Heart rate-induced modifications of concentric left ventricular hypertrophy: exploration of a novel therapeutic concept

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Klein FJ, Bell S, Runte KE, Lobel R, Ashikaga T, Lerman LO, LeWinter MM, Meyer M. Heart rate-induced modifications of concentric left ventricular hypertrophy: exploration of a novel therapeutic concept. *Am J Physiol Heart Circ Physiol* 311: H1031–H1039, 2016. First published September 2, 2016; doi:10.1152/ajpheart.00301.2016.—Lowering the heart rate is considered to be beneficial in heart failure (HF) with reduced ejection fraction (HFrEF). In a dilated left ventricle (LV), pharmacological heart rate lowering is associated with a reduction in LV chamber size. In patients with HFrEF, this structural change is associated with better survival. HF with preserved ejection fraction (HFpEF) is increasingly prevalent but, so far, without any evidence-based treatment. HFpEF is typically associated with LV concentric remodeling and hypertrophy. The effects of heart rate on this structural phenotype are not known. Analogous with the benefits of a low heart rate on a dilated heart, we hypothesized that increased heart rates could lead to potentially beneficial remodeling of a concentrically hypertrophied LV. This was explored in an established porcine model of concentric LV hypertrophy and fibrosis. Our results suggest that a moderate increase in heart rate can be used to reduce wall thickness, normalize LV chamber volumes, decrease myocardial fibrosis, and improve LV compliance. Our results also indicate that the effects of heart rate can be titrated, are reversible, and do not induce HF. These findings may provide the rationale for a novel therapeutic approach for HFpEF and its antecedent disease substrate.

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Heart rate; hypertrophy; pacemaker; remodeling

NEW & NOTEWORTHY

This study is the first to explore if moderate heart rate elevations could be therapeutically employed to induce beneficial structural changes in the concentrically hypertrophied left ventricle. The result that heart rate elevations can induce structural changes that improve diastolic filling challenges the canonical thought that they are harmful.

HEART FAILURE (HF) is a leading cause of hospitalizations and a major public health problem (7). About half of patients with HF have a preserved ejection fraction (HFpEF), and the prevalence of HFpEF is growing relative to HF with reduced ejection fraction (HFrEF) (19, 24). No treatment has been shown to improve survival in patients with HFpEF, and few interventions improve symptoms or quality of life. Thus, identification of novel treatment targets for the underlying myocardial substrate is an important goal.

At a structural level, HFpEF is typically associated with concentric remodeling with an increased left ventricular (LV) mass-to-volume ratio or overt LV hypertrophy and fibrosis (29, 30). A high prevalence of this structural phenotype in HFpEF was recently confirmed in the Treatment of Preserved Cardiac Function Heart Failure with an Aldosterone Antagonist (TOPCAT) trial (21). Despite the inclusion of patients with LV chamber dilatation in this trial, about a quarter of patients had below-normal chamber volumes. A 20–50% reduction in LV chamber size has also been documented with normal aging in population studies (1, 12). Small chamber volumes are associated with a poor exercise capacity and contribute to an increase in the LV mass-to-volume ratio, which is a strong independent predictor of the future development of HF (12, 18).

In line with the established beneficial structural effects of lower heart rates in dilated cardiomyopathy, we hypothesized that a moderate increase in heart rate could be used to favorably alter the basic structure of the concentrically remodeled LV. This would be accomplished by a reduction in LV wall thickness, normalization of LV chamber volumes, and reorganization of the interstitial matrix that, together, may improve diastolic LV filling. In a first attempt to explore the potential and safety of this approach, we studied the effects of continuous atrial pacing in a well-characterized porcine model of concentric LV hypertrophy and fibrosis induced by unilateral renal artery stenosis (RAS) (10, 27). Because no previous work provides guidance on the heart rates needed to achieve these goals, we used an exploratory study design that would allow us to first find and then confirm a target heart rate.

