiological conditions and we found them
suitable for such a trial (33).

The three technologically modified dairy fats induced an
intermediary atherosclerotic status (e.g., \%aorta cholesteryl
ester to phospholipid ratio) compared with nonprocessed dairy
fat (RB, higher atherosclerosis) or to the oil mixture (VM,
lower atherosclerosis). The decholesterolized and desaturated
dairy fat (LCSB) performed the best among the transformed
dairy fats.

Test-Fat-Specific Nutrient Content and Atherogenic Status

To explain our findings further, we evaluated whether the
various methods used to change the fatty acid and cholesterol
content in our test fats also changed the content of other
important “anti”-atherogenic nutrients, such as phytosterols or
polyunsaturated (PUFA) C18 fatty acids, thus influencing the
atherogenic status. If indeed the technological strategies that
we used modified the phytosterol and PUFA content of the
diets, these modifications were not obviously correlated with
the atherogenic outcome. For instance, within a similar content
of cholesterol, LCSB fat contained 400-fold less phytosterols
and 3.4-fold less PUFA than VLCB but was still less athero-
genic.

Overall, the lower atherogenic impact of the LCSB diet
could seem paradoxical compared with the other dairy fat diets,
particularly the VLCB diet, which is very close in composition
but exhibited comparatively more atherogenic features. A dif-
ference in the amount of oxidative by-products in the diet
cannot be argued to explain that difference. In fact, whereas we
found lipoperoxidative fatty acids easily detectable under our
LC-MS conditions (49), such compounds that could have
occurred from oxidized diets were not found to be associated
with atherosclerosis in the present study. One possibility is that
the LCSB fat has higher levels of rumenic acid (2.5- to 5-fold

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Table 3. Quantitative contributions of individual clusters to the atherogenic index values (expressed as relative %cholesterol esters to total aortic phospholipids) according to the atherogenic status around the background level of atherosclerosis calculated from the predictive PLS equation

<table>
<thead>
<tr>
<th>Clusters</th>
<th>Low1</th>
<th>High1</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blood cholesterol related</td>
<td>-0.14</td>
<td>-0.15</td>
</tr>
<tr>
<td>Endothelium-derived</td>
<td>0.17</td>
<td>0.18</td>
</tr>
<tr>
<td>Dietary fat-derived factors</td>
<td>0.04</td>
<td>0.08</td>
</tr>
<tr>
<td>Resultant lipid transport and</td>
<td>0.05</td>
<td>0.06</td>
</tr>
<tr>
<td>Inflammation</td>
<td>0.01</td>
<td>0.02</td>
</tr>
<tr>
<td>Mitochondrion-related</td>
<td>0.04</td>
<td>0.06</td>
</tr>
<tr>
<td>Oxidation stress</td>
<td>0.00</td>
<td>0.00</td>
</tr>
</tbody>
</table>

Values are expressed as the means ± SD. Compared with the control diet, the atherogenic status was divided into two subgroups: Low1 and High1. The atherogenic status was calculated from the predictive PLS equation to 3.270.32±2.47 on July 3, 2017 http://ajpheart.physiology.org/ Downloaded from}

Modeling into Scores the Biological Events Related to the Atherogenic Status Across the Dietary Treatments

To investigate further how the atherogenic status was achieved across the dietary treatments, we determined which biological events were associated with the modulation of the disease by dairy fats. The heterogeneity in the origins of the 82 variables found to be associated with atherosclerosis provided a good illustration of the multifactorial basis of the disease that we intended to capture. To facilitate data interpretation and reporting, we aggregated the variables into 10 biological sets or clusters (Figs. 1 and 2 and Supplemental Table S2). When introduced into a PLS model, the 10 biological clusters described up to 81% of the atherogenic index variability [p = 0.903, R² = 3.27 × 10⁻⁷ after cross validated-ANOVA, ROC-area under the curve (AUC) > 0.90]. In comparison, total fasted plasma cholesterol, a well-validated biomarker of atherosclerosis, was only associated with cholesterol content (p = 0.53; non-HDL/HDL-C ratio: p = 0.07, ROC-AUC = 0.36) or nonfasted plasma cholesterol values (total cholesterol: p = 0.72, P < 0.0001, ROC-AUC = 0.84; non-HDL-C: p = 0.66, P < 0.0001, ROC-AUC = 0.72; non-HDL/HDL-C ratio: p = 0.33, P = 0.07, ROC-AUC = 0.60). Indeed, assessing all of the fasted and nonfasted plasma cholesterol values into a PLS predictive atherogenic score did not perform as well as the 10 biological clusters (p = 0.76, P = 0.000024, ROC-AUC = 0.84).
Our quantitative approach, based on the biological clusters, also allowed for an interesting stratification of the relative contributions of complex biological events to the development of the disease. For instance, we found the four lipid-related bioclusters to be the main contributors to the atherogenic index, whether positively or negatively (Fig. 2 and Table 3). The amino acid and vitamin E metabolism bioclusters were secondary. For the latter, consistent with our finding of an inverse relationship between vitamin E metabolites and atherosclerosis, the vitamin E biocluster was the most quantitative contributor to atheroprotection in the LCSB-fed hamsters. All of these clusters were sensitive to the dietary fat quality, whereas the remainder of the other bioclusters were not (mitochondrion function, hemostasis, inflammation, miscellaneous, and unidentified). Beyond our study, such an approach could be relevant for choosing the most efficient strategy in situations of multitarget therapy and eventually in assessing the efficacy of therapy.

