Physiological and pathological left ventricular hypertrophy of comparable degree is associated with characteristic differences of in vivo hemodynamics

Attila Oláh, Balázs Tamás Németh, Csaba Mátyás, László Hidi, Árpád Lux, Mihály Ruppert, Dalma Kellermayer, Alex Ali Sayour, Lilla Szabó, Marianna Török, Anna Meltzer, László Gellér, Béla Merkely,* and Tamás Radovits*

Heart and Vascular Center, Semmelweis University, Budapest, Hungary

CARDIAC HYPERTROPHY HAS BEEN referred to as an increase of cardiac mass by cardiomyocyte enlargement, which can develop in response to increased load of the heart. Pathological and physiological cardiac hypertrophies are caused by different stimuli and are associated with distinct structural and molecular phenotypes (1).

When pressure or volume overload is induced by diseases (e.g., hypertension, valvular disorders), it results in pathological hypertrophy, which is initially a compensatory response to the increased biomechanical stress in an attempt to maintain normal cardiovascular function (24). Pathological hypertrophy is also associated with loss of cardiomyocytes (apoptosis and necrosis), increased interstitial fibrosis, and the reactivation of fetal cardiomyocyte gene program (1, 9). This condition can eventually decompensate leading to ventricular dilatation, which, over time, may progressively lead to systolic and diastolic myocardial dysfunction (heart failure). With the rising incidence among the aging population, heart failure has remained a leading cause of death, and this major health problem facilitated numerous investigations to identify new therapeutic targets to prevent or reverse pathological cardiac hypertrophy (25).

Long-term exercise training also induces a cardiac hypertrophy response, called athlete’s heart. This balanced increase of myocardial mass is associated with preserved or even enhanced cardiac function without cardiac fibrosis, cardiomyocyte apoptosis, or changes in fetal gene expression (9, 24). Furthermore, the protective role of exercise in cardiovascular prevention and treatment has been widely recognized (35, 41). The use of physiological stimuli can be one potential therapeutic strategy to inhibit pathological processes leading to heart failure and sudden cardiac death (23).

Despite some morphological similarities, pathological and physiological cardiac hypertrophy are induced by substantially distinct signaling pathways and characterized by different gene and protein expression profile (1, 16, 43). Although numerous studies have investigated these distinctions, the functional differences between pathological and physiological hypertrophy are poorly described. So far noninvasive approaches have been used to distinguish pathological from physiological left ventricular (LV) hypertrophy in experimental animals (8, 24) and in human subjects (7, 15). These works describe several aspects of LV function in the two forms of hypertrophy; however, most of these noninvasively measured indexes are dependent on loading conditions, and these characterizations are not detailed enough to deeply understand LV mechanics. Assessment of LV pump properties can be fundamental to advance the understanding of cardiovascular pathophysiology and therapeutics. Pressure-volume analysis provides the most comprehensive method to evaluate in vivo cardiac systolic and diastolic function.
diastolic function as well as mechanoeffectives, less affected by arterial and venous loading conditions and heart rate (HR) (30). Advances in miniature sensor technology established this approach to be successfully translated to rodents; thus LV pressure-volume analysis has become standard in basic cardiovascular research for detailed characterization of cardiovascular function (14).

The aim of this study was to give a direct, reliable hemodynamic comparison of exercise- (physiological) and pressure overload-induced (pathological) LV hypertrophy using the sophisticated method of LV pressure-volume analysis in two widely used, well-established rat models.

MATERIALS AND METHODS

Ethical approval and animals. All animals received humane care in compliance with the Principles of Laboratory Animal Care formulated by the National Society for Medical Research and the Guide for the Care and Use of Laboratory Animals prepared by the Institute of Laboratory Animal Resources and published by the National Institutes of Health (Publication No. 85-23, revised 1996). All procedures and handling of the animals during the study were reviewed and approved by the Ethical Committee of Hungary for Animal Experimentation.

Young adult male Wistar rats (n = 41, m = 200–250 g; Toki-Coop, Dunakeszi, Hungary) were housed in a room with constant temperature of 22 ± 2°C with a 12-h/12-h light-dark cycle and fed standard laboratory rat diet ad libitum and free access to water.

Swimming training: rat model of physiological cardiac hypertrophy. For long-term exercise training, after an adequate adaptation, exercised (Ex; n = 12) rats swam for a total period of 12 wk, for 200 min/day, 5 days a week as previously described (28). Untrained control (Co; n = 11) rats were placed into the water for 5 min each day during the 12-wk-long training program.