MATERIALS AND METHODS

Animal Studies

All aspects of animal care were in accordance with National Institutes of Health guidelines and the standards of the Animal Care and Use Committee of the University of Vermont, which approved the protocol used in this study. Because the effects of moderately elevated heart rates on the structural (volume) and functional (ejection fraction [EF]) effects are unknown, a “heart rate finding study” allowed us to determine a target heart rate that would be associated with a structural change without affecting the EF. This was followed by a “heart rate confirmatory study” and a “high heart rate study.” A schematic outline of the experimental protocols is provided in Fig. 1.

A total of 10 adult female Yucatán miniature swine (9 mo old, 40–45 kg body wt; Sinclair Bio Resources, Windham, ME) were used for these studies. A pacemaker was implanted in all pigs. At the same time, a unilateral RAS was established in eight pigs to induce LV hypertrophy. Two pigs underwent a sham procedure without RAS. At
each study time point, meloxicam (0.2 mg/kg po) was administered as a preoperative analgesic followed by ketamine (20 mg/kg im) and atropine (0.024 mg/kg im). Anesthesia was induced with 5% isoflurane to allow endotracheal intubation and maintained at 1–3%. Thermal support was provided by a heated water blanket. Oxygen saturation, expired carbon dioxide levels, limb lead ECG, and rectal temperature were recorded throughout the surgical and diagnostic procedures. At each time point, a blood sample was collected and centrifuged (10 min at 3,000 rpm). The resulting supernatant was transferred to a 80°C freezer for later analyses.

During the entire course of the study, the animals were assessed on a daily basis for signs of discomfort, the incision site was evaluated, and respiratory abnormalities were assessed. The lungs were auscultated once a week, and the pacemaker was interrogated intermittently to ensure proper function. Upon study termination, the heart and both kidneys were removed. The atria were removed, and the right ventricle and LV were weighed. Tibia length and body weight were measured and used to index LV mass. Transmural samples were excised from the LV anterior wall. One sample was frozen in liquid nitrogen and stored in an 80°C freezer, and another sample was preserved in 10% formaldehyde solution.

Pacing Protocols

At 3 wk after the initial procedure, the animals were sedated and intubated to document the cardiac effect of RAS. In the first exper-
mental group (heart rate dose finding), the pacemaker was programmed to pace the atrium at 150/min for the following 2 wk (weeks 3–5 after the initial surgeries). Ventricular pacing was limited by selection of a rate-adaptive atrioventricular algorithm that prioritizes physiological atrioventricular conduction. All rates are well below those reported to result in tachycardia-induced dilated cardiomyopathy in large-animal models (10).

Echocardiography

Transthoracic and transesophageal echocardiography (Sequoia C256, Siemens Acuson) was performed to quantify LV volume and assess diastolic function. All measurements were obtained in sinus rhythm without pacing. Optimized two-dimensional transthoracic echocardiographic views that maximize endocardial definition were obtained from the parasternal long- and short-axis views, and two transesophageal LV chamber views were obtained at 0° and 90°. Echocardiographic images were recorded over at least two cardiac cycles. LV end-diastolic and end-systolic volumes were obtained using the method of disks (modified Simpson’s analysis) according to clinical guideline recommendations (13). LV end-diastolic and end-systolic volumes and EF were calculated by averaging the data from the three echocardiographic windows. LV mass was calculated from the parasternal short-axis dimensions (3). Doppler-based measurements of peak early mitral inflow (E) and the early displacement of the mitral annulus tissue (E’) were obtained to calculate E/E’ as a marker of diastolic function.