Interplay Between the Biological Clusters Associated to the Atherogenic Atatus

Interestingly, the regulation of lipid metabolism and transport biocluster, consisting exclusively of gene transcripts, was the main quantitative contributor to the atherogenic index (16%) among the hamsters of high or low atherogenic status, and it was especially greatly impacted by the RB diet (Table 3). It was also directly connected the atherogenic index (e.g., %CE to PL ratio in the aorta) and to the remainder of the other bioclusters within the partial correlation interaction network (Fig. 3). This partial correlations network was efficient in detecting highly regulatory functions with hub positions (1), and it possesses a topology that could correspond to metabolic reactions (28). This biocluster thus appeared central to the concerted action of multiple biological events that led to atherosclerosis development, with an impact that was strongly correlated with dietary fat quality. Notable was that stearoyl CoA-desaturase I (SCD1) gene expression had greater influence in determining the score of that biocluster and was also correlated with many variables arising from other bioclusters (Supplemental Fig. S2). In contrast, SCD1 was also coregulated with other lipogenic genes regulated by the nuclear receptor SREBP1c, such as ACC, LDL-r, and MTP (Fig. 3), as well as other genes involved in lipid export from the liver (supplemental Fig. S2 and Fig. 3). Thus nonprocessed dairy fat (RB diet) fed in excess caused an overflow of circulating lipids that was aggravated by the triggering of endogenous lipogenesis, which ultimately led to more severe atherosclerosis. This process was illustrated well by the proximity with the other
lipid biological clusters in hierarchical clustering analysis (blood cholesterol-related, endogenous-derived fatty acids, and dairy fat-derived fatty acids; Fig. 2E). Interestingly, such overflow was not observed with the transformed dairy fats and the plant oil mix control, despite the fat and energy intake being similar, underlying the qualitative impact of fats. Our finding also emphasized that the global regulatory role of SCD1 gene extended far beyond lipid metabolism, as demonstrated elsewhere (9), which was well illustrated in our correlation network (Supplemental Fig. S2).

Data Integration with the Dietary Challenge and the Atherogenic Status

Our results also stressed the relevance of decreasing both cholesterol levels (and/or availability) and saturated fatty acids to lower their atherogenic potential under conditions of high intake. In such inflammationist intake conditions, the lipid overflow shifts the medium-chain saturated fatty acids lauric acid (C12:0) and myristic acid (C14:0) from neutral to atherogenic. Flooding the liver with fatty acids, such as the C14:0 compound, specifically inhibits cholesterol esterification and thus alters the distribution of excess cellular cholesterol in the liver between the storage pool of cholesteryl esters and the metabolically active, regulatory pool, as nicely explained some years ago (53). The link of such flooding with atherogenic status was well illustrated in our study by the association we found between the lipid species exhibiting dairy fat-derived fatty acids and atherosclerosis, particularly those esterifying myristic acid (Supplemental Fig. S2 and Fig. 3). In addition, the excess could also be responsible for some of the defects of the mitochondrion function biocluster, illustrated by the connection of this biocluster to the regulation of lipid transport and metabolism cluster in the interaction network. Hence, some of its components, i.e., hydroxybutyryl-, octanoyl-, and tiglyl-carnitines, associated with fatty acid oxidation and ketone body production in the mitochondria (13, 26), were higher in the more atherogenic hamsters. These metabolites likely reflected the defect in the management of lipid overflow discussed above through insufficient lipids. This downregulation of the UCP2 gene could also be viewed as a failure in mitochondrial fatty acid metabolism, contributing to atherosclerosis development (6). Additionally, butyryl/isobutyryl- and isovaleryl-, as well as tiglyl-carnitine, would reflect defects in branched-chain amino acids mitochondrion metabolism (leucine, isoleucine and valine) (13, 22, 26).

Finally, if we refer to LCSB hamsters, their acylcarnitine profile is close to that found in the highly atherogenic RB hamsters. However, the LCSB hamsters were poorly atherogenic. This proatherogenic feature was thus balanced by other regulations that could counteract such mitochondrial defects, exemplifying well how complex diseases arose when multiple disruptions remained unrepairred or were not offset and relying on a small set of biomarkers can be insufficient to conclude (34).