Abdominal aortic banding: rat model of pathological cardiac hypertrophy. Pressure overload-induced cardiac hypertrophy was produced by abdominal aortic banding (AB; n = 10). All surgical procedures were performed under anesthesia with pentobarbital sodium (60 mg/kg) administered by intraperitoneal injection. Abdominal aortic constriction was performed as described before (17), with some modification. Briefly, after proper anesthesia, a midline laparotomy was performed. The abdominal aorta was constricted at suprarenal level by tightening a 2-0 surgical suture against a blunted 20-gauge needle, which was pulled out thereafter. The abdominal musculature and the skin incisions were closed by standard surgery techniques. Sham-operated rats (Sham; n = 8) served as controls and underwent an identical procedure, but without ligation of abdominal aorta. Thereafter, animals were observed for a 6-wk period after surgical processes.

Echocardiography. At the completion of swimming training program or 6 wk after abdominal aortic banding procedure, LV morphological alterations were observed by echocardiography as described before (21). Rats were anesthetized with pentobarbital sodium (60 mg/kg ip). Animals were placed on controlled heating pads, and the core temperature, measured via rectal probe, was maintained at 37°C. A polyethylene catheter was inserted into the left external jugular vein for fluid administration. A 2-Fr pressure-conductance microphone (SPR-838; Millar Instruments, Houston, TX) was inserted into the right carotid artery and advanced into the ascending aorta. After stabilization, aortic pressure curves were recorded to calculate mean arterial blood pressure (MAP).

The catheter was thereafter advanced into the LV under pressure control. After stabilization for 10 min, baseline P-V relations were recorded. Signals were continuously registered at a sampling rate of 1,000 samples/s using a P-V conductance system (MPVS-Ultra; Millar Instruments) connected to the PowerLab 16/30 data acquisition system (AD Instruments, Colorado Springs, CO), stored and displayed on a personal computer by the LabChart 7 Software System (AD Instruments). With the use of a special P-V analysis program (PVAN; Millar Instruments), HR, LV end-systolic pressure (LVESP), LV end-diastolic pressure (LVEDP), the maximal slope of LV systolic pressure increment (dP/dtmax) and diastolic pressure decrement (dP/dmin), time constant of LV pressure decay (τ; according to the Glantz method (30)), LV end-diastolic volume (LVEDV), LV end-systolic volume (LVESV), SV, EF, cardiac output (CO), and stroke work (SW) were calculated and corrected according to in vitro and in vivo calibration of the SPR-838 software using the total peripheral resistance (TPR) was calculated by the following equation: TPR = MAP/CO. To exclude the influence of body weight differences, CO was normalized to body weight [cardiac index (CI)].

In addition to the above parameters, P-V loops recorded at different preloads can be used to derive useful indexes of LV function that are less influenced by loading conditions and cardiac mass. Therefore, LV P-V relations were measured by transiently compressing the inferior vena cava (reducing preload) under the diaphragm with a cotton-tipped applicator. The slope of the LV end-systolic P-V relationship (ESPVR; according to the parabolic curvilinear model), preload recruitable SW (PRSW), and the slope of the dP/dmax-end-diastolic volume relationship (dP/dmax-EDV) were calculated as load-independent indexes of LV contractility. The slope of the LV end-diastolic PV relationship (EDPVR) was calculated as a reliable indicator of LV stiffness (18).

Cardiac mechanoeffective parameters were calculated as suggested by Suga (37) and Sunagawa et al. (39). SW and P-V area (PVA) were calculated by the PVAN software, and LV mechanical efficiency was calculated as SW/PVA. Arterial elastance (Ea) was calculated as LVESP/SV. Ventriculoarterial coupling (VAC) was described by the quotient of Ea and ESPVR (44).

At the end of each experiment, 100 μl of hypertonic saline were injected intravenously, and from the shift of volume relations, parallel conductance was calculated by the software and used for the correction of the LV volume values. The volume calibration of the conductance system was performed as previously described (30). Shortly, nine cylindrical holes in a block 1 cm deep and with known diameters...
ranging from 2 to 11 mm were filled with fresh heparinized whole rat blood. In this calibration, the linear volume-conductance regression of the absolute volume in each cylinder versus the raw signal acquired by the conductance catheter was used as the volume calibration formula.

To remove erythrocytes from myocardial tissue, an in vivo perfusion was performed. After the thoracic cavity was opened and the inferior caval vein in the thorax was dissected, a total volume of 40 ml oxygenated Ringer solution (37°C) was infused into the LV through the apex of the heart.

Thereafter, the heart was quickly removed and placed into cold (4°C) Ringer solution. Heart weight was measured, and LV myocardial tissue samples were collected immediately for histology and molecular biology. Subsequently, post-mortem TL measurements were done.

**Histology.** Hearts were harvested immediately after animals were euthanized. Heart tissue samples were fixed in buffered paraformaldehyde solution (4%) and embedded in paraffin. Transverse transmural slices of the ventricles were sectioned (5 μm), deparaffinized, and processed conventionally for histological examination. The sections were stained with methoxylin-eosin (HE) and picrosirius red staining. Light microscopic examination was performed with a Zeiss microscope (Axio Observer.Z1; Carl Zeiss, Jena, Germany), and digital images were captured using an imaging software (QCapture Pro 6.0; QImaging, Canada).