End-Diastolic Pressure and Volume Measurements

We also assessed the effect of pacing-induced structural changes after pacing at 125/min on LV end-diastolic pressures (EDP) at prespecified time points in six animals (heart rate dose confirmation and high heart rate studies). The right carotid artery was accessed via a surgical cutdown. A catheter was advanced into the LV cavity to allow for pressure measurements accompanied by simultaneous echocardiographic volume assessments. After the first pressure-volume measurement, three sequential 250-ml infusions of normal saline were administered over 5–10 min. Pressure-volume measurements were made after each infusion. LV compliance was calculated as the ratio of LV EDP change to LV end-diastolic volume (EDV) change made after each infusion. LV compliance was calculated as the ratio of LV EDP change to LV end-diastolic volume (EDV) change (ΔEDV/ΔEDP) from baseline to the completion of the last saline infusion.

Blood Samples

Blood samples were collected from each pig at baseline and at each time point. The following biomarkers were assayed: B-type natriuretic peptide (BNP) and norepinephrine (NE) for neurohumoral activation and galectin-3, a cardiac fibrosis marker (2, 9). BNP and NE have been reported to be elevated in HF models of tachycardia-induced cardiomyopathy (10, 22, 23). Porcine-specific BNP, NE, and porcine-specific galectin-3 levels were determined using commercial competitive ELISAs: Phoenix Pharmaceuticals (Burlingame, CA) for BNP, LDN Labor Diagnostika Nord (Nordhorn, Germany) for NE, and BlueGene Biotech (Shanghai, China) for galectin-3. In addition to the samples provided to establish the standard curve, all kits provided for negative and positive controls.

PicroSirius Red Stain

The LV anterior wall samples preserved in formaldehyde were embedded in paraffin. The paraffin blocks were cut in 5-μm sections with a standard low-profile steel microtome blade on a Leica paraffin microtome (Buffalo Grove, IL). Sections on glass slides were air-dried overnight and then baked at 70°C for 30 min. PicroSirius red staining was performed as previously described (11). Tissue sections were deparaffinized, rehydrated, and stained with filtered 0.1% Sirius red F3BA (Pfaltz & Bauer, Waterbury, CT) in saturated aqueous picric acid, pH 2, for 90 min at room temperature, as previously described by Dolber and Spach (4). An upright light microscope (model BX50, Olympus) was used to electronically overlay a grid on the slide, and each box was assigned a number. Ten grid squares were selected for imaging via a random number generator. Images were acquired at ×10 magnification with an UPLANFL ×10 (numerical aperture 0.3) objective lens. Polarized light images were captured as eight-bit gray-scale images with a digital charge-coupled device camera (Retiga 2000R, QImaging, Burnaby, BC, Canada). The images were analyzed using MetaMorph Imaging Series 7.1 image analysis software (Molecular Devices, Downingtown, PA). Each image was opened and calibrated to ×10 magnification. A threshold overlay was applied to the image to detect PicroSirius red staining. Values for threshold area and threshold area percentage were recorded and automatically logged to a spreadsheet. Ten random areas were analyzed, and the individual percent areas of fibrosis were averaged for each sample. This randomized systematic uniform analysis avoids selection bias, as it does not focus on perivascular connective tissue accumulations.

For comparison, myocardial sections from five additional non-paced female pigs with unilateral RAS were analyzed to provide porcine reference values (unstained slides were provided by L. O. Lerman). The animals had similar weights and were matched for RAS duration and level of LV hypertrophy. The same automated and blinded procedures were used to process and analyze the tissue slides.

Protein Slot Blot

LV myocardial samples from all animals were homogenized using a glass tissue grinder followed by ultrasonication. A total of 3 μg of the homogenate from each animal were bound to a nitrocellulose membrane using a slot-blot apparatus (Bio-Dot SF, Bio-Rad, Hercules, CA). After it was blocked, the membrane was incubated with a
collagen I/III-specific polyclonal rabbit antibody raised against pig collagen (Bio-Rad) followed by a secondary horseradish peroxidase anti-rabbit antibody and detection by chemiluminescence. Signal linearity was confirmed in a dilution series using two additional samples. The densitometric analysis was performed using ImageJ 1.50b software (National Institutes of Health, Bethesda, MD).