Additionally, beyond the impact on cholesterol metabolism, we also found that the dairy fat-derived fatty acids accumulated in lysoPCs in the blood, in correlation with the severity of atherosclerosis. In addition to reflecting dairy food intake (38), perhaps such lipid species esterifying medium-chain saturated fatty acids play a specific role in the progression of atherosclerosis, as reported with plasma long-chain lysoPCs (15, 44, 54). This possibility certainly requires a precise evaluation. One of the mechanisms of lysoPC signaling occurs through interaction with either a uncharacterized receptor and/or the platelet-activating factor receptor in the monocytes, giving rise to arachidonic acid release and the synthesis of proinflammatory mediators (43). Along with lysoPCs, our untargeted metabolomics analysis also revealed platelet-activating factors (lyso-PAF-O-C16:0 and PAF-O-C17:0/2:0) that were decreased in the low atherogenic status hamsters (Fig. 2E). As mentioned above, along with lysoPCs, such lipid messengers could also be involved in the progression of atherosclerosis. Nevertheless, not all PAF species exert the same biological response, and the one bearing the margaric alkyl (C17:0 alkyl) that we found has not yet been documented.

The amino acid metabolism biocluster formed a cross roads in the interaction network, underlying its significant importance in metabolic regulation, especially in relating vitE metabolism to inflammation. This statistical connection could illustrate the importance of unappreciated interplays between metabolites and/or biological clusters in the development of a complex disease such as atherosclerosis. Our results were also consistent with findings in humans relating branched-chain amino acids (amino acids biocluster) and their acylcarnitine products (mitochondrion function biocluster) to arterial disease (5), although the significance of the association could be different from that in our study.

We found within the inflammation biocluster that IL1a and CRP genes expression was upregulated in the least atherogenic high plant oil fed hamsters (VM) compared with the other dairy fat fed hamsters and the antioxidant PON1 was downregulated. This finding could seem paradoxical because such a profile is clearly proatherogenic. However, dairy fat intake has also frequently been associated with improvement of the inflammatory and oxidative status in obese subjects (27, 68), which could be consistent with our observations.

In conclusion, we found that, under conditions of atherogenic intake, the impact of dairy fats on the disease (RB diet) could be dramatically reduced by technological processing, such as by decreasing cholesterol and total saturated fatty acids (VRB, VLCB, and LCSB diets) and, further, by augmenting the level of antiatherogenic nutrients, such as CLA (59) and more likely γ-tocopherol (LCSB diet) (69). We thus additionally demonstrated that dairy fats cannot be considered monolithically with regard to cardiovascular diseases and that quality improvements with regard to this outcome could be achieved. For instance, marketing products including dairy fats, such as the LCSB, could provide health benefits for consumers or at least could help in the setting of a prudent diet. Very recently, such a strategy, using a canola oil-modified butter as part of a Mediterranean diet, was found to be quite effective for improving blood lipids in a randomized, controlled study of overweight individuals (4).

In our context, the atherogenic impact of individual saturated fatty acids could not be formally revealed, and PUFA levels did not seem to exert obvious effects. Of course, the hyperlipidemic hamster does not represent the general features of atherosclerosis and restricted our conclusions to these specific settings. Other animal models with genetically induced atherosclerosis could yield complementary information with regard to the background biological events underlying the disease outcome.

When comparing the various biological components related to our atherosclerosis conditions, it was possible to distinguish factors that were sensitive to dairy fat composition from those...
that were not or that were less sensitive to dairy fats. This distinction is important to know when addressing complex multifactorial diseases to choose the most appropriate and efficient dietary intervention strategy. From this perspective, we believe that the integrated analysis we performed, which allowed for the calculation of a predictive equation consisting of biological clusters, could be a powerful strategy for targeting the most relevant factors. For instance, we identified the cluster regulation of lipid metabolism and transport, including SCD1, as the main driver of the atherogenic status across our dietary settings, along with other biological events. The accuracy of our analysis could be further improved by even wider coverage of the “omics” dimensions. A very recent human study, using a similar multi-omics biological module concept applied in the context of cardiometabolic risk, was published when our article was in revision (63). This study and ours were the first examples, to our knowledge, that allowed for the precise quantitative contribution of various biological sets to a defined disease outcome. They illustrated how such tools could be constructed to identify, quantify, and stratify specifically the contributions of various components to the development of a complex disease such as atherosclerosis. This ability could be advantageous in situations of growing multimorbidity (2). Therefore, we consider our results to be very encouraging for the identification and validation of multiplex predictors, instead of single predictors of cardiovascular disease risk (50).

GRANTS
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DISCLOSURES
D. Dalemans is employed by Corman, which provided the transformed dairy fats and partially financed the study.

AUTHOR CONTRIBUTIONS

REFERENCES


Nestel PJ. 34.