On the HE-stained LV myocardial sections, transverse transmural widths of randomly selected, longitudinally oriented cardiomyocytes were measured by a single investigator (with a magnification of 400×). The mean value of 100 LV cardiomyocytes represents each sample.

To investigate myocardial collagen content, picrosirius red positive area was calculated using ImageJ (National Institutes of Health, Bethesda, MD) image analysis software. Five subendocardial and five subepicardial images (magnification 200×) were taken randomly from the free LV wall on each sections. After background subtraction, eye controlled auto-threshold have been determined to detect positive areas. The fractional area (picrosirius red positive area-to-total area ratio) was determined on each image, and the mean value of the images represents each animal.

**Cardiac mRNA analysis.** LV myocardial tissue samples were harvested immediately after animals were euthanized, snap-frozen in liquid nitrogen, and stored at −80°C. LV tissue of eight animals from each group was homogenized in a lysis buffer (RLT buffer; Qiagen, Hilden, Germany); RNA was isolated from the ventricular samples using the RNeasy Fibrous Tissue Mini Kit (Qiagen) according to the manufacturer’s instructions and quantified by measuring optical density (OD) at 260 nm. RNA purity was ensured by obtaining a 260/280 nm OD ratio ~2.0. Reverse transcription reaction (1 μg total RNA of each sample) was completed using the QuantiTect Reverse Transcription Kit (Qiagen). Quantitative real-time PCR was performed with the StepOnePlus Real-Time PCR System (Applied Biosystems, Foster City, CA) in triplicates of each sample in a volume of 10 μl in each well containing cDNA (1 μl), TaqMan Universal PCR MasterMix (5 μl), and a TaqMan Gene Expression Assay (0.5 μl) (Applied Biosystems) for the following genes: myocardial hypertrophy markers, such as α-myosin heavy chain (α-MHC; assay ID: Rn00691721_g1), β-myosin heavy chain (β-MHC; assay ID: Rn00568328_m1), and atrial natriuretic factor (ANF; Rn00561661_m1); endogenous antioxidants, such as catalase (Cat; assay ID: Rn00560930_m1), SOD-2 (assay ID: Rn00690690_m1), and thioredoxin-1 (TRX-1; assay ID: Rn00587437_m1); inflammatory markers, such as IL-1β (assay ID: Rn00508043_m1), TNF-α (assay ID: Rn99999017_m1), and transforming growth factor β (assay ID: Rn00572010_m1); and markers related to mitochondrial function, such as peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC-1α; assay ID: Rn00580241_m1), nuclear respiratory factor 1 (NRF1; assay ID: Rn01455958_m1), estrogen-related receptor α (ERR-α; assay ID: Rn00433142_m1), peroxisome proliferator-activated receptor α (PPAR-α; assay ID: Rn00566193_m1), and cytochrome c (CytC; assay ID: Rn00470541_g1). Gene expression data were normalized to GAPDH (reference gene; assay ID: Rn01775763_g1), and expression levels were calculated using the CT comparative method (2−ΔΔCT). All results are expressed as values normalized to a positive calibrator (a pool of cDNA from all samples of the control group).

**Statistical analysis.** Statistical analysis was performed on a personal computer by using GraphPad Prism software (GraphPad Software, San Diego, CA). Results are expressed as means ± SE. We confirmed normal distribution of our data by the Shapiro-Wilk test. Thereafter, an unpaired two-sided Student’s t-test was used to compare parameters of control and corresponding hypertrophic rats (Co vs. Ex, Sham vs. AB).

To compare hypertrophy-induced alterations of cardiac function and myocardial gene expression between the two hypertrophy models, individual values of animals with cardiac hypertrophy were normalized to the mean value of the corresponding control group. These normalized data (given in percentage) were compared by using two-sided Student’s t-test after confirming normal distribution. Differences were considered statistically significant when P < 0.05.

The relations between LV mass values and sensitive indexes of LV contractility were examined by Pearson’s correlation.

**RESULTS**

**Body weight and heart weight data.** Body weight was decreased in exercised rats compared with control ones (402 ± 10 g Ex vs. 469 ± 10 g Co; P < 0.001), whereas it did not differ in the model of pathological hypertrophy (421 ± 14 g Sham vs. 435 ± 16 g AB; P = 0.517). Post-mortem measured heart weight values normalized to TL were increased in both physiological and pathological hypertrophy (Fig. 1A).

**Echocardiography.** Echocardiography data are shown in Table 1. There was no difference in HR between the hypertrophied and corresponding control groups. Anterior and posterior wall thickness values, either in diastole or systole, were increased in both models. LVEDD remained unaltered, whereas LVESD was decreased in physiological hypertrophy. Pathological hypertrophy was associated with unaltered ventricular dimensions, despite a tendency toward higher values that could be observed. Consequently, FS and EF, two conventional parameters of systolic function, were improved in exercised animals, whereas they remained unchanged in AB rats. The calculated LV mass and LV mass index (normalized to TL) values were markedly elevated in the hypertrophic groups, clearly underpinning LV hypertrophy (Table 1 and Fig. 1A).