**Data Presentation and Statistical Methods**

Individual data items were examined using a linear mixed model for within-animal repeated-measure observations with a decomposition of the overall repeated-measure effect into single degree-of-freedom polynomial contrasts followed by pair-wise exploratory contrasts using paired t-tests. Sham and RAS group comparisons were conducted using two-sample tests and confirmed using the nonparametric Kruskal-Wallis rank-sum test when small sample sizes were involved. All rate-mediated postintervention and recovery effects were compared with the baseline measurements made 3 wk after surgery. Values are means ± SD if not otherwise indicated. No adjustments for multiple testing were made in this exploratory study; P ≤ 0.05 was used for significance testing. The data analysis was conducted using SYSTAT version 11 (SYSTAT Software, Richmond, CA). Box-and-whisker plots are used to present the biomarker results. A linear regression analysis was performed to evaluate the interaction between change in chamber volume and degree of hypertrophy.

**RESULTS**

**RAS-Induced LV Concentric Hypertrophy**

In the eight animals with high-grade unilateral RAS, the right kidney size was markedly smaller (Fig. 2B). In RAS animals, the LV septal and posterior wall thickness increased from 7.7 ± 0.1 to 10.6 ± 0.1 mm (P < 0.01) and from 7.1 ± 0.1 to 9.4 ± 0.1 mm (P < 0.01), respectively, at 3 wk after surgery prior to initiation of pacing. The average LV EDV was 28 ± 6 ml at baseline and 21 ± 2 ml after 3 wk (P = 0.18). The LV mass-to-volume ratio was substantially higher in RAS than sham-operated animals (4.0 ± 0.5 vs. 2.9 ± 0.3 g/ml, P < 0.05).

**Heart Rate Effects on LV Structure and Function**

Heart rate finding study. The average intrinsic heart rate prior to the pacing intervention was 93 ± 6/min. After 2 wk of predominant atrial pacing at 150/min, LV volumes markedly increased (Fig. 3). LV end-diastolic and systolic volumes assessed in sinus rhythm increased by 54 ± 3% and 106 ± 23%, respectively (both P < 0.01), whereas the EF fell from 58 ± 1% to 45 ± 1% (P < 0.05). LV mass was unchanged due to wall thinning. Because of the substantial increase in LV volume and the decrease in EF after 2 wk, the atrial pacing rate was subsequently decreased to 125/min. After 2 wk, the LV cavity size was smaller but still enlarged compared with baseline. LV end-diastolic and systolic volumes were increased by 29 ± 24% (P = 0.09) and 50 ± 19% (P < 0.01), respectively. Importantly, the EF normalized to 52 ± 1%. Thereafter, the pacing was stopped. After 2 wk, the LV chamber size had returned to baseline levels prior to the pacing (Fig. 3).

Heart rate confirmation study. In another four animals, we confirmed the effects of 125/min pacing in the absence of the initial period of pacing at 150/min (Fig. 4). The heart rate in the first 3 wk prior to pacing was 88 ± 4/min. After 2 wk of pacing at 125/min, the LV EDV in sinus rhythm increased by 25 ± Fig. 3. Pacing-induced changes in LV dimensions and biomarkers in the heart rate finding study. LV dimensions and plasma biomarkers were assessed 3 wk after the initial surgery. Thereafter, right atrial pacing was started at a rate of 150/min. After 2 wk, pacing was stopped to obtain all measurements before atrial pacing was restarted at a rate of 125/min. After 2 wk, pacing was stopped to assess LV dimensions, and a final assessment was carried out after 2 wk of sinus rhythm. Error bars represent SE. Biomarker [B-type natriuretic peptide (BNP) and norepinephrine (NE)] results are presented as box-and-whisker plots. EDV, end-diastolic volume; ESV, end-systolic volume; EF, ejection fraction.
14% (P < 0.05), with a trend toward an increased EDV (P = 0.09). The EF was 55 ± 1%, not different from baseline. The LV mass remained unchanged as well. After cessation of pacing, LV volumes returned to baseline values within 2 wk.

**High heart rate study.** In two additional animals, we set out to induce pathological LV dilation at a rate of 170/min. The goal was to explore if a pronounced dilation would regress once the rate was lowered to 125/min. After 4 wk at 170/min, the animals did not develop any signs of HF, despite a 141% increase in the average LV end-diastolic and a 246% increase in average end-systolic chamber volume. EF was reduced from 51% to 37% and from 59% to 33%. Within 2 wk of lowering the rate from 170/min to 125/min, the cavity size regressed to previously established 125/min levels and the EF was restored to 54% and 59%, respectively.

Table 1 summarizes the cumulative 125/min data under the following conditions: prior to pacing (baseline), immediately after 2 wk of pacing at 125/min, and 2 wk after pacing cessation (recovery). This recapitulates the significant increase in LV chamber dimensions and thinning of the LV wall, which in effect reduced the average mass-to-volume ratio by ~27% to normal levels. The two animals without RAS were included, reproducing the natural distribution of hypertrophy in HFpE.

**Heart Rate Effects on Biomarkers**

None of the animals developed HF, and the biomarkers did not change (Figs. 3 and 4). Galectin-3 levels were also unchanged at 150/min compared with baseline (27.2 ± 4.9 vs. 27.6 ± 6.0 ng/ml). In addition, BNP and NE levels remained in the normal range throughout the high heart rate study at all time points (BNP <8 ng/ml and NE <0.5 ng/ml). The cumulative biomarker results at 125/min are presented in Table 1.

**EDP-EDV Relationships**

In six animals, we were able to serially assess the LV EDP-EDV relationship before and after the heart rate-induced structural change (Fig. 6). After 2 wk of pacing at 125/min, we found a rightward shift of the pressure-volume relationship due to the increase in chamber volume. Despite nominally lower baseline filling pressures after 2 wk of pacing at 125/min, this did not reach significance (P = 0.24). However, diastolic compliance, expressed as ΔEDV/ΔEDP with the volume challenge, increased from 1.95 ± 0.72 ml/mmHg at baseline to 3.18 ± 0.57 ml/mmHg (P < 0.05) after 2 wk of pacing at 125/min.

**Tissue Fibrosis Levels**

Analysis of the myocardial connective tissue levels at the end of the study protocol revealed very low levels of interstitial connective tissue in all paced animals. Importantly, there was no difference between RAS animals and sham animals (Fig. 7). The relative area of total myocardial interstitial fibrosis was 0.69 ± 0.08% in sham and 0.52 ± 0.16% in RAS animals. Levels of myocardial connective tissue were significantly higher in referent control sections from five RAS animals that were not paced than in the paced RAS animals (1.45 ± 0.98%, P < 0.05).
results also suggest that rate-induced (re)modeling reduces myocardial fibrosis and improves diastolic compliance.

The rationale for heart rate as a treatment target originates from several physiological and clinical observations. Heart rate, through its high variability, plays a central role in cardiac workload. It is believed to play a key part in many physiological adaptations such as athlete’s heart and in pregnancy (8). Heart rate is an established treatment target in dilated cardiomyopathy, where a lower heart rate reduces LV chamber size (25). In contrast, extreme tachycardia is known to severely dilate the LV, as evident in tachycardia-induced cardiomyopathy. Accordingly, we speculated that modest heart rate elevations could be used to restore chamber volumes, reduce LV wall thickness, and trigger an interstitial remodeling process, which is very common and is frequently encountered in patients with concentric remodeling and HfPEF.

There was no significant difference in collagen I and III levels in homogenates from RAS and sham animals (Fig. 7).

**DISCUSSION**

In this study we explored whether moderately increased heart rates could be used to favorably modify the LV structure in pigs with concentric LV hypertrophy. This myocardial substrate is very common and is frequently encountered in patients with concentric remodeling and HfPEF.