According to LV mass index values, the degree of LV hypertrophy was similar in these two models (Table 1 and Fig. 1A).

**Hemodynamic measurements.** Figure 2 shows typical steady-state P-V loops obtained from Co, Ex, Sham, and AB animals. The widening of the baseline loops can be observed. Consequently, FS and EF, two conventional parameters of systolic function, were improved in exercised animals, whereas they remained unchanged in AB rats. The calculated LV mass and LV mass index (normalized to TL) values were markedly elevated in the hypertrophic groups, clearly underpinning LV hypertrophy (Table 1 and Fig. 1A).

As shown in Table 2, exercise training was associated with decreased LVESV along with unaltered HR, MAP, LV pressure values, dp/dtmax, and dp/dtmin. Consequently, SV, EF, CO, and CI were increased in Ex rats compared with Co rats, suggesting increased systolic performance. In the model of pressure overload-induced hypertrophy, increased MAP, LVESP, and dp/dtmax values were observed along with un-
changed HR, LVEDP, and dP/dt. LV end-diastolic volume, LVEDV, and SV tended toward higher values, but without reaching statistical significance; thus EF, CO, and CI did not differ between the Sham and AB groups. TPR and Ea were decreased in Ex rats compared with Co rats, whereas higher values were shown in AB rats compared with Sham ones (Table 2).

A load-independent index of LV relaxation, τ, was significantly improved in physiological hypertrophy, whereas it showed a marked deterioration in pathological hypertrophy, resulting in an important difference between the two kinds of hypertrophy (Table 2 and Fig. 4).

Figure 3 displays representative P-V loops obtained during inferior vena cava occlusions in Co, Sham, and AB rats. Overall results of ESPVR, PRSW, and dP/dtmax-EDV are depicted in Fig. 3. All of these sensitive contractility parameters were increased in both hypertrophy models, and no significant difference was calculated between the two kinds of hypertrophy (Figs. 3 and 4). Moreover, load-independent contractility parameters showed positive linear correlation with calculated LV mass values involving the data of all animals (LV mass vs. ESPVR: r = 0.535, P < 0.001, Fig. 3; LV mass vs. PRSW: r = 0.490, P = 0.003; LV mass vs. dP/dtmax-EDV: r = 0.434, P = 0.012).

Table 1. Echocardiographic data

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<th>Physiological Hypertrophy</th>
<th>Pathological Hypertrophy</th>
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<tr>
<td></td>
<td>Co</td>
<td>Ex</td>
</tr>
<tr>
<td>HR, beats/min</td>
<td>357 ± 15</td>
<td>351 ± 10</td>
</tr>
<tr>
<td>LVAWTd, mm</td>
<td>1.96 ± 0.02</td>
<td>2.17 ± 0.02</td>
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<td>LVAWTs, mm</td>
<td>3.02 ± 0.05</td>
<td>3.37 ± 0.06</td>
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<tr>
<td>LVPWTd, mm</td>
<td>1.82 ± 0.03</td>
<td>1.93 ± 0.02</td>
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<tr>
<td>LVPWTs, mm</td>
<td>2.91 ± 0.04</td>
<td>3.09 ± 0.04</td>
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<tr>
<td>LVEDD, mm</td>
<td>6.85 ± 0.05</td>
<td>6.89 ± 0.03</td>
</tr>
<tr>
<td>LVESD, mm</td>
<td>4.06 ± 0.08</td>
<td>3.45 ± 0.09</td>
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<tr>
<td>FS, %</td>
<td>41.4 ± 0.7</td>
<td>50.0 ± 1.1</td>
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<tr>
<td>EF, %</td>
<td>65.5 ± 1.3</td>
<td>73.0 ± 1.3</td>
</tr>
<tr>
<td>LV mass, g</td>
<td>0.87 ± 0.01</td>
<td>0.98 ± 0.01</td>
</tr>
<tr>
<td>LV mass index, g/cm</td>
<td>0.202 ± 0.004</td>
<td>0.246 ± 0.003</td>
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Values are means ± SE. Co, control; Ex, exercised; AB, aortic banding; HR, heart rate; LVAWTd and LVAWTs, left ventricular (LV) anterior wall thickness at diastole and systole, respectively; LVPWTd and LVPWTs, LV posterior wall thickness at diastole and systole, respectively; LVEDD, LV end-diastolic dimension; LVESD, LV end-systolic dimension; FS, LV fractional shortening; EF, LV ejection fraction. LV mass index is LV mass normalized to tibial length.
Picrosirius staining revealed no collagen deposition in the myocardium of exercise-trained rats. In contrast, increased subendocardial fractional area was observed in pathological hypertrophy, whereas the subepicardial collagen content did not differ between the Sham and AB rats (Fig. 1B).