The following novel observations were made: 1) a moderate elevation in heart rate can increase LV volumes and lower mass-to-volume ratios in concentrically hypertrophied LVs without reducing the EF, and 2) this change in LV structure is reversible, can be titrated, and is not associated with HF. Our

**Table 1. Cumulative data prior to pacing, after 2 wk of pacing, and after 2 wk without pacing**

<table>
<thead>
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<th></th>
<th>Collective Data</th>
<th>P Values</th>
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<tr>
<td></td>
<td>Baseline (n = 10)</td>
<td>125/min (n = 10)</td>
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<tr>
<td>Body wt/heart rate</td>
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<tr>
<td>Body wt, kg</td>
<td>42 ± 3</td>
<td>44 ± 2</td>
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<td>Heart rate, min⁻¹</td>
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<td>Atrial pacing, %</td>
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<td>LV mass, g</td>
<td>31.9 ± 2.8</td>
<td>35.2 ± 2.8</td>
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<td>ESV, ml</td>
<td>20.7 ± 3.8</td>
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<td>A2-D, cm²</td>
<td>9.7 ± 1.7</td>
<td>11.0 ± 2.4</td>
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<td>A2-S, cm²</td>
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<td>LV EDV, ml</td>
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<td>34.2 ± 11.1</td>
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<tr>
<td>LV ESV, ml</td>
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<td>EF, %</td>
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<td>Mass-to-volume ratio, g/ml</td>
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<td>E/Eʹ lat</td>
<td>7.6 ± 2.0</td>
<td>7.0 ± 2.9</td>
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<td>Tachypnea, %</td>
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<td>Crackles, %</td>
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<td>BNP, ng/ml</td>
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<td>NE, ng/ml</td>
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<tr>
<td>Galectin-3, ng/ml</td>
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<td>23 ± 6</td>
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Values are means ± SD. Baseline, prior to pacing; 125/min, after 2 wk of pacing; Recovery, after 2 wk without pacing; LV, left ventricle; RV, right ventricle; EDD, end-diastolic diameter; ESD, end-systolic diameter; A2-D, parasternal short-axis LV area at end diastole; A2-S, parasternal short-axis LV area at systole; LV EDV, end-diastolic volume; ESV, systolic volume; EF, ejection fraction; E/Eʹ lat, ratio of early ventricular filling peak velocity to lateral early diastolic mitral annular velocity; BNP, B-type natriuretic peptide; NE, norepinephrine. *Significant difference (P < 0.05).
which in aggregate could improve LV filling. If so, patients with concentric hypertrophy and impaired LV filling might potentially benefit from modest heart rate elevations, if the structural remodeling can be safely accomplished.

The unilateral RAS model is well characterized and recapitulates concentric LV hypertrophy and myocardial fibrosis (14, 15, 27). Although the lack of overt HF and only moderate levels of fibrosis are clear limitations of this model, they allowed us to establish the safety of moderately elevated heart rates in a common antecedent disease substrate of HFpEF. Because there is no established large-animal model that recapitulates HFpEF, this limitation cannot be overcome (10).

The potential to titrate the effects of heart rate was first apparent after the rate was lowered from 150/min to 125/min. The second experimental series confirmed that this reduction in LV chamber size was not merely a result of an ongoing regression of chamber volumes after pacing at 150/min but was specific to pacing at 125/min. We also determined that even a very pronounced chamber dilation induced by a heart rate of 170/min was not pathological, as determined by the lack of HF and biomarker elevations, and could be reversed with a rate of 125/min. Within 2 wk, chamber dimensions regressed and EF normalized. These rapid structural changes demonstrate that the swiftness of the physiological cardiac remodeling process is preserved in concentric hypertrophy, even when pushed to levels that exceed the physiological adaptations reported in human subjects exposed to extreme bed rest, spaceflight, or reconditioning (16, 20).