**Cardiac mRNA analysis.** Myocardial gene expressions of ANF and β-MHC were markedly increased, whereas expression of α-MHC was markedly decreased (resulting in decreased α/β-MHC ratio) in the model of pathological hypertrophy; the expression values of these pathological hypertrophy markers did not differ between the control and exercised groups (Fig. 6A). There was no difference neither within nor between the models regarding the myocardial expression of major antioxidants (catalase, SOD-2, thioredoxin-1) and inflammation-related markers (TGF-β, TNF-α, and IL-1β) (Fig. 6, B and C). Expression of markers related to mitochondrial function (PGC-1α, NRF1, ERR-α, PPAR-α, and CytC) were

### Table 2. Hemodynamic data

<table>
<thead>
<tr>
<th></th>
<th>Physiological Hypertrophy</th>
<th>Pathological Hypertrophy</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>Co</td>
<td>Ex</td>
</tr>
<tr>
<td>HR, beats/min</td>
<td>412 ± 9</td>
<td>400 ± 8</td>
</tr>
<tr>
<td>MAP, mmHg</td>
<td>145.3 ± 2.8</td>
<td>142.1 ± 5.2</td>
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<tr>
<td>LVESP, mmHg</td>
<td>154.8 ± 3.4</td>
<td>152.2 ± 8.2</td>
</tr>
<tr>
<td>LVEDP, mmHg</td>
<td>3.0 ± 0.2</td>
<td>3.7 ± 0.4</td>
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<tr>
<td>dP/dtmax, mmHg/s</td>
<td>9,237 ± 397</td>
<td>9,847 ± 659</td>
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<tr>
<td>dP/dtmin, mmHg/s</td>
<td>−12,191 ± 443</td>
<td>−12,213 ± 670</td>
</tr>
<tr>
<td>τ, ms</td>
<td>12.1 ± 0.3</td>
<td>11.2 ± 0.3</td>
</tr>
<tr>
<td>LVESV, μl</td>
<td>234.4 ± 5.2</td>
<td>239.2 ± 6.1</td>
</tr>
<tr>
<td>SV, μl</td>
<td>111.5 ± 2.0</td>
<td>98.6 ± 2.6</td>
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<tr>
<td>EF, %</td>
<td>123.1 ± 5.5</td>
<td>140.6 ± 4.8</td>
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<tr>
<td>CO, ml/min</td>
<td>52.2 ± 1.4</td>
<td>58.3 ± 0.8</td>
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<tr>
<td>CI, (ml/min)/100 g BW</td>
<td>50.6 ± 2.0</td>
<td>56.2 ± 1.5</td>
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<tr>
<td>SW, mmHg·ml</td>
<td>10.9 ± 0.5</td>
<td>14.2 ± 0.4</td>
</tr>
<tr>
<td>Slope of EDPRV, mmHg/μl</td>
<td>14.7 ± 0.8</td>
<td>18.0 ± 0.8</td>
</tr>
<tr>
<td>Slope of dP/dV, mmHg/μl</td>
<td>0.036 ± 0.003</td>
<td>0.042 ± 0.003</td>
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<tr>
<td>TPR, mmHg·(ml/min)</td>
<td>2.91 ± 0.09</td>
<td>2.54 ± 0.12</td>
</tr>
<tr>
<td>PVR, mmHg·(ml/min)</td>
<td>1.33 ± 0.05</td>
<td>1.09 ± 0.08</td>
</tr>
<tr>
<td>Ees, mmHg/μl</td>
<td>0.53 ± 0.03</td>
<td>0.32 ± 0.03</td>
</tr>
<tr>
<td>VAC</td>
<td>0.08 ± 0.02</td>
<td>0.48 ± 0.07</td>
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</table>

Values are means ± SE. Hemodynamic parameters were measured by the Millar pressure-volume conductance catheter system. MAP, mean arterial pressure; LVESP, LV end-systolic pressure; LVEDP, LV end-diastolic pressure; dP/dtmax and dP/dtmin, maximal slope of the systolic pressure increment and the diastolic pressure decrement, respectively; τ, time constant of LV pressure decay; LVEDV, LV end-diastolic volume; LVESV, LV end-systolic volume; SV, stroke volume; CO, cardiac output; CI, cardiac index; BW, body weight; SW, stroke work; EDPRV, end-diastolic pressure-volume relationship; TPR, total peripheral resistance; Ees, arterial elastance; VAC, ventriculoarterial coupling.

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Hemodynamics of Physiological versus Pathological LV Hypertrophy

We also determined EDPVR, a sensitive marker of LV stiffness, which tended toward higher values in both hypertrophy (Table 2). This slight increase did not differ between the two models (Fig. 4).

Regarding mechanoenergetics, our results showed increased SW in both models. Values of PVA were elevated in only two models (Fig. 4).

Histology. Increased cardiomyocyte width values were observed in exercised and aortic banding groups compared with controls and was unchanged in pathological hypertrophy (Table 2). This slight increase did not differ between the models regarding the myocardial expression of major antioxidants (catalase, SOD-2, thioredoxin-1) and inflammation-related markers (TGF-β, TNF-α, and IL-1β) (Fig. 6, B and C). Expression of markers related to mitochondrial function (PGC-1α, NRF1, ERR-α, PPAR-α, and CytC) were...
not altered in physiological hypertrophy. In contrast, pathological hypertrophy was associated with myocardial downregulation of these genes, which led to characteristic differences between the two hypertrophy models (Fig. 6D).

DISCUSSION

According to our knowledge, this is the first study to provide a direct hemodynamic comparison of physiological and pathological LV hypertrophy using pressure-conductance catheter system in relevant small animal models. Despite the similar degree of LV hypertrophy, distinct characteristics of energy-dependent LV performance were observed. According to our data, different myocardial expression of mitochondrial markers might indicate the alteration of mitochondrial regulation as a possible underlying mechanism.

Characterization of LV hypertrophy. Cardiac hypertrophy is described as the response of the heart to a variety of stimuli that impose increased biomechanical stress. An increase in cardiomyocyte size, enhanced protein synthesis, and a higher organization of the sarcomere lead to morphological alterations resulting in hypertrophic phenotype (12). According to our results, in response to chronic pressure overload or exercise training, the heart weight values of rats increased significantly. Both aortic banding and intense swimming training resulted in a significant increase of LV wall thickness values (Table 1). LV mass index data revealed the degree of LV hypertrophy ∼20–25% after exercise training and ∼25–30% after chronic pressure overload stimulus (Fig. 1A), which are comparable with the results of our previous study (29) and other experimental investigations (24, 42, 45). Histological evaluation of LV cardiomyocyte width on HE-stained sections underpinned the LV hypertrophy in both models (Fig. 1). All of these data suggest marked LV hypertrophy in our animal models with similar degree of myocardial enlargement. Relatively unchanged LV cavity dimensions as well as FS and EF revealed no pathological hypertrophy-associated dilatation in AB rats, which is in line with previous investigations using abdominal aortic banding (Table 1) (8, 17).

The increase in collagen was localized only in subendocardial area after 6-wk banding period (Fig. 1B). This is in line with a previous investigation using abdominal aortic banding model, where subendocardial collagen deposition was ob-
served after 2 mo of chronic pressure overload (8). The absence of fibrotic remodeling supports the physiologic nature of LV hypertrophy after long-term exercise training (45). In contrast with exercise training-induced cardiac hypertrophy, in the case of pathological hypertrophy the observed LV enlargement is accompanied by the reinduction of myocardial fetal gene program (12). Myocardial gene expression values of widely used pathological hypertrophy markers α-MHC, β-MHC, and ANF clearly demonstrated the physiological and pathological nature of LV hypertrophy in Ex and AB rats, respectively (Fig. 6A) (16, 24). Pathological hypertrophy advanced to heart failure is associated with myocardial oxidative stress and activated inflammatory mechanisms (1). Unaltered myocardial gene expressions of endogenous antioxidants and inflammatory cytokines after 6 wk of aortic banding might indicate that intensive oxidative stress and consecutive inflammatory response are not present in this compensated phase (Fig. 6, B and C). This is in line with results of an experiment comparing compensated myocardial hypertrophy with failing heart in a pressure overload model (2). No alterations in expression of oxidative stress- and inflammatory-related markers were expected in exercise-induced hypertrophy according to literature data (16).

Baseline hemodynamics. As it is clearly depicted on Fig. 2, unaltered end-diastolic dimensions along with decreased end-systolic dimensions—accompanied by similar LV pressure relations and heart rate—were observed in exercised animals under anesthesia (Fig. 2 and Table 2), which is contradictory to observations in humans, where athlete’s heart induced by aerobic training was associated with decreased heart rate and increased end-diastolic volume (22). As a substantial point, in line with human studies using noninvasive methods, baseline pressure-volume relations revealed increased stroke volume in athlete’s heart (34, 36).