Although the results after pacing at >125/min suggest a gradual rate-dependent decrease in EF, no clinical signs or biomarker changes suggestive of HF were seen at any of the studied heart rates. This suggests that the observed structural changes are within the limits of physiological adaptations without causing decompensation. This is an important finding, as it is well established that pigs and other similar-sized animals develop overt HF with increased BNP and NE levels when exposed to pacing at >200/min (10).

Our analysis of the EDP-EDV relationship after pacing at 125/min demonstrates a rightward shift with a trend toward lower LV EDP with a substantial improvement in diastolic compliance. This may be primarily facilitated by wall thinning, but a pacing-induced reduction in myocardial connective tissue could contribute to this finding. Compared with nonpaced RAS animals, the level of myocardial connective tissue was low in all paced animals. The overall very low levels of connective tissue in all paced animals were very likely the result of a concurrent interstitial matrix remodeling process that reduces collagen levels to support the elongation and thinning of individual myocytes, as shown in animal models of tachycardia-induced cardiomyopathy (5, 22). Because the analyzed tissues originate from hearts that were allowed to return to their baseline geometry, it is conceivable that intermittent increases in heart rate may suffice to reduce tissue fibrosis without a permanent chamber enlargement.

It is important to acknowledge that it is not known if a reversal of concentric remodeling would be beneficial in patients. However, it is equally possible that a lack of structural remodeling in clinical HFpEF trials may also have contributed to their failure. It is also unclear if a moderate increase in heart rate would be feasible and safe in patients, as it could result in symptoms and tachycardia-induced cardiomyopathy. It is also possible that older age, advanced myocardial fibrosis, and other changes in the composition of the myocardium may reduce myocardial plasticity (28). High resting heart rates are generally viewed as an unfavorable prognostic sign (6). This may be the reason why only very few clinical studies have investigated

Fig. 7. A: PicroSirius-stained myocardial section from a RAS animal and a sham animal after pacing demonstrating similar amounts of interstitial connective tissue. B: slot-blot analysis of collagen I/III protein levels in myocardial tissue homogenates. Collagen levels were not different in RAS and sham animals. au, Arbitrary units.
interventions that allow increased heart rates. An important trial in this regard is the guideline-influencing Rate Efficacy in Permanant Atrial Fibrillation (RACE-2) study (26). This trial compared two heart rate control strategies in patients with atrial fibrillation and included patients with HFrEF. The strict rate control group had a target heart rate of <80/min at rest. The lenient rate control allowed heart rates of up to 110/min. This study established the noninferiority of higher heart rates with a numerical signal toward better outcomes. Similar to our own preliminary experience of nocturnal pacing at comparable rates, no untoward safety signals were observed. This may also provide some reassurance for older patient populations with a high prevalence of coronary artery disease.

Despite the discussed limitations of the experimental model and the exploratory approach, our proof-of-concept study suggests that a heart rate-mediated innate cardiac adaptation mechanism could be employed to safely induce potentially beneficial myocardial remodeling. This may present a therapeutic opportunity in symptomatic patients with concentric remodeling and increased LV mass-to-volume ratios, as commonly encountered in HFrEF. Furthermore, it appears possible that such an intervention could achieve favorable effects, even if applied intermittently and at lower rates.

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DISCLOSURES

Medtronic supported this research. The University of Vermont has received intellectual property protection for the use of pacemakers to prevent and treat heart failure with preserved ejection fraction and related conditions.

AUTHOR CONTRIBUTIONS

F.J.K., S.B., K.E.R., and M.M. performed the experiments; F.J.K. and M.M. analyzed the data; F.J.K., T.A., L.O.L., M.M.L., and M.M. interpreted the results of the experiments; F.J.K., S.B., K.E.R., L.O.L., M.M.L., and M.M. edited and revised the manuscript; F.K., S.B., K.E.R., T.A., M.M.L., and M.M. approved the final version of the manuscript; K.E.R., M.M.L., and M.M. drafted the manuscript; M.M.L. and M.M. developed the concept and designed the research; M.M. prepared the figures.

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