As shown in Table 2, abdominal aortic banding was associated with a marked increase in MAP (≈40 mmHg) and in peak LV systolic pressure (≈60 mmHg), which degree is comparable with other studies using aortic constriction to induce pathological hypertrophy in rodents (19, 24), whereas LV end-diastolic pressure remained unchanged. The baseline pressure-volume loops showed a slight shift rightward since it is described in developing cardiac hypertrophy induced by pressure overload (26). However, no marked dilation was observed since LV volume values showed only a tendency toward higher values without reaching the level of significance.
Fig. 6. Myocardial gene expression analysis. Relative myocardial expression of genes related to pathological hypertrophy [A: α-myosin heavy chain (α-MHC), β-myosin heavy chain (β-MHC), and atrial natriuretic factor (ANF)], oxidative stress [B: catalase (Cat), superoxide dismutase-2 (SOD-2), and thioredoxin-1 (TRX-1)], inflammation [C: transforming growth factor β (TGF-β), TNF-α, and IL-1β], and mitochondrial function [D: peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC-1α), nuclear respiratory factor 1 (NRF1), estrogen-related receptor α (ERR-α), peroxisome proliferator-activated receptor α (PPAR-α), and cytochrome c (CytC)] in Co, Ex, Sham, and AB rats is shown. P values in the middle of the graphs indicate differences between the 2 models. Values are means ± SE. #P < 0.05 vs. Sham. n.s., not significant.
HEMODYNAMICS OF PHYSIOLOGICAL VERSUS PATHOLOGICAL LV HYPERTROPHY

In summary, when physiological and pathological stimuli-induced alterations of baseline pressure-volume loops are compared, the main characteristics are the widening of the P-V loop reflecting increased stroke volume in case of physiological hypertrophy, whereas increased systolic pressure values along with relatively unaltered stroke volume were shown in pathological hypertrophy by P-V analysis.

Systolic function and LV contractility. Conventional parameters of systolic function including LV FS and EF by echocardiography and EF by P-V analysis were elevated in the exercised rats, suggesting increased LV systolic performance in physiological hypertrophy (Table 2). These parameters remained unchanged, which demonstrated preserved systolic function after 6 wk of pressure overload, indicating clearly compensated pathological cardiac hypertrophy in accordance with a previous study using relatively short-term (2 mo) abdominal aortic banding (8). Another traditional index of LV systolic function is the peak rate of systolic pressure increment (dP/dtmax). However, it is influenced especially by preload but also by extreme grade of afterload and heart rate (5). Thus the potential alteration of cardiac preload in trained animals and the extremely increased LV afterload in AB rats turn this parameter unreliable.

LV contractility can be assessed precisely and reliably by pressure-volume analysis. By the help of pressure-volume relations being recorded during a transient preload reduction maneuver (vena cava inferior occlusion), the load-independent relations being recorded during a transient preload reduction (dP/dtmin) was unchanged in both types of LV hypertrophy; however, the value of this parameter is very limited because of dependence on many hemodynamic factors, such as cardiac loading conditions (13). In contrast, the time constant of LV pressure decay has been described as a relatively load-independent index of LV active relaxation (46). Physiological hypertrophy was associated with enhanced relaxation as shown by decreased τ, thus a shorter isovolumetric relaxation period (Table 2). This is in line with our previously published data in a rat model of athlete’s heart (31). In contrast, we observed a markedly delayed relaxation in pathological hypertrophy (Table 2). In accordance, investigations for functional characterization of LV hypertrophy induced by pressure overload have yielded similar results regarding active relaxation. Animal models that included aortic constriction (19, 40) or spontaneously hypertensive rats (5) have described a marked impairment in active relaxation. This distinction might be the most characteristic difference between the two types of cardiac hypertrophy (Fig. 4). This is in accordance with recent human echocardiographic studies, where hypertrophic cardiomyopathy and athlete’s heart could be distinguished by early diastolic components (3, 20).

Ventricular end-diastolic stiffness, although it can be influenced by different factors, is affected predominantly by alterations in myocardial intracellular and extracellular structural components (e.g., fibrosis, edema) (30). This passive viscoelastic property of the myocardium can be quantified by examination of the relationship between diastolic pressure and volume (47). EDPVR showed a slight tendency in both types of hypertrophy toward higher values—without reaching statistical significance—, and no difference could be observed between the hypertrophy models (Table 2 and Fig. 4). These data are in good agreement with the lack of collagen deposition observed in the hearts of exercised animals. The observed slight increase of subendocardial (but not subepicardial) fibrosis in AB rats might be not so relevant as resulting in characteristic functional consequences (Fig. 1B). Unchanged LVEDP was also consistent with these observations (Table 2).

Mechanoenergetics. SW, the effective external mechanical work of LV in one cardiac cycle, can be calculated as the area enclosed by the PV loop. Increased mechanical work was observed both in exercise training- and pressure overload-induced LV hypertrophy due to increased stroke volume and elevated systolic pressure values, respectively (Fig. 5 and Table 2). PVA, the specific area in the P-V plane bounded by the end-systolic and end-diastolic P-V relationship lines and the systolic segment of the P-V loop, serves as a reliable index of total mechanical energy and is directly proportional to myocardial oxygen consumption (38). Although physiological hypertrophy was related to similar energy consumption as compared with control animals, pathological hypertrophy was associated with an increased energy consumption of LV myocardium (Fig. 5). As a consequence, mechanical efficiency of ventricular performance was increased in exercise training-induced hypertrophy, whereas it remained unaltered in pathological hypertrophy (Fig. 5). These results might reflect the previously described distinction of metabolic efficiency between physiological and pathological myocardial conditions: exercise training is associated with enhanced fatty acid and glucose oxidation, whereas pathological hypertrophy is related to a decrease in fatty acid oxidation and increase in glucose metabolism (1).
As an important factor of cardiac energetics, ventricular-arterial coupling expresses the interaction between left ventricle and arterial system by the ratio of arterial elastance and LV end-systolic elastance (39). Arterial system can be characterized by effective arterial elastance ($E_a$), an integrative index that includes, among others, peripheral vascular resistance, arterial compliance, and characteristic impedance. Decreased $E_a$ in exercised rats reveals a better compliance of the arterial system in physiological hypertrophy, whereas increased $E_a$ in pathological hypertrophy clearly reflects the increased afterload due to pressure overload (Table 2). These alterations were underpinned by changes in TPR (Table 2). ESPVR, the substantial index of contractility, corresponds to LV end-systolic elastance (32). Improved VAC in exercise-trained animals reflects a more appropriate matching between the LV and the arterial system, which results in an optimal transfer of blood from the LV to the periphery without excessive changes in pressure. Unaltered VAC in pressure overload-induced pathological hypertrophy indicates that increased systolic LV performance compensates the increased arterial load (Table 2). These observations are in accordance with human investigations describing ventriculo-arterial relations in athletes and in patients with hypertension-induced LV hypertrophy (11, 27).

Mitochondrial function. The distinct cardiac functional parameters as active relaxation, SW, and mechanical efficiency have been each referred to as energy-dependent indexes. Thus we examined the myocardial expression of a mitochondrial transcriptional coactivator PGC-1α, the master regulator of myocardial energy metabolism, by regulating mitochondrial biogenesis and function (10). In line with previous literature data (32), both PGC-1α and its downstream coactivators and targets (ERR-β, NRF1, PPAR-α, and Cytc) were downregulated in pathological hypertrophy, suggesting mitochondrial dysfunction and a shift from lipid to glucose utilization (Fig. 6D). Physiological cardiac hypertrophy is associated with normal or enhanced mitochondrial biogenesis (32). According to our data, there was no significant upregulation of these genes in our swim training model of athlete’s heart (Fig. 6D). This difference between our cardiac hypertrophy models supports the concept that targeting regulator molecules of mitochondrial biogenesis and function might play an important role in the treatment of pathological hypertrophy.

Limitations and perspectives. This study was designed to compare functional and molecular alterations in the most widely used pressure overload-induced compensated pathological LV hypertrophy and dynamic exercise-induced LV hypertrophy models. To compare these conditions to decompensated hypertrophy or volume overload-induced pathological hypertrophy, further investigations are needed.

Changes in coronary angiogenesis and capillary density might be an important contributing factor to myocardial hypertrophic situations because the enlarged cardiac mass requires increased blood supply to maintain energy-dependent cellular processes; thus examining myocardial capillary density might be an important issue in future investigations comparing physiological and pathological hypertrophic phenotypes (28).

We should also mention that for a more accurate analysis of cardiomyocyte size, hearts should be arrested in diastole using intravenous injection of potassium chloride solution, which was not possible in the present study, due to methodological reasons.

Conclusions. In conclusion, we provided the first detailed hemodynamic comparison of physiological and compensated pathological hypertrophy in relevant rodent models. We confirmed LV hypertrophy of comparable degree in swim-trained and AB animals with characteristic attributes of physiological and pathological stimuli-induced histological and gene expression alterations. Increased LV contractility could be observed in both types of myocardial conditions, and contractility correlated with the degree of hypertrophy. Although the degree of hypertrophy was similar in the two models, characteristic differences were detected in early diastolic function: active relaxation was improved in physiological hypertrophy, whereas a marked deterioration was observed in pathological hypertrophy. A tendency toward increased myocardial stiffness was observed in both types of myocardial hypertrophy. LV mechan energetics (mechanical efficiency and ventriculo-arterial coupling) were more optimal in exercise-training-induced hypertrophy, whereas they remained unaltered in pressure overload-induced pathological hypertrophy. Altered myocardial expression of markers related to mitochondrial function and biogenesis might explain the described energy-dependent functional differences.

ACKNOWLEDGMENTS

We thank Henriett Biró, Tímea Fischinger, Viktória Gregor, Dóra Juhász, Gábor Fritz, and Gábor Alt for expert technical assistance and Dr. Gábor Kőkény for scientific advice.

GRANTS

This work was supported by a grant from the National Development Agency of Hungary (TÁMOP-4.2.2/B-10/1-2010-0013), by the Hungarian Scientific Research Fund (OTKA 105555 to B. Merkely), and by the János Bolyai Research Scholarship of the Hungarian Academy of Sciences (to T. Radovits).

DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the author(s).

AUTHOR CONTRIBUTIONS


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